

DAFTAR PUSTAKA

- Aldavood, S. J., Abbott, L. C., Evans, Z. R., Griffin, D. J., Lee, M. D., Quintero-Arevalo, N. M., & Villalobos, A. R. (2020). Effect of Cadmium and Nickel Exposure on Early Development in zebrafish (*Danio rerio*) embryos. *Water (Switzerland)*, 12(11), 1–16. <https://doi.org/10.3390/w12113005>
- Arrusyda, M., & Retnoaji, B. (2022). Pengaruh Koloid Nanoperak terhadap Perkembangan Embrio Ikan Wader Pari (*Rasbora lateristriata*) [The Effect of Nanosilver Colloids on the Embryo Development of the Wader Fish (*Rasbora lateristriata*)]. *Berita Biologi*, 21(2), 111–118.
- Barjhoux, I., Baudrimont, M., Morin, B., Landi, L., Gonzalez, P., & Cachot, J. (2012). Effects of Copper and Cadmium Spiked-Sediments on Embryonic Development of Japanese medaka (*Oryzias latipes*). *Ecotoxicology and Environmental Safety*, 79, 272–282. <https://doi.org/10.1016/j.ecoenv.2012.01.011>
- Barjhoux, I., Gonzalez, P., Baudrimont, M., & Cachot, J. (2016). Molecular and Phenotypic Responses of Japanese medaka (*Oryzias latipes*) Early Life Stages to Environmental Concentrations of Cadmium in Sediment. *Environmental Science and Pollution Research*, 23(18), 17969–17981. <https://doi.org/10.1007/s11356-016-6995-4>
- Berger, A. (2010). Molecular Analysis of The *Oryzias latipes* (Medaka) Transcriptome. *Freien Universität Berlin*, 1–166.
- Berniyanti, T. (2018). Biomarker Toksisitas Paparan Logam Tingkat Molekuler. Universitas Airlangga.
- Bertoni, Í., Sales, B. C. P., Viriato, C., Peixoto, P. V. L., & Pereira, L. C. (2024). Embryotoxicity Induced by Triclopyr in Zebrafish (*Danio rerio*) Early Life Stage. *Toxics*, 12(4). <https://doi.org/10.3390/toxics12040255>
- Budiardi, T., Cahyaningrum, W., & Effendi, I. (2005). Efisiensi Pemanfaatan Kuning Telur Embrio dan Larva Ikan Maanvis (*Pterophyllum scalare*) pada Suhu Inkubasi yang Berbeda. *Jurnal Akuakultur Indonesia*, 4(1), 57–61.
- Burggren, W. W., Dubansky, B., & Bautista, N. M. (2017). Cardiovascular Development in Embryonic and Larval Fishes. In *Fish Physiology* (1st ed., Vol. 36, Issue PartB). Elsevier Inc. <https://doi.org/10.1016/bs.fp.2017.09.002>
- Cahyanti, W., Mumpuni, F., & Yani, F. (2021). Perkembangan Embrio dan Performaa Awal Larva Ikan Tengadak (*Barbonymus schwanenfeldii*). *Jurnal Mina Sains*, 7(2). <https://doi.org/10.30997/jmss.v7i2.4687>
- Cho, J. G., Kim, K. T., Ryu, T. K., Lee, J. W., Kim, J. E., Kim, J., Lee, B. C., Jo, E. H., Yoon, J., Eom, I. C., Choi, K., & Kim, P. (2013). Stepwise Embryonic Toxicity of Silver Nanoparticles on *Oryzias latipes*. *BioMed Research International*, 2013. <https://doi.org/10.1155/2013/494671>
- Chou, Y., Zhu, P., Huang, X., Lin, J., Liu, J., & Gu, Y. (2018). Comparison Between Heart Rate Variability and Pulse Rate Variability for Bradycardia and Tachycardia Subjects. *2018 International Conference on Control, Automation and Information Sciences (ICCAIS)*, 1–6.

