

DAFTAR PUSTAKA

- Agustina, E., Andiarna, F., & Hidayati, I. (2020). Antioxidant Activity of Black Garlic Extract with Variations of the Heating Time. *Al-Kauniyah: Jurnal Biologi*, *13*(1), 39–50.
- Ameer, K., Shahbaz, H. M., & Kwon, J. (2017). Green Extraction Methods for Polyphenols from Plant Matrices and Their Byproducts : A Review. *Comprehensive Reviews in Food Science and Food Safety*, *16*(2), 295–315. <https://doi.org/10.1111/1541-4337.12253>
- Artania, N. P. A. I., Harta, I. K. G. G. G., Pratama, G. W. A. P., Ayu, N. P. A. S., Sukmarani, I. G. A. P., & Arisanti, C. I. S. (2020). Optimasi Propilenglikol dalam Sediaan Sirup Obat Batuk Ekstrak Rimpang Jahe. *Journal of Chemistry*, *14*(2), 182–187.
- Bera, K., Nosalova, G., Sivova, V., & Ray, B. (2016). Structural Elements and Cough Suppressing Activity of Polysaccharides from *Zingiber officinale* Rhizome. *Phytotherapy Research*, *30*, 105–111.
- Candra, L. M. M., Andayani, Y., & Wirasisya, D. G. (2021). Pengaruh Metode Ekstraksi Terhadap Kandungan Fenolik Total dan Flavonoid Total Pada Ekstrak Etanol Buncis (*Phaseolus vulgaris* L.). *Jurnal Pijar Mipa*, *16*(3), 397–405. <https://doi.org/10.29303/jpm.v16i3.2308>
- Chaiklahan, R., Chirasuwan, N., Triratana, P., Loha, V., Tia, S., & Bunnag, B. (2013). Polysaccharide extraction from *Spirulina* sp. and its antioxidant capacity. *International Journal of Biological Macromolecules*, *58*, 73–78. <https://doi.org/10.1016/j.ijbiomac.2013.03.046>
- Chen, G., Yuan, B., Wang, H., Qi, G., & Cheng, S. (2019). Characterization and Antioxidant Activity of Polysaccharides Obtained from Ginger Pomace Using Two Different Extraction Processes. *International Journal of Biological Macromolecules*, *139*, 801–809. <https://doi.org/10.1016/j.ijbiomac.2019.08.048>
- Cho, Y. J., Getachew, A. T., Saravana, P. S., & Chun, B. S. (2019). Optimization and Characterization of Polysaccharides Extraction from Giant African snail (*Achatina fulica*) Using Pressurized Hot Water Extraction (PHWE). *Bioactive Carbohydrates and Dietary Fibre*, *18*, 100179. <https://doi.org/10.1016/j.bcdf.2019.100179>
- Deglas, W. (2019). Pengaruh Lama Perendaman dan Konsentrasi Etanol Terhadap Rendemen pada Pembuatan Minyak Esensial Kulit Buah Jeruk Pontianak. *TEKNOLOGI PANGAN: Media Informasi Dan Komunikasi Ilmiah Teknologi Pertanian*, *10*(2), 88–94. <https://doi.org/10.35891/tp.v10i2.1645>
- Fernández, J. P., Rodr, J. A., Tojo, E., & Andrade, J. M. (2003). Quantitation of k-, i- and λ-Carrageenans by Mid-Infrared Spectroscopy and PLS Regression. *Analytica Chimica Acta*, *480*, 23–37. [https://doi.org/10.1016/S0003-2670\(02\)01592-1](https://doi.org/10.1016/S0003-2670(02)01592-1)
- Helmalia, A. W., Putrid, & Dirpan, A. (2019). The Potential of Traditional Spices as a Source of Natural Antioxidants for Functional Food Raw Materials. *Canrea Journal: Food Technology, Nutritions, and Culinary Journal*, *2*(1), 26–31.
- Herawati, I. E., & Saptarini, N. M. (2019). Studi Fitokimia pada Jahe Merah (*Zingiber officinale* Roscoe Var. Sunti Val). *Majalah Farmasetika*, *4*, 22–27.
- Li, S., & Shah, N. P. (2014). Antioxidant and Antibacterial Activities of Sulphated Polysaccharides from *Pleurotus eryngii* and *Streptococcus thermophilus* ASCC 1275. *Food Chemistry*, *165*, 262–270. <https://doi.org/10.1016/j.foodchem.2014.05.110>

