

## DAFTAR PUSTAKA

- Abdul, NS, Shenoy, M, Reddy, NR, Sangappa, SB, & ... (2024). Gene sequencing applications to combat oral-cavity related disorders: a systematic review with meta-analysis. *BMC Oral Health*, Springer, <https://doi.org/10.1186/s12903-023-03541-7>
- Abellan-Schneyder, I, Matchado, MS, Reitmeier, S, & ... (2021). Primer, pipelines, parameters: issues in 16S rRNA gene sequencing. *MspHERE*, Am Soc Microbiol, <https://doi.org/10.1128/msphere.01202-20>
- Afraz, M, Muhammad, F, Nisar, J, Shah, A, Munir, S, & ... (2023). Production of value added products from biomass waste by pyrolysis: An updated review. *Waste Management* ..., Elsevier, <https://www.sciencedirect.com/science/article/pii/S2949750723000263>
- Agarwal, PK (2018). A biophysical perspective on enzyme catalysis. *Biochemistry*, ACS Publications, <https://doi.org/10.1021/acs.biochem.8b01004>
- Aggarwal, S, & Ikram, S (2023). A comprehensive review on bio-mimicked multimolecular frameworks and supramolecules as scaffolds for enzyme immobilization. *Biotechnology and Bioengineering*, Wiley Online Library, <https://doi.org/10.1002/bit.28282>
- Appels, R, & Honeycutt, RL (2019). rDNA: evolution over a billion years. *DNA systematics*, taylorfrancis.com, <https://doi.org/10.1201/9780429286285-5>
- Aransiola, SA, Afolabi, F, Joseph, F, & ... (2022). Soil Enzymes: Distribution, Interactions, and Influencing Factors. *Agricultural* ..., taylorfrancis.com, <https://doi.org/10.1201/9781003313106-15>
- Arfah, R. A., Patong, A. R., Ahmad, A., dan Djide, M. N., 2014. Isolasi dan Identifikasi Bakteri Termofil Penghasil Amilase dari Sumber Air Panas Lejia Sulawesi Selatan. *Al-Kimia*, 2(2), 36-46.
- Asnawi, I., Natsir, H., dan Hariani, N., 2014. Eksplorasi Mikroba Penghasil Enzim Lipolitik Pada Sumber Air Panas Lemo Susu, Pinrang, Sulawesi Selatan. *Jurnal Sains Dasar*, 2(1): 1-6.
- Baehaki, A, Nopianti, R, Nofriansyah, H, & ... (2024). Isolation and production of proteases from bacteria isolate from Bekasam and Rusip (Traditional Food from South Sumatera, Indonesia). *World Journal of* ..., wjarr.com, <<https://wjarr.com/content/isolation-and-production-proteases-bacteria-isolate-bekasam-and-rusip-traditional-food-south>>
- Baksi, S, Sarkar, U, Villa, R, Basu, D, & ... (2023). Conversion of biomass to biofuels through sugar platform: A review of enzymatic hydrolysis highlighting the trade-off between product and substrate inhibitions. ... *Energy Technologies and* ..., Elsevier, <https://www.sciencedirect.com/science/article/pii/S2213138822010116>
- Bilal, M, Zhao, Y, Noreen, S, Shah, SZH, & ... (2019). Modifying bio-catalytic properties of enzymes for efficient biocatalysis: A review from immobilization strategies viewpoint. *Biocatalysis and* ..., Taylor & Francis, <https://doi.org/10.1080/10242422.2018.1564744>

- Bisswanger, H (2014). Enzyme assays. Perspectives in Science, Elsevier, <https://www.sciencedirect.com/science/article/pii/S2213020914000068>
- Buller, R, Lutz, S, Kazlauskas, RJ, Snajdrova, R, & ... (2023). From nature to industry: Harnessing enzymes for biocatalysis. Science, science.org, <https://doi.org/10.1126/science.adh8615>
- Burek, BO, Dawood, AWH, Hollmann, F, Liese, A, & ... (2022). Process intensification as game changer in enzyme catalysis. Frontiers in ..., frontiersin.org, <https://doi.org/10.3389/fctls.2022.858706>
- Cao, H, Wang, X, Liu, J, Sun, Z, Yu, Z, & ... (2023). Mechanistic insights into the changes of enzyme activity in food processing under microwave irradiation. ... Reviews in Food ..., Wiley Online Library, <https://doi.org/10.1111/1541-4337.13154>
- Cavanaugh, SE, & Bathrick, AS (2018). Direct PCR amplification of forensic touch and other challenging DNA samples: a review. *Forensic science international: Genetics*, Elsevier, <https://www.sciencedirect.com/science/article/pii/S1872497317302119>
- Cerutti, L, Görtler, A, Griener, T, Zelazny, A, & ... (2020). Performance and application of 16S rRNA gene cycle sequencing for routine identification of bacteria in the clinical microbiology laboratory. *Clinical microbiology* ..., Am Soc Microbiol, <https://doi.org/10.1128/cmr.00053-19>
- Cheung, SF, Yee, MF, Le, NK, Wu, BM, & ... (2018). A one-pot, isothermal DNA sample preparation and amplification platform utilizing aqueous two-phase systems. *Analytical and bioanalytical* ..., Springer, <https://doi.org/10.1007/s00216-018-1178-4>
- Choudhary, G, Kumari, S, Anu, K, & Devi, S (2024). Deciphering the microbial communities of alkaline hot spring in Panamik, Ladakh, India using a high-throughput sequencing approach. Brazilian Journal of Microbiology, Springer, <https://doi.org/10.1007/s42770-024-01346-6>
- Church, DL, Cerutti, L, Görtler, A, Griener, T, & ... (2020). Performance and application of 16S rRNA gene cycle sequencing for routine identification of bacteria in the clinical microbiology laboratory. *Clinical microbiology* ..., Am Soc Microbiol, <https://doi.org/10.1128/cmr.00053-19>
- Chusna, W., Hidayati, K., dan Noor, F. M. 2023. Analisis Pengaruh Perbedaan Background Sekolah Terhadap Pemahaman Materi IPA Mahasiswa Tadris IPA. In NCOINS: National Conference Of Islamic Natural Science 1(2): 214-223.
- Daba, GM, Elnahas, MO, & Elkhateeb, WA (2021). Contributions of exopolysaccharides from lactic acid bacteria as biotechnological tools in food, pharmaceutical, and medical applications. *International Journal of Biological* ..., Elsevier, <https://www.sciencedirect.com/science/article/pii/S0141813021001410>
- Dadwal, A, Sharma, S, & Satyanarayana, T (2021). Thermostable cellulose saccharifying microbial enzymes: Characteristics, recent advances and biotechnological applications. International Journal of Biological ..., Elsevier, <https://www.sciencedirect.com/science/article/pii/S0141813021016834>