- Chow, E. S. H., & Cheng, S. H. (2003). Cadmium Affects Muscle Type Development and Axon Growth in zebrafish Embryonic Somitogenesis. *Toxicological Sciences*, 73(1), 149–159. <https://doi.org/10.1093/toxsci/kfg046>
- Cong, Y., Jin, F., Wang, J., & Mu, J. (2017). The Embryotoxicity of ZnO Nanoparticles to Marine medaka, *Oryzias melastigma*. *Aquatic Toxicology*, 185, 11–18. <https://doi.org/10.1016/j.aquatox.2017.01.006>
- Damayani, I. A., Kurniawan B, A., Yaqin, K., & Irmawati. (2022). Pengaruh Paparan Logam Timbal Terhadap Kematian Embrio Ikan Medaka Embrio (*Oryzias celebensis* Weber, 1894) di Indonesia. *SIGANUS: Journal of Fisheries and Marine Science*, 3(2), 215–221. <https://doi.org/10.31605/siganus.v3i2.1507>
- Dubińska-Magiera, M., Daczewska, M., Lewicka, A., Migocka-Patrzałek, M., Niedbalska-Tarnowska, J., & Jagl, K. (2016). Zebrafish: A Model for The Study of Toxicants Affecting Muscle Development and Function. *International Journal of Molecular Sciences*, 17(11). <https://doi.org/10.3390/ijms17111941>
- Emilia, I., Suheryanto, & Hanafiah, Z. (2013). Distribusi Logam Kadmium dalam Air dan Sedimen di Sungai Musi Kota Palembang. *Jurnal Penelitian Sains*, 16(2), 59–64.
- Fahmi, M. R., Prasetyo, A. B., & Vidiakusuma, R. (2015). Potensi Ikan Medaka (*Oryzias woworae*, *O. javanicus* dan *O. profundicola*) sebagai ikan hias dan ikan model. *Prosiding Seminar Nasional Ikan Ke 8*, 227–233.
- Gerber, R., Smit, N. J., van Vuren, J. H. J., Ikenaka, Y., & Wepener, V. (2018). Biomarkers in Tigerfish (*Hydrocynus vittatus*) as Indicators of Metal and Organic Pollution in Ecologically Sensitive Subtropical Rivers. *Ecotoxicology and Environmental Safety*, 157(March), 307–317.
- González-Doncel, M., Okihiro, M. S., Villalobos, S. A., Hinton, D. E., & Tarazona, J. V. (2005). A Quick Reference Guide to The Normal Development of *Oryzias latipes* (Teleostei, Adrianichthyidae). *Journal of Applied Ichthyology*, 21(1), 39–52. <https://doi.org/10.1111/j.1439-0426.2004.00615.x>
- Hastjarjo, T. D. (2019). Rancangan Eksperimen-Kuasi. *Buletin Psikologi*, 27(2), 187–203. <https://doi.org/10.22146/buletinpsikologi.38619>
- Hornung, M. W., Spitsbergen, J. M., & Peterson, R. E. (1999). 2,3,7,8-tetrachlorodibenzo-p-dioxin Alters Cardiovascular and Craniofacial Development and Function in Sac Fry of Rainbow Trout (*Oncorhynchus mykiss*). *Toxicological Sciences*, 47(1), 40–51. <https://doi.org/10.1093/toxsci/47.1.40>
- Indriana, E. F., Risandiansyah, R., & Aini, N. (2019). Efek Paparan Kronik Cadmium Chloride (CdCl₂) Dosis Rendah Terhadap Tubulus Hiperplasia Lamela Sekunder Insang dan Nekrosis Sel Tubulus Proksimal Ginjal Ikan Zebra Dewasa (*Danio rerio*). *Jurnal Kedokteran Komunitas*, 1–10.
- Iwamatsu, T. (2011). Developmental Stages in the Wild Medaka, *Oryzias latipes*. *Bulletin of Aichi Univ. of Education*, 60, 71–81.
- Jain-Schlaepfer, S., Fakan, E., Rummer, J. L., Simpson, S. D., & McCormick, M. I. (2018). Impact of Motorboats on Fish Embryos Depends on Engine Type. *Conservation Physiology*, 6(1), 1–9. <https://doi.org/10.1093/conphys/cov014>
- Johnson, A., Carew, E., & Sloman, K. A. (2007). The Effects of Copper on The