- Mahmudati, N., Wahyono, & Djunaedi, D. (2020). Antioxidant activity and total phenolic content of three varieties of Ginger (*Zingiber officinale*) in decoction and infusion extraction method Antioxidant activity and total phenolic content of three varieties of Ginger (*Zingiber officinale*) in decoct. *Journal of Physics*, 102(1), 1–6. <https://doi.org/10.1088/1742-6596/1567/2/022028>
- Maran, J. P., Mekala, V., & Manikandan, S. (2018). Modeling and Optimization of Ultrasound-Assisted Extraction of Polysaccharide from Cucurbita Moschata. *Carbohydrate Polymers*, 92(2), 2018–2026. <https://doi.org/10.1016/j.carbpol.2012.11.086>
- Mingda, H., Jun, G., Ziqi, L., Yu, S., & Shuyong, L. (2021). Antioxidant and immunomodulatory activities in vitro of a neutral polysaccharide from ginger (*Zingiber officinale*). *Starch-Stärke*, 7, 9–10.
- Munadi, R. (2020). Analisis Komponen Kimia dan Uji Aktivitas Antioksidan Ekstrak Rimpang Jahe Merah (*Zingiber officinale* Rosc. Var rubrum). *Cokroaminoto Journal of Chemical Science*, 2(1), 1–6.
- Nie, C., Zhu, P., Ma, S., Wang, M., & Hu, Y. (2018). Purification, Characterization and Immunomodulatory Activity of Polysaccharides from Stem Lettuce. *Carbohydrate Polymers*, 188, 236–242. <https://doi.org/10.1016/j.carbpol.2018.02.009>
- Nielsen, S. S. (2017). *Total Carbohydrate by Phenol-Sulfuric Acid Method*. 137–141. <https://doi.org/10.1007/978-3-319-44127-6>
- Nursakinah, D., & Verawati, B. (2021). Pembuatan Permen Jeli Ekstrak Jahe Merah dengan Substitusi Ekstrak Jambu Biji Merah sebagai Sumber Antioksidan bagi Penderita Diabetes Melitus. *Jurnal Pangan Dan Gizi*, 11(2), 125–133.
- Panjaitan, R. S., & Natalia, L. (2021). Ekstraksi Polisakarida Sulfat dari Sargassum polycystum dengan Metode Microwave Assisted Extraction dan Uji Toksisitasnya. *Jurnal Pascapanen Dan Bioteknologi Kelautan Dan Perikanan*, 16(1), 23–32. <https://doi.org/10.15578/jpbkp.v16i1.692>
- Rahmi, H. (2017). ktivitas Antioksidan dari Berbagai Sumber Buah-buahan di Indonesia. *Jurnal Agrotek Indonesia*, 2(1), 34–38.
- Samudra, A. G., K, F. S., & Chintama, M. (2018). Polisakarida dan Senyawa Polifenol Alga Coklat (*Sargassum* sp.) Pada Mencit yang Diinduksi Aloksan. *Jurnal Ilmiah Manuntung*, 4(1), 48–52.
- Semwal, R. B., Semwal, D. K., Combrinck, S., & Viljoen, A. M. (2015). Phytochemistry Gingerols and Shogaols: Important Nutraceutical Principles from Ginger. *Phytochemistry*, 117, 554–568. <https://doi.org/10.1016/j.phytochem.2015.07.012>
- Sibuea, P. (2021). Review : Kajian Manfaat Makanan Fungsional di Saat Pandemi. *Jurnal Riset Teknologi Pangan Dan Hasil Pertanian (RETIPA)*, 2(1), 83–92.
- Soebagio, S. B., Soares, J. S., Indraswati, N., & Kurniawan, Y. (2014). Ekstraksi Polisakarida Pada Biji Tamarind (*Tamarindus Indica* L). *Jurnal Ilmiah Widya Teknik*, 13(2), 23–32.
- Souza, B. W. S., Cerqueira, M. A., Bourbon, A. I., Pinheiro, A. C., Martins, J. T., Teixeira, J. A., Coimbra, M. A., & Vicente, A. A. (2012). Food Hydrocolloids Chemical Characterization and Antioxidant Activity of Sulfated Polysaccharide from the Red Seaweed *Gracilaria birdiae*. *Food Hydrocolloids*, 27(2), 287–292. <https://doi.org/10.1016/j.foodhyd.2011.10.005>