- Damer, B, & Deamer, D (2020). The hot spring hypothesis for an origin of life. *Astrobiology*, liebertpub.com, <https://doi.org/10.1089/ast.2019.2045>
- Drozdzik, M, Busch, D, Lapczuk, J, & ... (2018). Protein abundance of clinically relevant drug-metabolizing enzymes in the human liver and intestine: a comparative analysis in paired tissue specimens. *Clinical ...*, Wiley Online Library, <https://doi.org/10.1002/cpt.967>
- Ent, F van der, Skagseth, S, Lund, BA, Sočan, J, & ... (2023). Computational design of the temperature optimum of an enzyme reaction. *Science ...*, science.org, <https://doi.org/10.1126/sciadv.ad0963>
- Fida, M, Khalil, S, Saleh, O Abu, & ... (2021). Diagnostic value of 16S ribosomal RNA gene polymerase chain reaction/Sanger sequencing in clinical practice. *Clinical Infectious ...*, academic.oup.com, <https://academic.oup.com/cid/article-abstract/73/6/961/6354408>
- Firliani, W, Agustien, A, & Febria, FA (2015). Karakterisasi bakteri termofilik penghasil enzim protease netral. *Jurnal Biologi UNAND*, jbioua.fmipa.unand.ac.id, <http://jbioua.fmipa.unand.ac.id/index.php/jbioua/article/view/112>
- Gabler, F, Nam, SZ, Till, S, Mirdita, M, & ... (2020). Protein sequence analysis using the MPI bioinformatics toolkit. ... in *Bioinformatics*, Wiley Online Library, <https://doi.org/10.1002/cbpi.108>
- Gao, D, Chundawat, SPS, Sethi, A, & ... (2013). Increased enzyme binding to substrate is not necessary for more efficient cellulose hydrolysis. *Proceedings of the ...*, National Acad Sciences, <https://doi.org/10.1073/pnas.1213426110>
- Gao, X, Lin, H, Revanna, K, & Dong, Q (2017). A Bayesian taxonomic classification method for 16S rRNA gene sequences with improved species-level accuracy. *BMC bioinformatics*, Springer, <https://doi.org/10.1186/s12859-017-1670-4>
- Gaston, DC (2023). Clinical metagenomics for infectious diseases: progress toward operational value. *Journal of Clinical Microbiology*, Am Soc Microbiol, <https://doi.org/10.1128/jcm.01267-22>
- Ghosh, AC, Duboc, C, & Gennari, M (2021). Synergy between metals for small molecule activation: Enzymes and bio-inspired complexes. *Coordination Chemistry Reviews*, Elsevier, <https://www.sciencedirect.com/science/article/pii/S0010854520306986>
- Guengerich, FP, & Yoshimoto, FK (2018). Formation and cleavage of C–C bonds by enzymatic oxidation–reduction reactions. *Chemical Reviews*, ACS Publications, <https://doi.org/10.1021/acs.chemrev.8b00031>
- Guevara, EE, Frankel, DC, Ranaivonasy, J, & ... (2018). A simple, economical protocol for DNA extraction and amplification where there is no lab. *Conservation genetics ...*, Springer, <https://doi.org/10.1007/s12686-017-0758-5>
- Gurung, N, Ray, S, Bose, S, & Rai, V (2013). A broader view: microbial enzymes and their relevance in industries, medicine, and beyond. *BioMed research ...*, Wiley Online Library, <https://doi.org/10.1155/2013/329121>

- Hardi, J., Jusman., Razak, A. R., dan Silva., 2016. Produksi Dan Uji Aktivitas Enzim Kitinase Dari Isolat Bakteri Termofilik B1211 Asal Air Panas Bora, *Jurnal Kovalen*, 2(3): 67-72.
- Hardi, J., Ruslan, R., Razak, A. R., dan Silva, S., 2017. Karakterisasi Enzim Kitinase Dari Isolat Bakteri Termofilik B1211 Asal Air Panas Bora. *Kovalen, Jurnal Riset Kimia*, 3(2): 172-179.
- Hassler, HB, Probert, B, Moore, C, Lawson, E, & ... (2022). Phylogenies of the 16S rRNA gene and its hypervariable regions lack concordance with core genome phylogenies. *Microbiome*, Springer, <https://doi.org/10.1186/s40168-022-01295-y>
- Herrialfian, H, Lubis, MMN, Darmawi, D, & ... (2021). 7. Inhibition Activity of Ethanolic Extract of Binahong Leaf (*Anredera cordifolia* (Ten.) Steenis) on *Staphylococcus aureus* Bacteria. *Jurnal Medika* ..., jurnal.usk.ac.id, <https://jurnal.usk.ac.id/JMV/article/view/9988>
- Hollenberg, PF (2022). Enzyme inhibition. *Drug Metabolism Handbook: Concepts and ...*, Wiley Online Library, <https://doi.org/10.1002/9781119851042.ch13>
- Huang, Y, Ren, J, & Qu, X (2019). Nanozymes: classification, catalytic mechanisms, activity regulation, and applications. *Chemical reviews*, ACS Publications, <https://doi.org/10.1021/acs.chemrev.8b00672>
- Huggins, LG, Colella, V, Atapattu, U, & ... (2022). Nanopore sequencing using the full-length 16S rRNA gene for detection of blood-borne bacteria in dogs reveals a novel species of hemotropic mycoplasma. *Microbiology* ..., Am Soc Microbiol, <https://doi.org/10.1128/spectrum.03088-22>
- Idris, SA, Rasak, A, & Ado, MS (2023). Isolasi Dan Identifikasi Bakteri Kitinolitik Pada Air Payau Menggunakan Media Kitin Dari Tulang Sotong. *JKEMS-Jurnal Kesehatan* ..., rumahjurnal.or.id, <http://rumahjurnal.or.id/index.php/jkems/article/view/406>
- Irianto., 2006. Mikrobiologi: Menguak Dunia Mikroorganisme. Universitas Negeri Malang.
- Jones, CB, White, JR, Ernst, SE, Sfanos, KS, & ... (2022). Incorporation of data from multiple hypervariable regions when analyzing bacterial 16S rRNA gene sequencing data. *Frontiers in* ..., frontiersin.org, <https://doi.org/10.3389/fgene.2022.799615>
- Kalidas, C, & Sangaranarayanan, MV (2023). Co-Enzymes and Vitamins. *Biophysical Chemistry: Techniques* ..., Springer, [https://doi.org/10.1007/978-3-031-37682-5\\_10](https://doi.org/10.1007/978-3-031-37682-5_10)
- Kang, TS (2019). Basic principles for developing real-time PCR methods used in food analysis: A review. *Trends in Food Science & Technology*, Elsevier, <https://www.sciencedirect.com/science/article/pii/S0924224418300074>
- Kartawiria, IS, Serafin, L, & Abimanyu, H (2019). EFFECT OF THE SUBSTRATE CONCENTRATION AND THE STIRRING RATE ON THE ENZYMATIC HYDROLYSIS OF CELLULOSE FROM PRE-TREATED .... *Journal of Chemical*.
- Koolman, J. 2005. Color atlas of biochemistry. New York.