- Morphological and Functional Development of Zebrafish Embryos. *Aquatic Toxicology*, 84(4), 431–438. <https://doi.org/10.1016/j.aquatox.2007.07.003>
- Khodadoust, D., Ismail, A., Zulfikli, S., Z., & Tayefeh, F., H. (2013). Short Time Effect of Cadmium and Copper on Java Medaka (*Oryzias javanicus*) as Bioindicator for Ecotoxicological Studies. *Life Science Journal*, 10 (1)(May 2014).
- Korwin-Kossakowski, M. (2012). Fish Hatching Strategies: A review. *Reviews in Fish Biology and Fisheries*, 22(1), 225–240. <https://doi.org/10.1007/s11160-011-9233-7>
- Kumar, R., Mohanty, U. L., & Pillai, B. R. (2021). Effect of Hormonal Stimulation on Captive Broodstock Maturation, Induced Breeding and Spawning Performance of Striped Snakehead, *Channa striata* (Bloch, 1793). *Animal Reproduction Science*, 224, 106650. <https://doi.org/10.1016/j.anireprosci.2020.106650>
- Lalitha, Sripathi, S. K., & Jayanthi. (2017). Acute Toxicity Study of Dermabin. *Current Issues in Pharmacy and Medicine: Science and Practice*, 0(1), 24–26. <https://doi.org/10.14739/2409-2932.2017.1.93456>
- Lalombo, Y. I. S., Yaqin, K., & Omar, S. B. A. (2021). Nutrient Absorption Rate of *Oryzias celebensis* Embryo. *Akuatikisile: Jurnal Akuakultur, Pesisir Dan Pulau-Pulau Kecil*, 5(2), 67–71. <https://doi.org/10.29239/j.akuatikisile.5.2.67-71>
- Li, H., Gu, X., Chen, H., Mao, Z., Zeng, Q., Yang, H., & Kan, K. (2021). Comparative Toxicological Effects of Planktonic Microcystis and Benthic *Oscillatoria* on zebrafish Embryonic Development: Implications for cyanobacteria risk assessment. *Environmental Pollution*, 274(xxxx), 115852.
- Liu, K., Song, J., Chi, W., Liu, H., Ge, S., & Yu, D. (2021). Developmental Toxicity in Marine Medaka (*Oryzias melastigma*) Embryos and Larvae Exposed to Nickel. *Comparative Biochemistry and Physiology Part - C: Toxicology and Pharmacology*, 248(7), 109082. <https://doi.org/10.1016/j.cbpc.2021.109082>
- Luderman, L. N., Michaels, M. T., Levic, D. S., & Knapik, E. W. (2023). Zebrafish Erc1b Mediates Motor Innervation and Organization of Craniofacial Muscles in Control of Jaw Movement. *Developmental Dynamics*, 252(1), 104–123.
- Malinda, U. S., Widyaningrum, I., Aini, N., Mt, J., Malang, H., Timur, J., Malinda, U. S., Widyaningrum, I., & Aini, N. (2019). Efek Teratogenik Cadmium Chloride (CdCl₂) Terhadap Stunting dan Kelengkungan Vertebrae Larva Ikan *Danio rerio*. 134–141.
- Manantung, V. O., Sinjal, H. J., & Monijung, R. D. (2013). Evaluasi Kualitas, Kuantitas Telur Dan Larva Ikan Patin Siam (*Pangasianodon Hiphopthalmus*) Dengan Penambahan Ovaprim Dosis Berbeda. *E-Journal BUDIDAYA PERAIRAN*, 1(3), 14–23. <https://doi.org/10.35800/bdp.1.3.2013.2718>
- Mandagi, I. F., Mokodongan, D. F., Tanaka, R., & Yamahira, K. (2018). A New Riverine Ricefish of the Genus *Oryzias* (Beloniformes, Adrianichthyidae) from Malili, Central Sulawesi, Indonesia. *Copeia*, 106(2), 297–304. <https://doi.org/10.1643/CI-17-704>
- Michibata, H., Nojima, Y., & Kojima, M. K. (1987). Stage Sensitivity of Eggs of the Teleost *Oryzias latipes* to Cadmium Exposure. *Environmental Research*, 42(2), 321–327. [https://doi.org/10.1016/S0013-9351\(87\)80197-4](https://doi.org/10.1016/S0013-9351(87)80197-4)
- Mulyani, Y. W. T., Solihin, D. D., & Affandi, R. (2015). Yolk Absorption Efficiency and Morphogenesis of The Silver Arawana *Osteoglossum bicirrhosum* (Cuvier, 1829)