- Srikandi, S., Humaeroh, M., & Sutamihardja, R. (2020). Kandungan Gingerol dan Shogaol dari Ekstrak Jahe Merah (*Zingiber Officinale* Roscoe) dengan Metode Maserasi Bertingkat. *Al-Kimiya*, 7(2), 75–81. <https://doi.org/10.15575/ak.v7i2.6545>
- Tepsongkroh, B., Jangchud, K., & Trakoontivakorn, G. (2019). Antioxidant Properties and Selected Phenolic Acids of Five Different Tray - Dried and Freeze - Dried Mushrooms Using Methanol and Hot Water Extraction. *Journal of Food Measurement and Characterization*, 13(4), 3097–3105. <https://doi.org/10.1007/s11694-019-00232-2>
- Tsubaki, S., Oono, K., Hiraoka, M., Onda, A., & Mitani, T. (2016). Microwave-Assisted Hydrothermal Extraction of Sulfated Polysaccharides from *Ulva* spp . and *Monostroma latissimum*. *Food Chemistry*, 210, 311–316. <https://doi.org/10.1016/j.foodchem.2016.04.121>
- Wang, C., He, Y., Tang, X., & Li, N. (2020). Sulfation, Structural Analysis, and Anticoagulant Bioactivity of Ginger Polysaccharides. *Journal of Food Science*, 85(8), 2427–2434. <https://doi.org/10.1111/1750-3841.15338>
- Wang, Y., Wei, X., Wang, F., Xu, J., Tang, X., & Li, N. (2018). Structural Characterization and Antioxidant Activity of Polysaccharide from Ginger. *International Journal of Biological Macromolecules*, 111, 862–869. <https://doi.org/10.1016/j.ijbiomac.2018.01.087>
- Widyastuti, I., Luthfah, H. Z., Hartono, Y. I., & Islamadina, R. (2020). Antioxidant Activity of Temulawak (*Curcuma xanthorrhiza* Roxb.) and its Classification with Chemometrics. *Indonesian Journal of Chemometrics and Pharmaceutical Analysis*, 02(1), 29–42.
- Xu, Y., Cai, F., Yu, Z., Zhang, L., Li, X., Yang, Y., & Liu, G. (2016). Optimisation of Pressurised Water Extraction of Polysaccharides from Blackcurranta its Antioxidant Activity. *Food Chemistry*, 194, 650–658. <https://doi.org/10.1016/j.foodchem.2015.08.061>
- Xu, Z., Li, X., Feng, S., Liu, J., Zhou, L., Yuan, M., & Ding, C. (2016). International Journal of Biological Macromolecules Characteristics and bioactivities of different molecular weight polysaccharides from camellia seed cake. *International Journal of Biological Macromolecules*, 91, 1025–1032. <https://doi.org/10.1016/j.ijbiomac.2016.06.067>
- Yang, W., Pei, F., Shi, Y., Zhao, L., Fang, Y., & Hu, Q. (2012). Purification, Characterization and Anti-Proliferation Activity of Polysaccharides from *Flammulina Velutipes*. *Carbohydrate Polymers*, 88(2), 474–480. <https://doi.org/10.1016/j.carbpol.2011.12.018>
- Yangthong, M., Ruensirikul, J., & Kaneko, G. (2022). The Hot-Water Extract of *Sargassum* sp . as a Feed Ingredient for Spotted Scat (*Scatophagus argus* Linnaeus , 1766) Reared in Songkhla Lake : Effects on Growth , Feed Efficiency , Hematological Data and Body Composition. *Fishes*, 7(170), 1–10.
- Ye, H., Wang, K., Zhou, C., Liu, J., & Zeng, X. (2008). Purification, Antitumor and Antioxidant Activities in Vitro of Polysaccharides from the Brown Seaweed *Sargassum pallidum*. *Food Chemistry*, 111, 428–432. <https://doi.org/10.1016/j.foodchem.2008.04.012>
- Yun, L., Li, D., Yang, L., & Zhang, M. (2019). Hot Water Extraction and Artificial Simulated Gastrointestinal Digestion of Wheat Germ Polysaccharide. *International Journal of Biological Macromolecules*, 123, 174–181. <https://doi.org/10.1016/j.ijbiomac.2018.11.111>