- Kuby, SA (2019). A study of enzymes: Enzyme catalysts, kinetics, and substrate binding., taylorfrancis.com, <https://doi.org/10.1201/9780429291579>
- Kuchibhatla, DB, Sherman, WA, Chung, BYW, & ... (2014). Powerful sequence similarity search methods and in-depth manual analyses can identify remote homologs in many apparently "orphan" viral proteins. *Journal of* ..., Am Soc Microbiol, <https://doi.org/10.1128/jvi.02595-13>
- Küchler, A, Yoshimoto, M, Luginbühl, S, Mavelli, F, & ... (2016). Enzymatic reactions in confined environments. *Nature* ..., nature.com, <https://www.nature.com/articles/nnano.2016.54>
- Kumari, M, Padhi, S, Sharma, S, Phukon, LC, Singh, SP, & ... (2021). Biotechnological potential of psychrophilic microorganisms as the source of cold-active enzymes in food processing applications. 3 Biotech, Springer, <https://doi.org/10.1007/s13205-021-03008-y>
- Leiva, LE, Zegarra, V, Bange, G, & ... (2023). At the crossroad of nucleotide dynamics and protein synthesis in bacteria. ... and Molecular Biology ..., Am Soc Microbiol, <https://doi.org/10.1128/mmbr.00044-22>
- Lestari, P. 2000., Eksplorasi Enzim Termostabil dari Mikroba Termofil, Fakultas Biologi, Universitas Jendral Sudirman, Purwokerto
- Lynch, C, & Fleming, R (2019). A review of direct polymerase chain reaction of DNA and RNA for forensic purposes. *Wiley Interdisciplinary Reviews: Forensic* ..., Wiley Online Library, <https://doi.org/10.1002/wfs2.1335>
- Lyu, Y, & Scrimin, P (2021). Mimicking Enzymes: The quest for powerful catalysts from simple molecules to nanozymes. ACS Catalysis, ACS Publications, <https://doi.org/10.1021/acscatal.1c01219>
- Mahmudah, R., Baharuddin, M., dan Sappewali, S., 2016. Identifikasi Isolat Bakteri Termofilik dari Sumber Air Panas Lejja Kabupaten Soppeng, Jurnal Al-Kimia, 4(1): 31-42.
- Mai, VQ, Nhan, TA, & Hammouch, Z (2021). A mathematical model of enzymatic non-competitive inhibition by product and its applications. Physica Scripta, iopscience.iop.org, <https://doi.org/10.1088/1402-4896/ac35c6>
- Nakatsu, CH, Byappanahalli, MN, & ... (2019). Bacterial community 16S rRNA gene sequencing characterizes riverine microbial impact on Lake Michigan. *Frontiers in Microbiology*, frontiersin.org, <https://doi.org/10.3389/fmicb.2019.00996>
- Natsir, H., 2000, Karakterisasi dan Purifikasi Enzim Pendegradasi Kitin dari Mikroba Asidofilik Asal Kawah Kamojang, Tesis, Program Pascasarjana, Institut Pertanian Bogor, Bogor
- Natsir, H., 2000, Karakterisasi dan Purifikasi Enzim Pendegradasi Kitin dari Mikroba Asidofilik Asal Kawah Kamojang, Tesis, Program Pascasarjana, Institut Pertanian Bogor, Bogor
- Natsir, H., Patong, A. R., Suhartono, M. T., and Ahmad, A., 2010 A. Production and Characterization of Chitinase Enzymes From Sulili Hot Spring In South Sulawesi Bacillus sp. HSA,3-1a, Indoneia Journal Chemistry, 10(2): 256-260.
- Natsir, H., Patong, A. R., Suhartono, M. T., dan Ahmad, A., 2012. Produksi dan Aplikasi Kitinase dari B. Licheniformis HSA3-1a dalam Menghidrolisis Kitin dari Limbah Udang dan Dinding Sel Jamur Ganoderma sp. Laporan Penelitian.