- prelarvae at various interactions of temperature and salinity. *Jurnal Iktiologi Indonesia*, 15(3), 179–191.
- Nisa, C., Saputra, P., & Setiawati, E. (2021). Pengembangan dan Validasi Metode Uji Cadmium (Cd) Pada Air Permukaan Secara Spektrometri Serapan Atom Nyala. *Pertemuan Dan Presentasi Ilmiah Standardisasi*, 2020(Cd), 249–258. <https://doi.org/10.31153/ppis.2020.84>
- Padilla, S., Cowden, J., Hinton, D. E., Yuen, B., Law, S., Kullman, S. W., Johnson, R., Hardman, R. C., Flynn, K., & Au, D. W. T. (2009). Use of Medaka in Toxicity Testing. In *Current Protocols in Toxicology* (Issue SUPPL. 39).
- Parenti, L. R. (2008). A Phylogenetic Analysis and Taxonomic Revision of Ricefishes, *Oryzias* and Relatives (Beloniformes, Adrianichthyidae)[CiNii Research. *Zoological Journal of the Linnean Society*, 154, 494–610.
- Piatkowska, A. M., Evans, S. E., & Stern, C. D. (2021). Cellular Aspects of Somite Formation in Vertebrates. *Cells and Development*, 168(August), 203732. <https://doi.org/10.1016/j.cdev.2021.203732>
- Pujihastuti, Y., Nirmala, K., & Effendi, I. (2009). The Effect of Cirata Reservoir Sediment on Early Developmental Stage of Common Carp (*Cyprinus carpio*) Embryo. *Jurnal Akuakultur Indonesia*, 8(2), 185. <https://doi.org/10.19027/jai.8.185-192>
- Purbonegoro, T. (2017). Faktor-Faktor Yang Mempengaruhi Toksisitas Bahan Pencemar Terhadap Organisme Perairan. *Oseana*, 42(2), 12–22.
- Rahadian, A., & Riani, E. (2019). Pencemaran Cd Pada Ekosistem Perairan Tawar dan Mekanisme Gangguannya Pada Hewan Air: Sebuah Tinjauan (*Cd Pollution in Fresh Water Ecosystem and Disturbance Mechanism to Aquatic An May 2018*.
- Rand, G. M. (1995). *Fundamentals of Aquatic Toxicology: Effects, Environmental Fate and Risk Assessment*. CRC press.
- Rumahlatu, D. (2011). Biomonitoring: Sebagai alat asesmen kualitas perairan akibat logam berat kadmium pada invertebrata perairan. *Sainstis*.
- Sahabuddin, H., Harisuseno, D., & Yuliani, E. (2014). Analisa Status Mutu Air dan Daya Tampung Beban Pencemaran Wungai wangu Kota Kendari. *Jurnal Teknik Pengairan*, 5(1), 19–28.
- Schweizer, M., von der Ohe, P. C., Gräff, T., Kühnen, U., Hebel, J., Heid, C., Kundy, L., Kuttler, J., Moroff, F.-M., & Schlösinger, A.-F. (2022). Heart Rate As an Early Warning Parameter and Proxy for Subsequent Mortality in *Danio rerio* embryos exposed to ionisable substances. *Science of The Total Environment*, 818, 151744.
- Wang, R. F., Zhu, L. M., Zhang, J., An, X. P., Yang, Y. P., Song, M., & Zhang, L. (2020). Developmental Toxicity of Copper in Marine Medaka (*Oryzias melastigma*) Embryos and Larvae. *Chemosphere*, 247, 125923.
- Wijayanti, T., & Lestari, D. E. G. (2017). Bioremediasi Limbah Tercemar Kadmium (Cd) Pada Perairan di Kabupaten Pasuruan Menggunakan Bakteri Indigen Secara Ex-Situ. *Jurnal Pena Sains*, 4(2), 114–123.
- Witeska, M., Sarnowski, P., Lugowska, K., & Kowal, E. (2014). The effects of cadmium and copper on embryonic and larval development of ide *Leuciscus idus* L . 151–

163. <https://doi.org/10.1007/s10695-013-9832-4>
- Yang, L., Ho, N. Y., Alshut, R., Legradi, J., Weiss, C., Reischl, M., Mikut, R., Liebel, U., Müller, F., & Strähle, U. (2009). Zebrafish Embryos As Models for Embryotoxic and Teratological Effects of Chemicals. *Reproductive Toxicology*, 28(2), 245–253. <https://doi.org/10.1016/j.reprotox.2009.04.013>
- Yaqin, K. (2019). Petunjuk Praktis Aplikasi Biomaker Sederhana (Issue October).
- Yaqin, K. (2021). Mengenal dengan cepat embriogenesis ikan binisi, *Oryzias celebensis* untuk studi ekotoksikologi. Deepublish.
- Yaqin, K., Rahim, S. W., Parawansa, B. S., Kudsiah, H., Tambaru, R., Sari, D. K., Riani, E., Hardiana, A. D., Meimulya, & Lalombo, Y. I. (2024). Toxicity Detection of Pollutants in the Tallo River Using Simple Biomarkers of *Oryzias celebensis* Embryo. *International Journal on Advanced Science, Engineering and Information Technology*, 14(2), 665–674. <https://doi.org/10.18517/ijaseit.14.2.19625>
- Yasumasu, S., Iuchi, I., & Yamagami, K. (1989). Isolation and Some Properties of Low Choriolytic Enzyme (LCE), a Component of the Hatching Enzyme of the Teleost , *Oryzias latipes* Shigeki Life Science Received for publication , July One of the two component proteases of the hatching enzyme of the fis. *J. Biochem*, 105, 212–218.
- Yu, R. M. K., Lin, C. C., Chan, P. K., Chow, E. S. H., Murphy, M. B., Chan, B. P., Müller, F., Strähle, U., & Cheng, S. H. (2006). Four-dimensional Imaging and Quantification of Gene Expression in Early Developing Zebrafish (*Danio rerio*) Embryos. *Toxicological Sciences*, 90(2), 529–538. <https://doi.org/10.1093/toxsci/kfj115>
- Yudasmara, G. A. (2014). Biologi Perikanan. *Plantaxia*, Yogyakarta.
- Zhang, H., Cao, H., Meng, Y., Jin, G., & Zhu, M. (2012). Ecotoxicology and Environmental Safety The toxicity of cadmium (Cd 2 þ) towards embryos and pro-larva of soldatov ' s catfish (*Silurus soldatovi*). *Ecotoxicology and Environmental Safety*, 80, 258–265. <https://doi.org/10.1016/j.ecoenv.2012.03.013>
- Zhang, Han, L., Wang, J., Shu, M., Liu, K., Zhang, Y., Hsiao, C. Der, Tian, Q., & He, Q. (2021). Clozapine Induced Developmental and Cardiac Toxicity on Zebrafish Embryos by Elevating Oxidative Stress. *Cardiovascular Toxicology*, 21(5), 399–409. <https://doi.org/10.1007/s12012-021-09632-7>