Zhang, S., Kou, X., Zhao, H., Mak, K. K., Balijepalli, M. K., & Pichika, M. R. (2022). Zingiber officinale var. rubrum: Red Ginger's Medicinal Uses. *Molecules*, 27(3), 1–31. <https://doi.org/10.3390/molecules27030775>

LAMPIRAN

Lampiran 1 Hasil Uji Statistik

ANOVA

		Sum of Squares	df	Mean Square	F	Sig.
Total Gula	Between Groups	8.692	2	4.346	3.213	.180
	Within Groups	4.057	3	1.352		
	Total	12.749	5			
Sulfat	Between Groups	472650.648	2	236325.324	24.603	.014
	Within Groups	28816.062	3	9605.354		
	Total	501466.710	5			
Rendemen	Between Groups	.143	2	.072	1.195	.415
	Within Groups	.180	3	.060		
	Total	.324	5			
Antioksidan	Between Groups	164991.796	2	82495.898	10.544	.044
	Within Groups	23472.981	3	7824.327		
	Total	188464.777	5			
Total Fenolik	Between Groups	4172.707	2	2086.354	1.108E3	.000
	Within Groups	5.648	3	1.883		
	Total	4178.356	5			

Lampiran 2 Hasil Uji Lanjut *Duncan* Antioksidan

Antioksidan

Duncan

Perlakuan	N	Subset for alpha = 0.05	
		1	2
4 Jam	2	171.2300	
3 Jam	2		472.8950
2 Jam	2		557.6310
Sig.		1.000	.409

Means for groups in homogeneous subsets are displayed.

Lampiran 3 Hasil Uji Lanjut *Duncan* Total Fenolik

Total Fenolik

Duncan

Perlakuan	N	Subset for alpha = 0.05		
		1	2	3
2 Jam	2	82.1250		
4 Jam	2		1.2556E2	
3 Jam	2			1.4525E2
Sig.		1.000	1.000	1.000

Means for groups in homogeneous subsets are displayed.

Lampiran 4 Hasil Uji Lanjut Duncan Sulfat

Sulfat

Duncan			
Perlakuan	N	Subset for alpha = 0.05	
		1	2
3 Jam	2	3.3650	
4 Jam	2	5.8650	
2 Jam	2		600.0000
Sig.		.981	1.000

Means for groups in homogeneous subsets are displayed.

Lampiran 5 Perhitungan

1. Rendemen

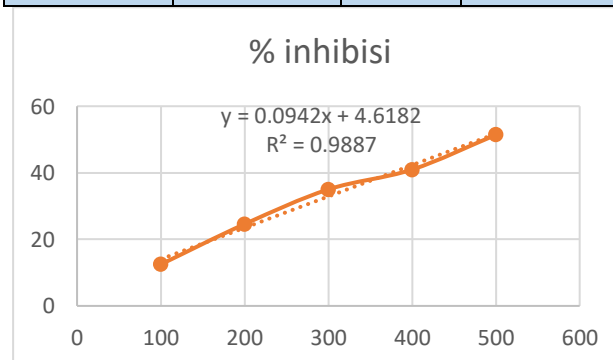
$$\% \text{ Yield} = \frac{\text{ekstrak yang dihasilkan (g)}}{\text{bahan baku yang digunakan (g)}} \times 100\%$$

- 2 jam ulangan 1 = $\frac{1.03}{100} \times 100\% = 1,03\%$
- 2 jam ulangan 2 = $\frac{1.03}{100} \times 100\% = 1,03\%$
- 3 jam ulangan 1 = $\frac{1.42}{100} \times 100\% = 1,42\%$
- 3 jam ulangan 2 = $\frac{0.83}{100} \times 100\% = 0,83\%$
- 4 jam ulangan 1 = $\frac{1.46}{100} \times 100\% = 1,46\%$
- 4 jam ulangan 2 = $\frac{1.34}{100} \times 100\% = 1,34\%$