- Ntarmouchant, A, Jeddi, EM, Carvalho, MR, & ... (2024). Thermal springs associated with the Melilla-Fès-Smaala-Oulmès fault (Morocco): The role of fluid geochemistry in identifying a major active geodynamic structure. *Applied* ..., Elsevier,  
<https://www.sciencedirect.com/science/article/pii/S0883292724001902>
- Nurikhsanti, M, Zulkifli, L, Rasmi, DAC, & ... (2024). Antagonistic Test of Bacteria Producing Siderophore and Protease Enzymes from The Rhizosfer of Peanut Plants on The Growth of Pathogenic Fungus *Colletotrichum* .... *Jurnal Biologi* ..., [jurnalfkip.unram.ac.id, <http://www.jurnalfkip.unram.ac.id/index.php/JBT/article/view/6459>](http://www.jurnalfkip.unram.ac.id/index.php/JBT/article/view/6459)
- Ogura, H, Okamoto, N, Nakamura, A, Takahashi, H, & ... (2024). Differences in the microbiome composition of the traditional fermented rice beverage Miki from Amami Island. *International Journal of* ..., Elsevier, <https://www.sciencedirect.com/science/article/pii/S1878450X24001045>
- Olivieri, C, Li, GC, Wang, Y, VS, M, Walker, C, Kim, J, & ... (2022). ATP-competitive inhibitors modulate the substrate binding cooperativity of a kinase by altering its conformational entropy. *Science* ..., science.org, <https://doi.org/10.1126/sciadv.abo0696>
- Pamungkas, SA, Puspita, ID, & ... (2022). Pengaruh pH, Suhu dan Jenis Substrat terhadap Aktivitas Kitinase *Bacillus* sp. RNT9. .... : Indonesian Journal of ..., ejournal.undip.ac.id, <https://ejournal.undip.ac.id/index.php/saintek/article/view/49516>
- Park, J. K., Morita, K., Fukumoto, I., Yukikazu, Y., Nakagawa, T., Kawamukai, M., and Matsuda, H., 1997. Purification And Characterization of The Chitinase (Chi A) From *Enterobacter* Sp. G-1. *Biosci. Biotech Biochem*, 61(4): 684-6.
- Pereira, R, Oliveira, J, & Sousa, M (2020). Bioinformatics and computational tools for next-generation sequencing analysis in clinical genetics. *Journal of clinical medicine*, mdpi.com, <https://www.mdpi.com/2077-0383/9/1/132>
- Pesaresi, A (2023). Mixed and non-competitive enzyme inhibition: underlying mechanisms and mechanistic irrelevance of the formal two-site model. *Journal of Enzyme Inhibition and Medicinal Chemistry*, Taylor &Francis, <https://doi.org/10.1080/14756366.2023.2245168>
- Pinney, MM, Mokhtari, DA, Akiva, E, Yabukarski, F, & ... (2021). Parallel molecular mechanisms for enzyme temperature adaptation. *Science*, science.org, <https://doi.org/10.1126/science.aay2784>
- Piumetti, M, Illanes, A, & Lygeros, N (2022). Molecular dynamics and complexity in catalysis and biocatalysis., Springer, <https://doi.org/10.1007/978-3-030-88500-7>
- Poedjiadi, A., 2006. Dasar-dasar Biokimia, UI Press, Jakarta.
- Punekar, NS, & Punekar, NS (2018). Regulation of Enzyme Activity. ENZYMES: Catalysis, Kinetics and ..., Springer, [https://doi.org/10.1007/978-981-13-0785-0\\_37](https://doi.org/10.1007/978-981-13-0785-0_37)
- Purwanto, M.G.M., 2014, Perbandingan Analisis Kadar Protein Terlarut dengan Berbagai Metode Spektroskopi UV-Visible, *Jurnal Ilmiah Sains dan Teknologi*, 7(2); 1-71.

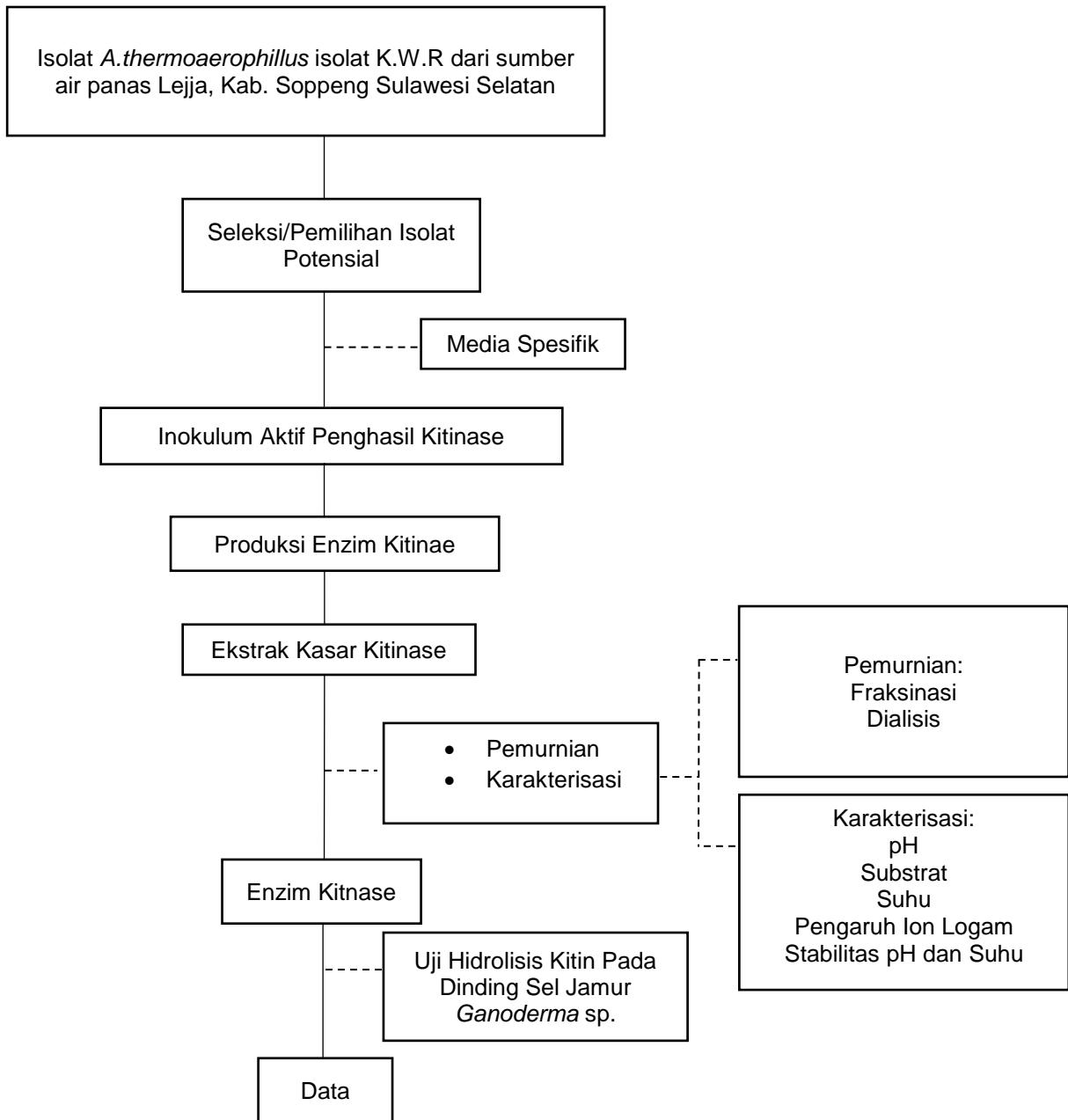
- Regueira-Iglesias, A, Balsa-Castro, C, & ... (2023). Critical review of 16S rRNA gene sequencing workflow in microbiome studies: From primer selection to advanced data analysis. ... *Oral Microbiology*, Wiley Online Library, <https://doi.org/10.1111/omi.12434>
- Rueda, N, Santos, JCS Dos, Ortiz, C, & ... (2016). Chemical modification in the design of immobilized enzyme biocatalysts: drawbacks and opportunities. The Chemical ..., Wiley Online Library, <https://doi.org/10.1002/tcr.201600007>
- Sakinah, AMM, Ismail, AF, Illias, RM, & ... (2014). Effect of substrate and enzyme concentration on cyclodextrin production in a hollow fibre membrane reactor system. Separation and ..., Elsevier, <https://www.sciencedirect.com/science/article/pii/S1383586614000136>
- Salaikumaran, MR, Badiger, VP, & Burra, VLSP (2022). 16S rRNA methyltransferases as novel drug targets against tuberculosis. *The Protein Journal*, Springer, <https://doi.org/10.1007/s10930-021-10029-2>
- Sarni, Natsir, H., dan Dali, S., 2015, Produksi dan Karakterisasi Enzim Kitosanase dari Isolat Bakteri Klebsiella sp, Jurnal Techno, 4(2): 8-15.
- Sarni, S (2023). Desain Dan Aplikasi Biosensor Menggunakan Enzim Urikase Dari Bakteri Termofil Mataumpana Untuk Analisis Asam Urat Pada ...., Repository.Unhas.Ac.Id, [Http://Repository.Unhas.Ac.Id/Id/Eprint/31072/3/H013191005\\_Disertasi\\_30-10-2023%20dp.Pdf](Http://Repository.Unhas.Ac.Id/Id/Eprint/31072/3/H013191005_Disertasi_30-10-2023%20dp.Pdf)
- Seetharaman, R, Advani, M, Mali, S, & Pawar, S (2020). Enzymes as targets of Drug Action: an Overview. International Journal, researchgate.net, [https://www.researchgate.net/profile/Rajmohan-Seetharaman/publication/341625530\\_Ezymes\\_as\\_targets\\_of\\_Drug\\_Action\\_an\\_Overview/links/5ecc04f7458515626ccc497c/Enzymes-as-targets-of-Drug-Action-an-Overview.pdf](https://www.researchgate.net/profile/Rajmohan-Seetharaman/publication/341625530_Ezymes_as_targets_of_Drug_Action_an_Overview/links/5ecc04f7458515626ccc497c/Enzymes-as-targets-of-Drug-Action-an-Overview.pdf)
- Setiarto, R. H. B., dan Karo, M. S. D. M. B., 2021. Pengantar Biokimia Klinis. GUEPEDIA.
- Shmidt, AA, & Egorova, TV (2021). PCR-based analytical methods for quantification and quality control of recombinant adeno-associated viral vector preparations. *Pharmaceuticals*, mdpi.com, <https://www.mdpi.com/1424-8247/15/1/23>
- Siliakus, MF, Oost, J van der, & Kengen, SWM (2017). Adaptations of archaeal and bacterial membranes to variations in temperature, pH and pressure. *Extremophiles*, Springer, <https://doi.org/10.1007/s00792-017-0939-x>
- Singla, P, & Bhardwaj, RD (2020). Enzyme promiscuity—A light on the “darker” side of enzyme specificity. *Biocatalysis and Biotransformation*, Taylor & Francis, <https://doi.org/10.1080/10242422.2019.1696779>
- Sousa, SF, Calixto, AR, Ferreira, P, Ramos, MJ, & ... (2020). Activation free energy, substrate binding free energy, and enzyme efficiency fall in a very narrow range of values for most enzymes. ... *Catalysis*, ACS Publications, <https://doi.org/10.1021/acscatal.0c01947>