LAMPIRAN

Lampiran 1. Hasil uji statistik *Kruskal-Wallis test* jumlah somit embrio *Oryzias celebensis*

Table Analyzed	JUMLAH SOMIT
Kruskal-Wallis test	
P value	0.9260
Exact or approximate P value?	Exact
P value summary	ns
Do the medians vary signif. ($P < 0.05$)?	No
Number of groups	4
Kruskal-Wallis statistic	0.5681
Data summary	
Number of treatments (columns)	4
Number of values (total)	12

Number of families	1
Number of comparisons per family	3
Alpha	0.05

Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	1.667	No	ns	>0.9999	B	0,05
Kontrol vs. 0,1	1.667	No	ns	>0.9999	C	0,1
Kontrol vs. 0,15	2.000	No	ns	>0.9999	D	0,15
Test details	Mean rank 1	Mean rank 2	Mean rank diff.	n1	n2	
Kontrol vs. 0,05	7.833	6.167	1.667	3	3	
Kontrol vs. 0,1	7.833	6.167	1.667	3	3	
Kontrol vs. 0,15	7.833	5.833	2.000	3	3	

Lampiran 2. Hasil uji statistik *Kruskal-Wallis test* detak jantung embrio *Oryzias celebensis***Stadia 24**

Kruskal-Wallis test	
P value	0.0052
Exact or approximate P value?	Approximate
P value summary	**
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4
Kruskal-Wallis statistic	12.74

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	9.050	No	ns	0.2462	B	0,05
Kontrol vs. 0,1	13.05	Yes	*	0.0365	C	0,1
Kontrol vs. 0,15	17.90	Yes	**	0.0018	D	0,15

Stadia 25

Kruskal-Wallis test	
P value	0.0004
Exact or approximate P value?	Approximate
P value summary	***
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4
Kruskal-Wallis statistic	18.11

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	9.750	No	ns	0.1782	B	0,05
Kontrol vs. 0,1	18.70	Yes	***	0.0009	C	0,1
Kontrol vs. 0,15	18.95	Yes	***	0.0007	D	0,15

Stadia 26

Kruskal-Wallis test	
P value	<0.0001
Exact or approximate P value?	Approximate
P value summary	****
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4
Kruskal-Wallis statistic	22.26

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	18.25	Yes	**	0.0013	B	0,05
Kontrol vs. 0,1	13.20	Yes	*	0.0322	C	0,1
Kontrol vs. 0,15	23.15	Yes	****	<0.0001	D	0,15

Stadia 27

Kruskal-Wallis test	
P value	<0.0001
Exact or approximate P value?	Approximate
P value summary	****
Do the medians vary signif. (P < 0.05)?	Yes
Number of groups	4
Kruskal-Wallis statistic	25.13

Number of families	1
Number of comparisons per family	3
Alpha	0.05

Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	24.25	Yes	****	<0.0001	B	0,05
Kontrol vs. 0,1	15.05	Yes	*	0.0112	C	0,1
Kontrol vs. 0,15	20.30	Yes	***	0.0003	D	0,15

Stadia 28

Kruskal-Wallis test	
P value	0.0092
Exact or approximate P value?	Approximate
P value summary	**
Do the medians vary signif. (P < 0.05)?	Yes
Number of groups	4
Kruskal-Wallis statistic	11.52

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	12.50	Yes	*	0.0147	B	0,05
Kontrol vs. 0,1	7.450	No	ns	0.2809	C	0,1
Kontrol vs. 0,15	13.45	Yes	**	0.0074	D	0,15