2. Aktivitas Antioksidan

- 2 jam ulangan 1

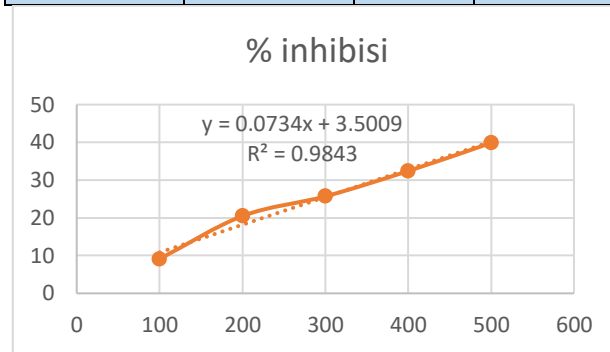
2.1			
Konsentrasi (ppm)	Absorbansi	Kontrol	% inhibisi
100	0.470	0.537	12.476723
200	0.405	0.537	24.581006
300	0.349	0.537	35.009311
400	0.317	0.537	40.968343
500	0.261	0.537	51.396648
IC50	481.760085		



- 2 jam ulangan 2

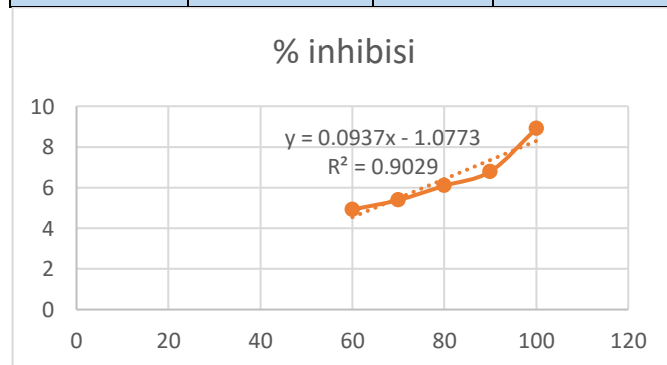
2.2

Konsentrasi (ppm)	Absorbansi	Kontrol	% inhibisi
100	0.488	0.537	9.1247672
200	0.427	0.537	20.484171
300	0.399	0.537	25.698324
400	0.363	0.537	32.402235
500	0.323	0.537	39.851024
IC50	633.502725		



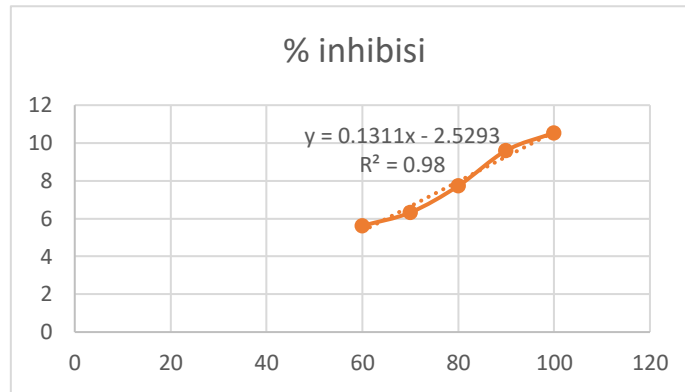
– 3 jam ulangan 1

3.1			
Konsentrasi (ppm)	Absorbansi	Kontrol	% inhibisi
60	0.406	0.427	4.918032787
70	0.404	0.427	5.386416862
80	0.401	0.427	6.088992974
90	0.398	0.427	6.791569087
100	0.389	0.427	8.899297424
IC50	545.1152615		