- Stover, NA, & Cavalcanti, ARO (2017). Using NCBI BLAST. *Current Protocols Essential ...*, Wiley Online Library, <https://doi.org/10.1002/cpet.8>
- Suryadi, Y., Priyanto, T. P., Susilowati, D. N., Samudra, I. M., Yudhistira, N., dan Purwakusumah, E. D., 2013. Isolasi dan Karakterisasi Kitinase Asal Bacillus Cereus 11 UJ, Jurnal Biologi Indonesia, 9(1): 51-62.
- Susanto, T. E., Wardani, Y. A. K., dan Sutrisno, A., 2013. Enzim Kitinase dan Aplikasi di Bidang Industri: Kajian Pustaka, Jurnal Pangan Dan Agroindustri, 3(3): 878-887.
- Tabassum, O (2022). Screening of carbapenemases in Acinetobacter species.., dspace.qau.edu.pk,  
<http://dspace.qau.edu.pk:8080/jspui/handle/123456789/28336>
- Teng, M, Young, DW, & Tan, Z (2022). The Pursuit of Enzyme Activation: A Snapshot of the Gold Rush. Journal of Medicinal Chemistry, ACS Publications, <https://doi.org/10.1021/acs.jmedchem.2c01291>
- Thapa, S, Li, H, OHair, J, Bhatti, S, Chen, FC, & ... (2019). Biochemical characteristics of microbial enzymes and their significance from industrial perspectives. Molecular ..., Springer, <https://doi.org/10.1007/s12033-019-00187-1>
- Vijayakumar, T, Divya, B, Vasantha, V, & ... (2023). Diagnostic utility of gram stain for oral smears—A review. Journal of Microscopy ..., journals.lww.com, [https://journals.lww.com/jmcu/\\_layouts/15/oaks.journals/downloadpdf.aspx?an=02087478-202311030-00002](https://journals.lww.com/jmcu/_layouts/15/oaks.journals/downloadpdf.aspx?an=02087478-202311030-00002)
- Wang, J, Tavakoli, J, & Tang, Y (2019). Bacterial cellulose production, properties and applications with different culture methods—A review. Carbohydrate polymers, Elsevier,  
<https://www.sciencedirect.com/science/article/pii/S0144861719305041>
- Washburne, AD, Morton, JT, Sanders, J, & ... (2018). Methods for phylogenetic analysis of microbiome data. Nature ..., nature.com, <https://www.nature.com/articles/s41564-018-0156-0>
- Yan, Y, Zhang, Z, Zhou, X, Wang, G, He, M, Tian, J, & ... (2024). Geochemical characteristics of hot springs in active fault zones within the northern Sichuan-Yunnan block: Geochemical evidence for tectonic activity. Journal of ..., Elsevier,  
<https://www.sciencedirect.com/science/article/pii/S0022169424005742>
- Yang, M, Derbyshire, MK, Yamashita, RA, & ... (2020). NCBI's conserved domain database and tools for protein domain analysis. Current protocols in ..., Wiley Online Library, <https://doi.org/10.1002/cpbi.90>
- Yusriadi, Ahmad, A., Khaerah, N., Arfah, R., Karim, A., dan Karim, H., 2019, Isolation, characterization and anticancer potential test of crude extract of L-asparaginase enzyme from siam weed leaf (Chromolaena odorata Linn): A novel source. Journal of Physics: Conference Series, 1341(3): 1-10.
- Zhang, X, Hao, T, Zhang, T, Hu, Y, Lu, R, Li, D, & ... (2024). Towards energy conservation and carbon reduction for wastewater treatment processes: A review of carbon-neutral anaerobic biotechnologies. Journal of Water ..., Elsevier,  
<https://www.sciencedirect.com/science/article/pii/S2214714424002587>