Stadia 29

Kruskal-Wallis test	
P value	0.0001
Exact or approximate P value?	Approximate
P value summary	***
Do the medians vary signif. (P < 0.05)?	Yes
Number of groups	4
Kruskal-Wallis statistic	20.42

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	22.75	Yes	****	<0.0001	B	0,05
Kontrol vs. 0,1	9.950	No	ns	0.1480	C	0,1
Kontrol vs. 0,15	12.30	Yes	*	0.0453	D	0,15
Kontrol vs. 0,15	31.75	19.45	12.30	10	10	

Stadia 30

Kruskal-Wallis test	
P value	0.0476
Exact or approximate P value?	Approximate
P value summary	*
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4
Kruskal-Wallis statistic	7.924

Number of families	1
Number of comparisons per family	3
Alpha	0.05

Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	-0.5000	No	ns	>0.9999	B	0,05
Kontrol vs. 0,1	-11.55	No	ns	0.0518	C	0,1
Kontrol vs. 0,15	-0.7500	No	ns	>0.9999	D	0,15

Stadia 31

Kruskal-Wallis test	
P value	0.0004
Exact or approximate P value?	Approximate
P value summary	***
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4
Kruskal-Wallis statistic	18.20

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	-11.10	No	ns	0.0735	B	0,05
Kontrol vs. 0,1	-6.000	No	ns	0.6723	C	0,1
Kontrol vs. 0,15	-20.30	Yes	***	0.0001	D	0,15

Stadia 32

Kruskal-Wallis test	
P value	<0.0001
Exact or approximate P value?	Approximate
P value summary	****
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4
Kruskal-Wallis statistic	22.64

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	23.30	Yes	****	<0.0001	B	0,05
Kontrol vs. 0,1	16.10	Yes	**	0.0044	C	0,1
Kontrol vs. 0,15	15.80	Yes	**	0.0054	D	0,15

Stadia 33

Kruskal-Wallis test	
P value	<0.0001
Exact or approximate P value?	Approximate
P value summary	****
Do the medians vary signif. (P < 0.05)?	Yes
Number of groups	4
Kruskal-Wallis statistic	27.01

Number of families	1
Number of comparisons per family	3
Alpha	0.05

Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	23.40	Yes	****	<0.0001	B	0,05
Kontrol vs. 0,1	6.250	No	ns	0.6293	C	0,1
Kontrol vs. 0,15	17.35	Yes	**	0.0015	D	0,15

Stadia 34

Kruskal-Wallis test	
P value	<0.0001
Exact or approximate P value?	Approximate
P value summary	****
Do the medians vary signif. (P < 0.05)?	Yes
Number of groups	4
Kruskal-Wallis statistic	22.71

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	-12.75	Yes	*	0.0323	B	0,05
Kontrol vs. 0,1	0.7500	No	ns	>0.9999	C	0,1
Kontrol vs. 0,15	-19.00	Yes	***	0.0004	D	0,15

Stadia 35

Kruskal-Wallis test	
P value	>0.9999
Exact or approximate P value?	Approximate
P value summary	ns
Do the medians vary signif. (P < 0.05)?	No
Number of groups	4
Kruskal-Wallis statistic	0.000

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	0.000	No	ns	>0.9999	B	0,05
Kontrol vs. 0,1	0.000	No	ns	>0.9999	C	0,1
Kontrol vs. 0,15	0.000	No	ns	>0.9999	D	0,15

Stadia 36

Kruskal-Wallis test	
P value	<0.0001
Exact or approximate P value?	Approximate
P value summary	****
Do the medians vary signif. (P < 0.05)?	Yes
Number of groups	4
Kruskal-Wallis statistic	33.05

Number of families	1
Number of comparisons per family	3
Alpha	0.05

Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	18.30	Yes	***	0.0008	B	0,05
Kontrol vs. 0,1	4.000	No	ns	>0.9999	C	0,1
Kontrol vs. 0,15	24.90	Yes	****	<0.0001	D	0,15

Stadia 37

Kruskal-Wallis test	
P value	<0.0001
Exact or approximate P value?	Approximate
P value summary	****
Do the medians vary signif. (P < 0.05)?	Yes
Number of groups	4
Kruskal-Wallis statistic	27.46

Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	-16.30	Yes	**	0.0020	B	0,05
Kontrol vs. 0,1	-16.20	Yes	**	0.0022	C	0,1
Kontrol vs. 0,15	-24.50	Yes	****	<0.0001	D	0,15

Lampiran 3. Hasil uji statistik *Kruskal-Wallis test* gerakan rahang embrio *Oryzias celebensis*

Stadia 36

Kruskal-Wallis test	
P value	0.0616
Exact or approximate P value?	Approximate
P value summary	ns
Do the medians vary signif. ($P < 0.05$)?	No
Number of groups	4
Kruskal-Wallis statistic	7.346