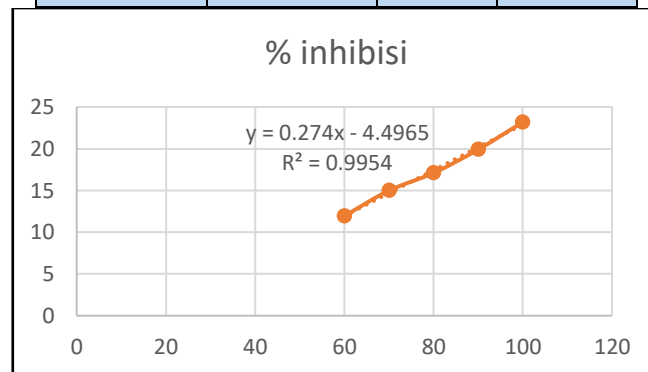
– 3 jam ulangan 2

3.2			
Konsentrasi (ppm)	Absorbansi	Kontrol	% inhibisi
60	0.403	0.427	5.620608899
70	0.400	0.427	6.323185012
80	0.394	0.427	7.728337237
90	0.386	0.427	9.601873536
100	0.382	0.427	10.53864169
IC50	400.6811594		



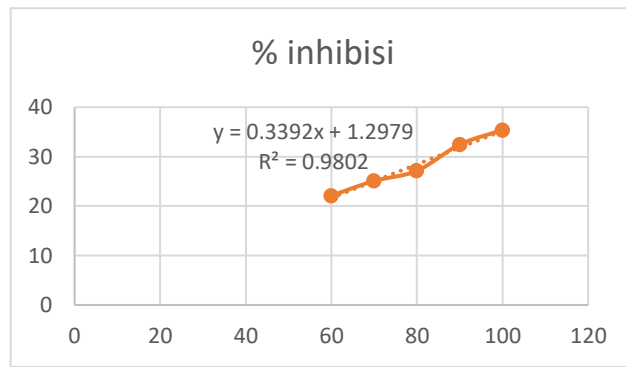
– 4 jam ulangan 1

4.1			
Konsentrasi (ppm)	Absorbansi	Kontrol	% inhibisi
60	0.376	0.427	11.94379
70	0.363	0.427	14.98829
80	0.354	0.427	17.09602
90	0.342	0.427	19.90632
100	0.328	0.427	23.18501
IC50	198.892336		



– 4 jam ulangan 2

4.2			
Konsentrasi (ppm)	Absorbansi	Kontrol	% inhibisi
60	0.264	0.339	22.12389
70	0.254	0.339	25.07375
80	0.247	0.339	27.13864
90	0.229	0.339	32.44838
100	0.219	0.339	35.39823
IC50	143.579304		



3. Total Fenolik

Perlakuan	Absorbansi	Berat hasil evaporasi (gram)	Berat sampel ditimbang (gr)/10 ml EtOH 70%	FP	Konsentrasi ($\mu\text{g/mL}$)	Total Fenol dalam 0,1 gr (μg)	Total Fenol dalam 1 gr (mg)	Total Fenol dalam berat hasil eva (mg)	Total Fenol mg GAE/gr	Rata-rata
2 jam	0.160	1.03	0.05	10 x	815.000	8150.000	81.500	83.945	81.500	82.125
	0.162	1.03	0.05		827.500	8275.000	82.750	85.233	82.750	
3 jam	0.262	1.42	0.05		1452.500	14525.000	145.250	149.608	145.250	145.250
	0.262	0.83	0.05		1452.500	14525.000	145.250	120.558	145.250	
4 jam	0.228	1.45	0.05		1240.000	12400.000	124.000	179.800	124.000	125.563
	0.233	1.34	0.05		1271.250	12712.500	127.125	170.348	127.125	