- Zhu, H, Zhang, H, Xu, Y, Laššáková, S, Korabečná, M, & ... (2020). PCR past, present and future. ..., Taylor & Francis, <https://doi.org/10.2144/btn-2020-0057>
- Zielezinski, A, Grgis, HZ, Bernard, G, Leimeister, CA, & ... (2019). Benchmarking of alignment-free sequence comparison methods. *Genome biology*, Springer, <https://doi.org/10.1186/s13059-019-1755-7>
- Zilda, D.S., dan Fawzya, Y.N., dan Chasanah, E., 2006, Karakterisasi Enzim Kitosanase dari Bakteri Kitinolitik T5a1 yang Diisolasi dari Terasi, Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan, 1(1): 43-50.

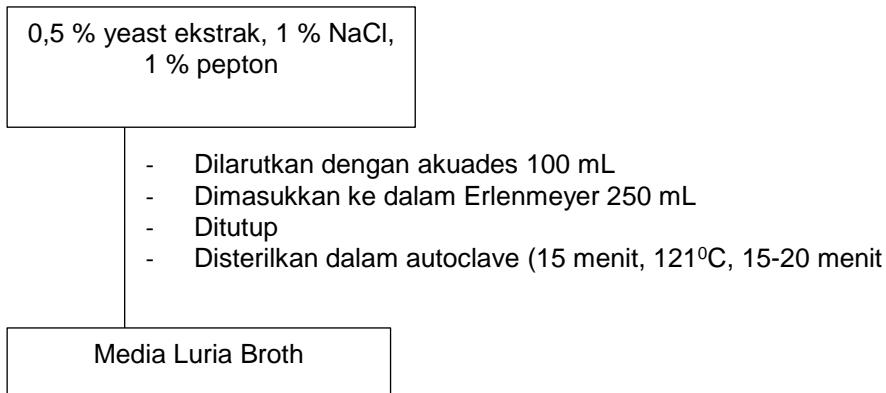
## LAMPIRAN

**Lampiran 1.** Diagram alir penelitian

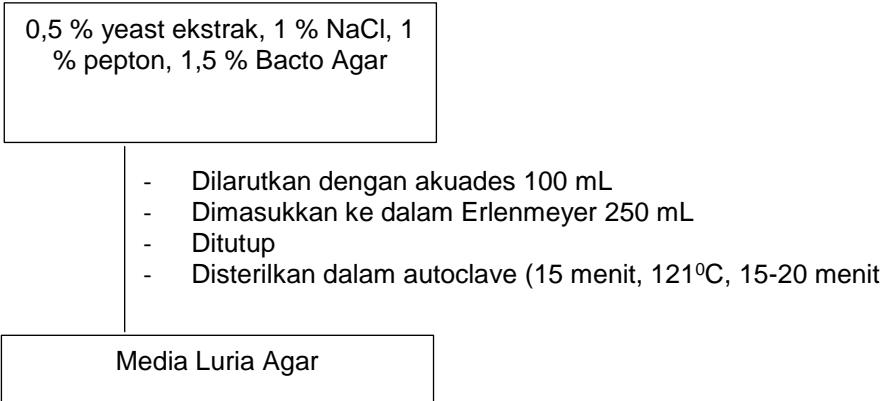


## Lampiran 2. Prosedur Kerja

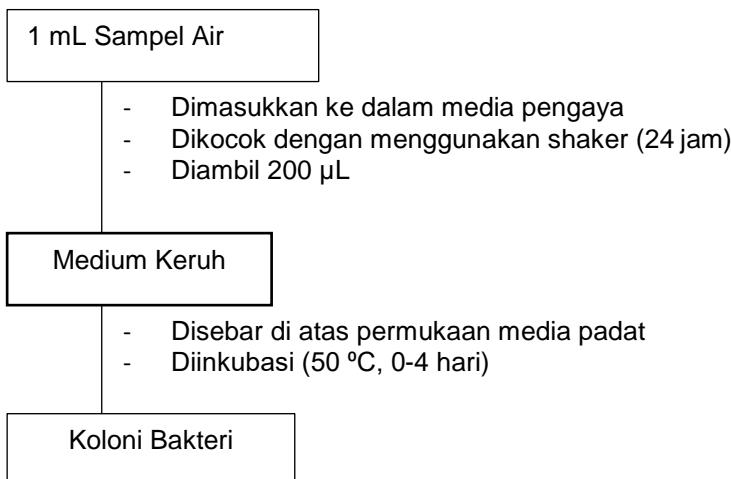
### Pembuatan Medium Luria Broth



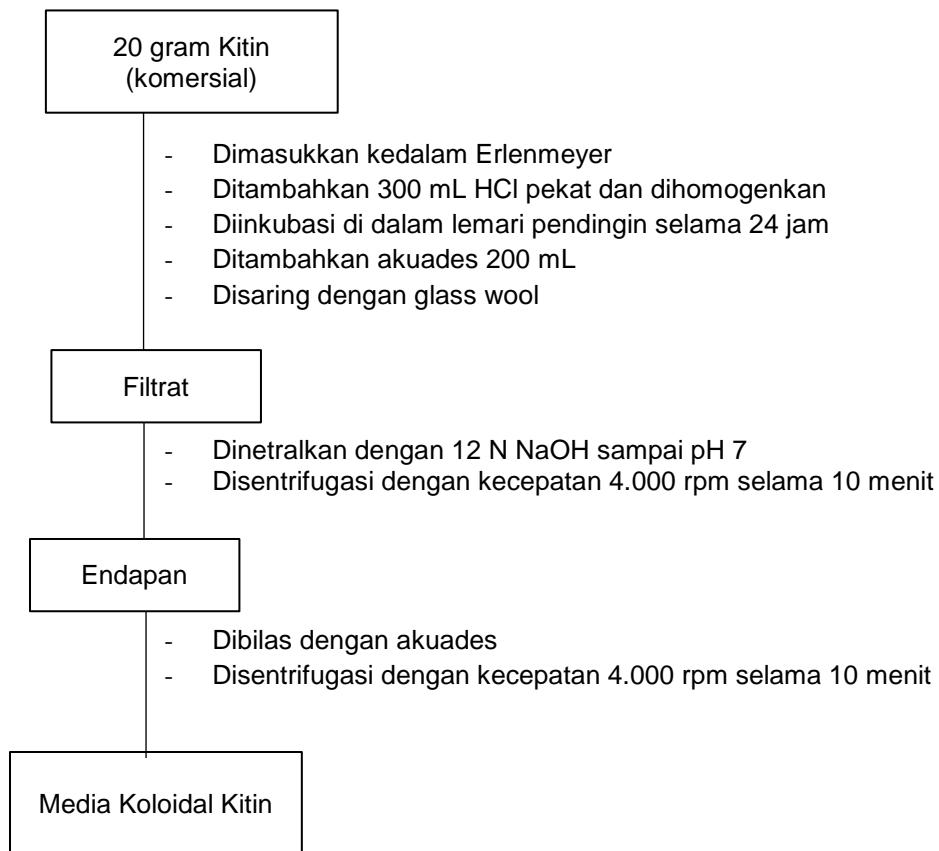
### Pembuatan Media Luria Agar



### Isolasi Mikroba dari Sumber Air Panas

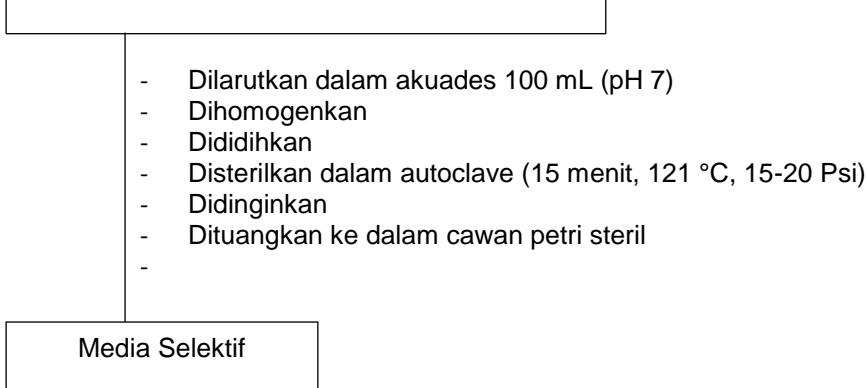


### Pembuatan Substrat Koloidal Kitin

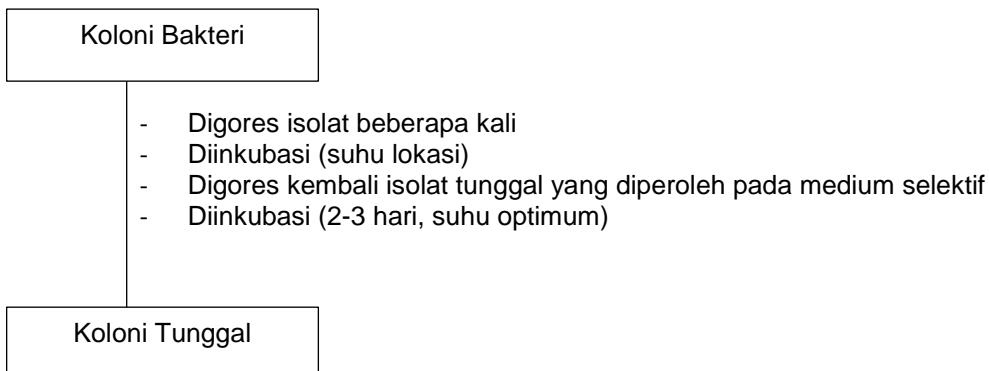