Number of families	1
Number of comparisons per family	3
Alpha	0.05

Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	4.450	No	ns	>0.9999	B	0,05
Kontrol vs. 0,1	6.400	No	ns	0.6273	C	0,1
Kontrol vs. 0,15	13.94	Yes	*	0.0232	D	0,15

Stadia 37

Kruskal-Wallis test	
P value	0.0240
Exact or approximate P value?	Approximate
P value summary	*
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4

Kruskal-Wallis statistic	9.434
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Number of families	1
Number of comparisons per family	3
Alpha	0.05

Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	-1.200	No	ns	>0.9999	B	0,05
Kontrol vs. 0,1	2.100	No	ns	>0.9999	C	0,1
Kontrol vs. 0,15	-12.70	Yes	*	0.0459	D	0,15

Lampiran 4. Hasil uji statistik *Kruskal-Wallis test* laju penyerapan kuning telur embrio *Oryzias celebensis*

Kruskal-Wallis test	
P value	0.0413
Exact or approximate P value?	Approximate
P value summary	*
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4
Kruskal-Wallis statistic	8.238

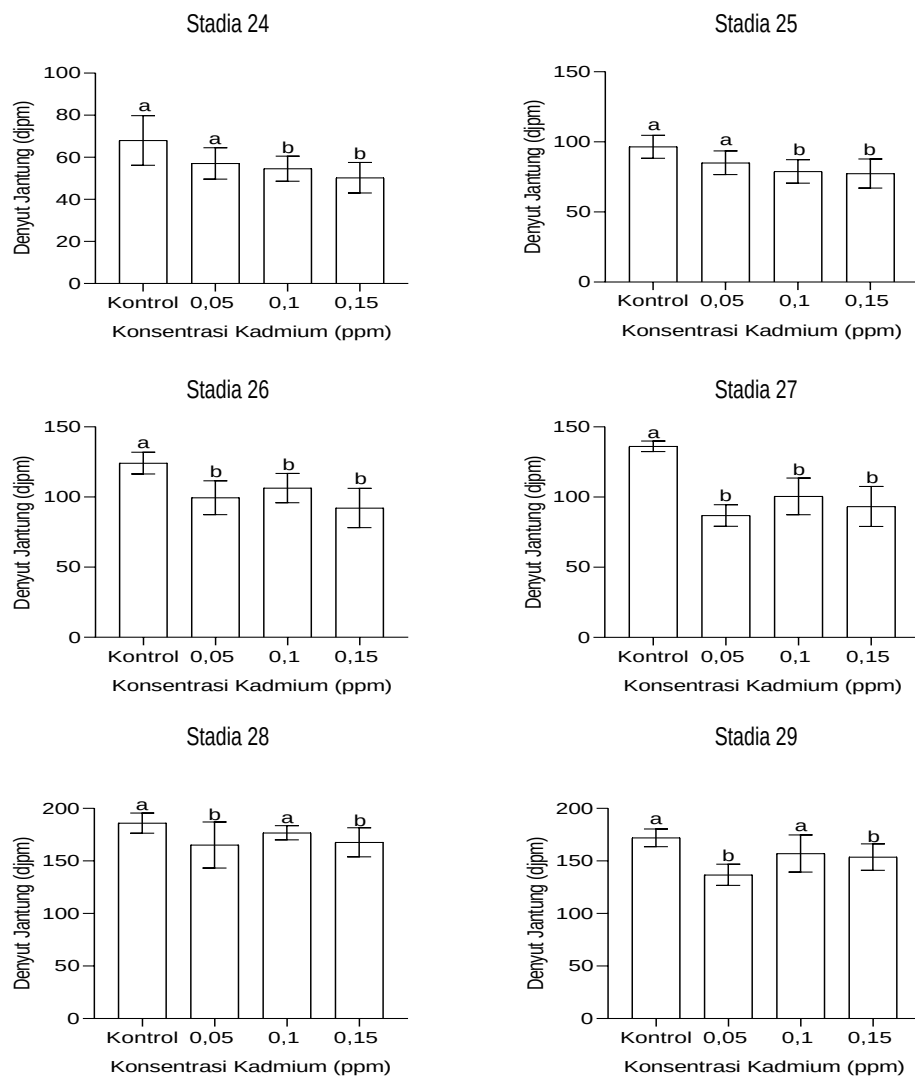
Number of families	1
Number of comparisons per family	3
Alpha	0.05

Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	-3.950	No	ns	>0.9999	B	0,05
Kontrol vs. 0,1	-6.500	No	ns	0.6255	C	0,1
Kontrol vs. 0,15	-14.35	Yes	*	0.0165	D	0,15