4. Total Gula

$$Y = 0,0063x + 0,0883$$

$$FP = 60$$

– 2 jam ulangan 1

$$0,237 = 0,0063x + 0,0883$$

$$x = \frac{0,1487}{0,0063} = 23 \times 60 = 1,380 \mu\text{g/ml}$$

– 2 jam ulangan 2

$$0,352 = 0,0063x + 0,0883$$

$$x = \frac{0,2637}{0,0063} = 41 \times 60 = 2,460 \mu\text{g/ml}$$

– 3 jam ulangan 1

$$0,491 = 0,0063x + 0,0883$$

$$x = \frac{0,4027}{0,0063} = 63 \times 60 = 3,780 \mu\text{g/ml}$$

– 3 jam ulangan 2

$$0,219 = 0,0063x + 0,0883$$

$$x = \frac{0,1307}{0,0063} = 20 \times 60 = 1,200 \mu\text{g/ml}$$

– 4 jam ulangan 1

$$0,615 = 0,0063x + 0,0883$$

$$x = \frac{0,5267}{0,0063} = 83 \times 60 = 4,980 \mu\text{g/ml}$$

– 4 jam ulangan 2

$$0,556 = 0,0063x + 0,0883$$

$$x = \frac{0,4677}{0,0063} = 74 \times 60 = 4,440 \mu\text{g/ml}$$

5. Sulfat

$$Y = 0,0012x + 0,3626$$

$$FP = 20$$

– 2 jam ulangan 1

$$0,333 = 0,0012x + 0,3626$$

$$x = \frac{0,0296}{0,0012} = 24 \times 20 = 480 \text{ mg/ml}$$

– 2 jam ulangan 2

$$0,406 = 0,0063x + 0,0883$$

$$x = \frac{0,0434}{0,0012} = 36 \times 20 = 720 \text{ mg/ml}$$

– 3 jam ulangan 1

$$0,530 = 0,0063x + 0,0883$$

$$x = \frac{0,1674}{0,0012} = 139,5 \times 20 = 2,790 \text{ mg/ml}$$

– 3 jam ulangan 2

$$0,599 = 0,0063x + 0,0883$$

$$x = \frac{0,2364}{0,0012} = 197 \times 20 = 3,940 \text{ mg/ml}$$

– 4 jam ulangan 1

$$0,548 = 0,0063x + 0,0883$$

$$x = \frac{0,1854}{0,0012} = 154,5 \times 20 = 3,090 \text{ mg/ml}$$

– 4 jam ulangan 2

$$0,882 = 0,0063x + 0,0883$$

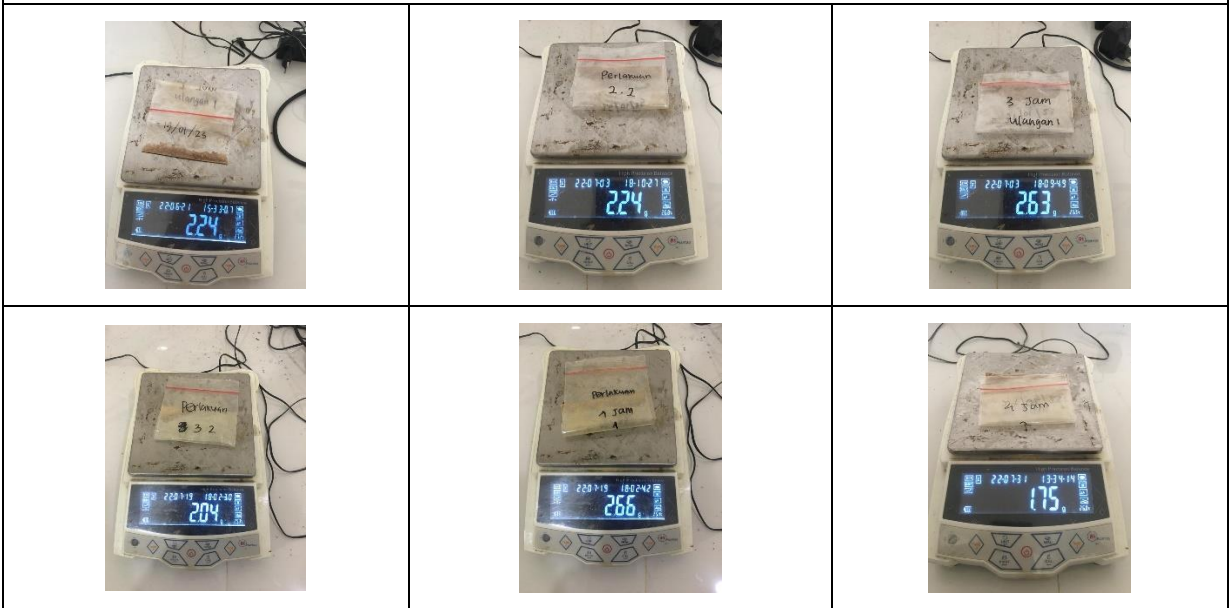
$$x = \frac{0,5194}{0,0012} = 432 \times 20 = 8,640 \text{ mg/ml}$$