### Pembuatan Media Selektif Kitinolitik

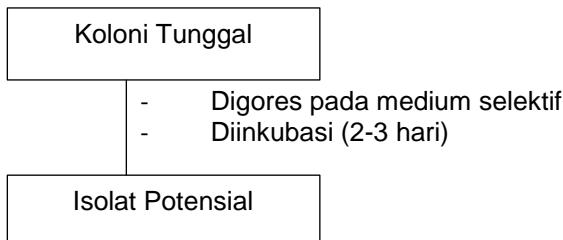
yeast ekstrak 0,1 %, pepton 0,1 %,  $\text{CaCl}_2$  0,01 %,  $\text{KH}_2\text{PO}_4$  0,01 %,  $\text{NaCl}$  0,1 %,  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$  0,01 %, koloidal kitin 0,5 %



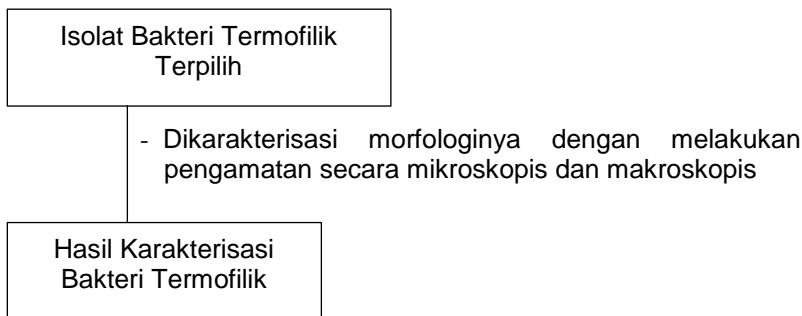
## Pemurnian Bakteri



## Pemilihan Isolat Potensial



## Uji Morfologi Bakteri Penghasil Enzim Kitinase



## Pembuatan dan Penyiapan Inokulum

yeast ekstrak 0,5 %, pepton 0,01 %, ammonium sulfat,  $\text{CaCl}_2$  0,01 %,  $\text{KH}_2\text{PO}_4$  0,01 %,  $\text{NaCl}$  0,1 %,  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$  0,01 % dan koloidal kitin 0,5 %

- Dilarutkan dalam akuades 100 mL
- Dihomogenkan, dipanaskan, dituang ke dalam Erlenmeyer steril
- Diambil 2 hingga 3 ose bakteri yang telah tumbuh dengan baik
- Diinokulasikan ke dalam medium inokulum yang telah disiapkan
- Dikocok ke dalam shaker (40 °C, 180 rpm, 18-24 jam) untuk diinokulasikan lebih lanjut dalam medium produksi