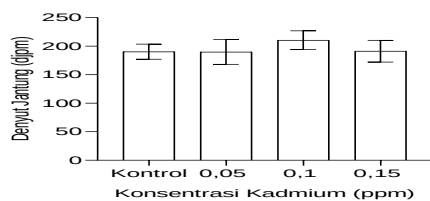
Lampiran 5. Hasil uji statistik *Kruskal-Wallis test* waktu penetasan embrio *Oryzias celebensis*

Kruskal-Wallis test	
P value	<0.0001
Exact or approximate P value?	Approximate
P value summary	****
Do the medians vary signif. ($P < 0.05$)?	Yes
Number of groups	4
Kruskal-Wallis statistic	33.80

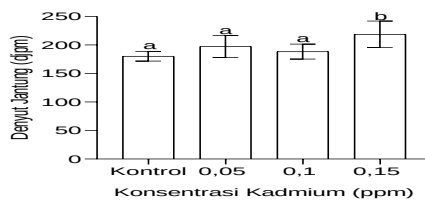
Number of families	1					
Number of comparisons per family	3					
Alpha	0.05					
Dunn's multiple comparisons test	Mean rank diff.	Significant?	Summary	Adjusted P Value	A-?	
Kontrol vs. 0,05	-3.600	No	ns	>0.9999	B	0,05
Kontrol vs. 0,1	-17.10	Yes	**	0.0017	C	0,1
Kontrol vs. 0,15	-25.30	Yes	****	<0.0001	D	0,15

Lampiran 6. Grafik detak jantung embrio *Oryzias celebensis* pada setiap stadia

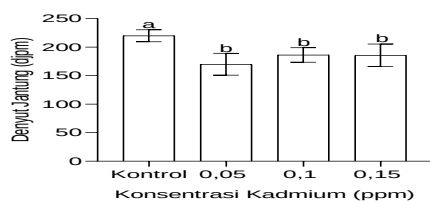
Stadia 30



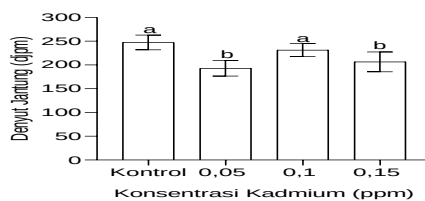
Stadia 31



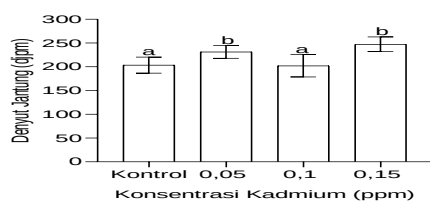
Stadia 32



Stadia 33



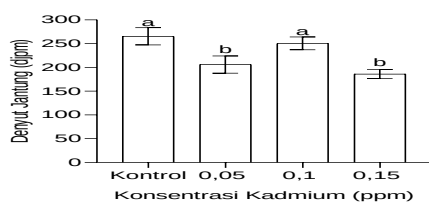
Stadia 34



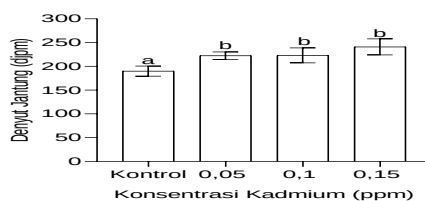
Stadia 35



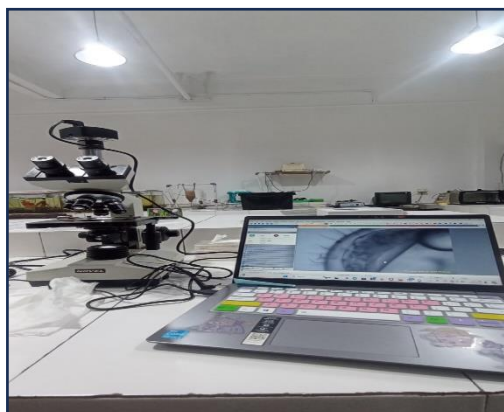
Stadia 36



Stadia 37



Lampiran 7. Dokumentasi Penelitian



CURRICULUM VITAE

A. Data Pribadi

1. Nama : Dwi Aryani
2. Tempat, Tanggal Lahir : Barata, 1 Februari 2002
3. Alamat : Barata, Kec. Marioriwawo, Kab. Soppeng
4. Kewarganegaraan : Indonesia

B. Riwayat Pendidikan

1. Tamat SD tahun 2014 di SD Negeri 143 Limpotenga
2. Tamat SMP tahun 2017 di SMP Negeri 3 Marioriwawo
3. Tamat SMA tahun 2020 di SMA Negeri 5 Soppeng