Lampiran 6 Dokumentasi Penelitian

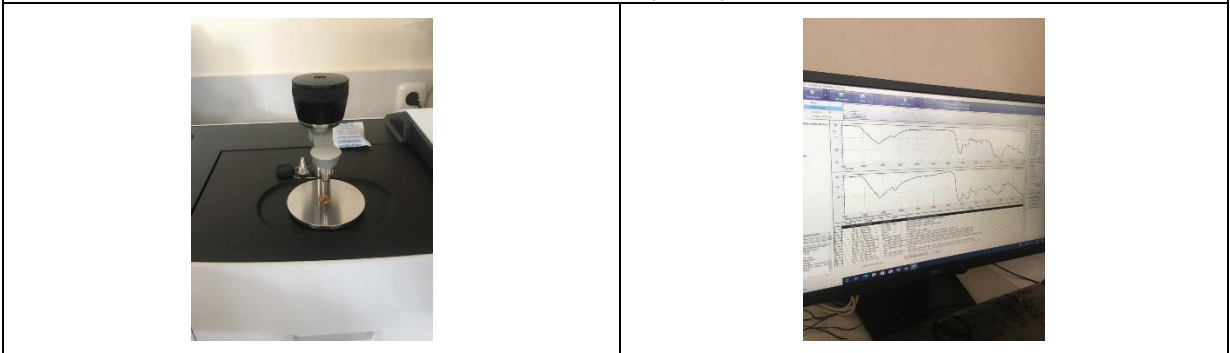




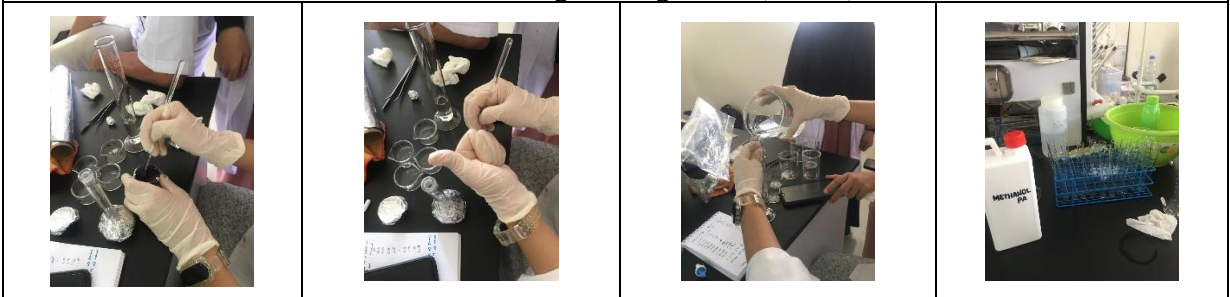
Ekstraksi Polisakarida Metode *Hot Water Extraction*

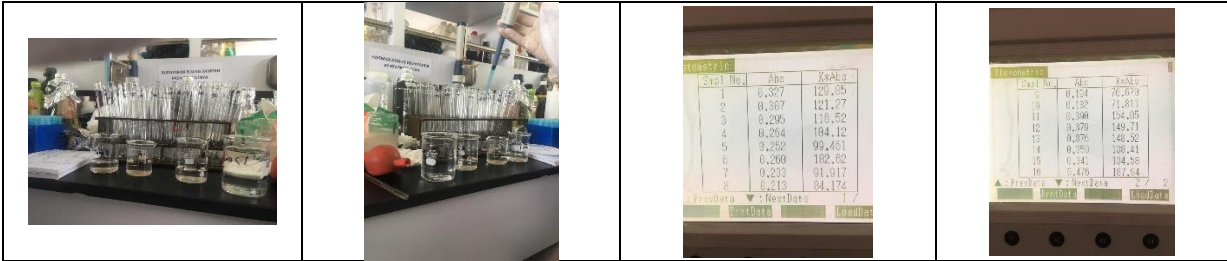


Rendemen (Yield)



Penentuan Gugus Fungsional (FT-IR)





Penentuan Aktivitas Antioksidan



Penentuan Total Gula



Penentuan Sulfat