Media Inokulum  
Aktif

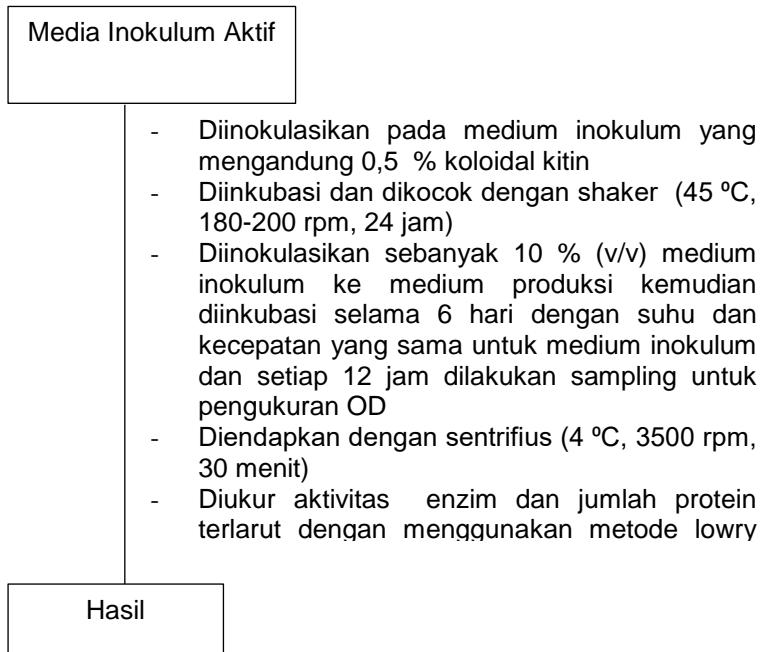
## Pembuatan Medium Produksi

amonium sulfat 0,7 %, yeast ekstrak 0,05 %, bakto pepton 0,1 %,  $\text{NaCl}$  0,1 %,  $\text{KH}_2\text{PO}_4$  0,01 %,  $\text{CaCl}$  0,01 %,  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$  0,01 %, koloidal kitin 0,5 %

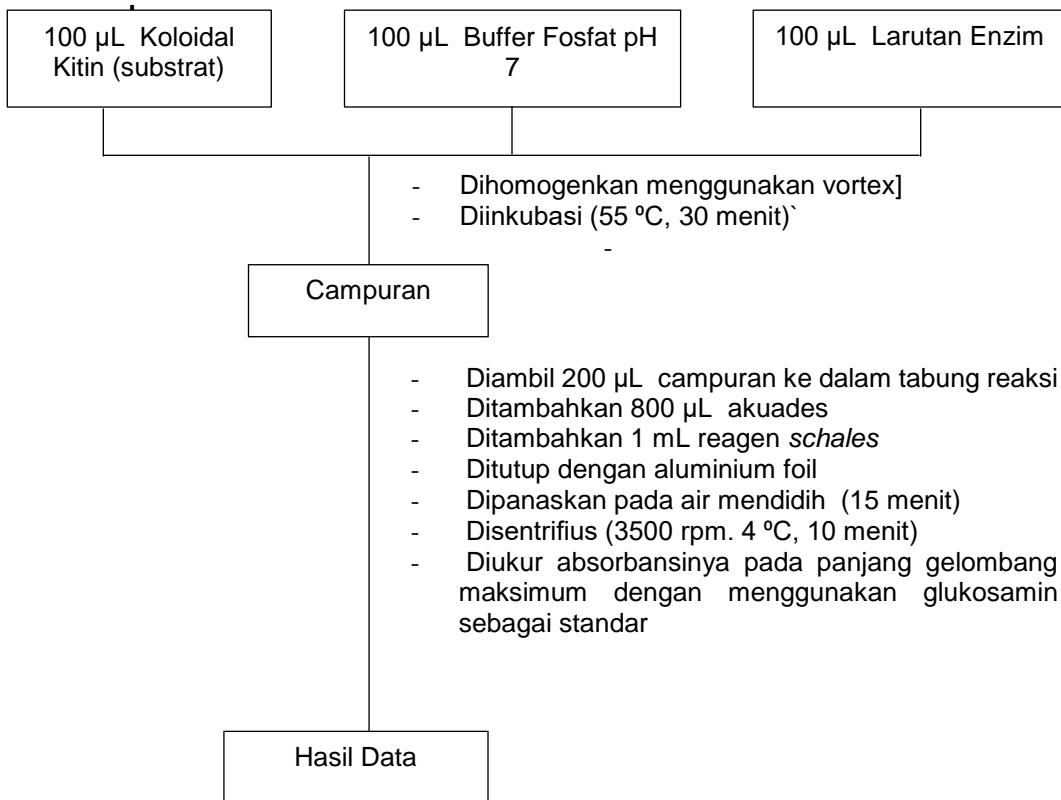
- Dilarutkan dengan akuades 500 mL
- Dihomogenkan
- Disterilkan dengan *autoclave*.

Medium Produksi

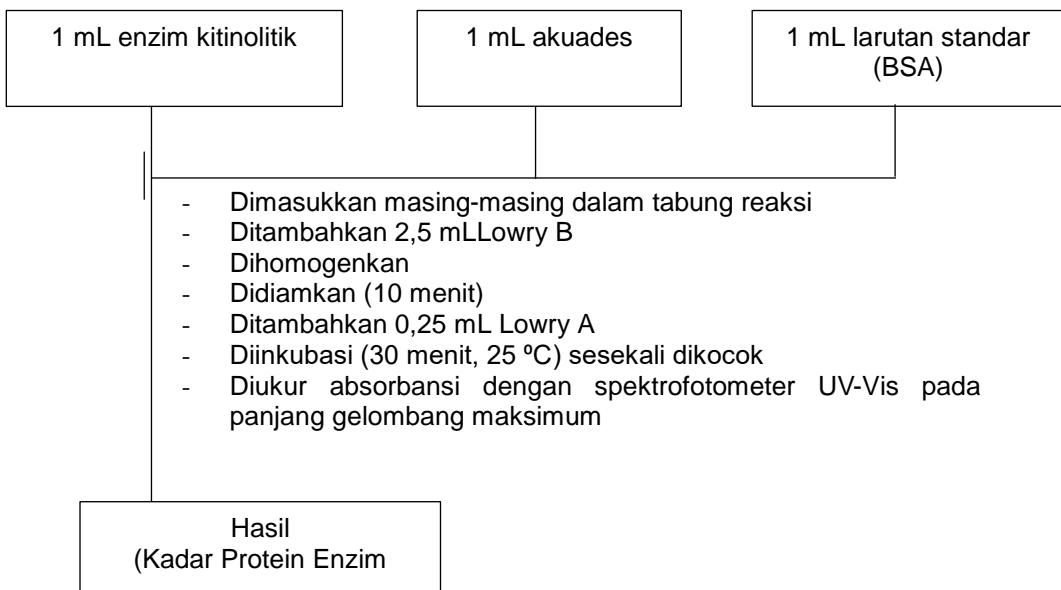
## Produksi Ekstrak Kitinase



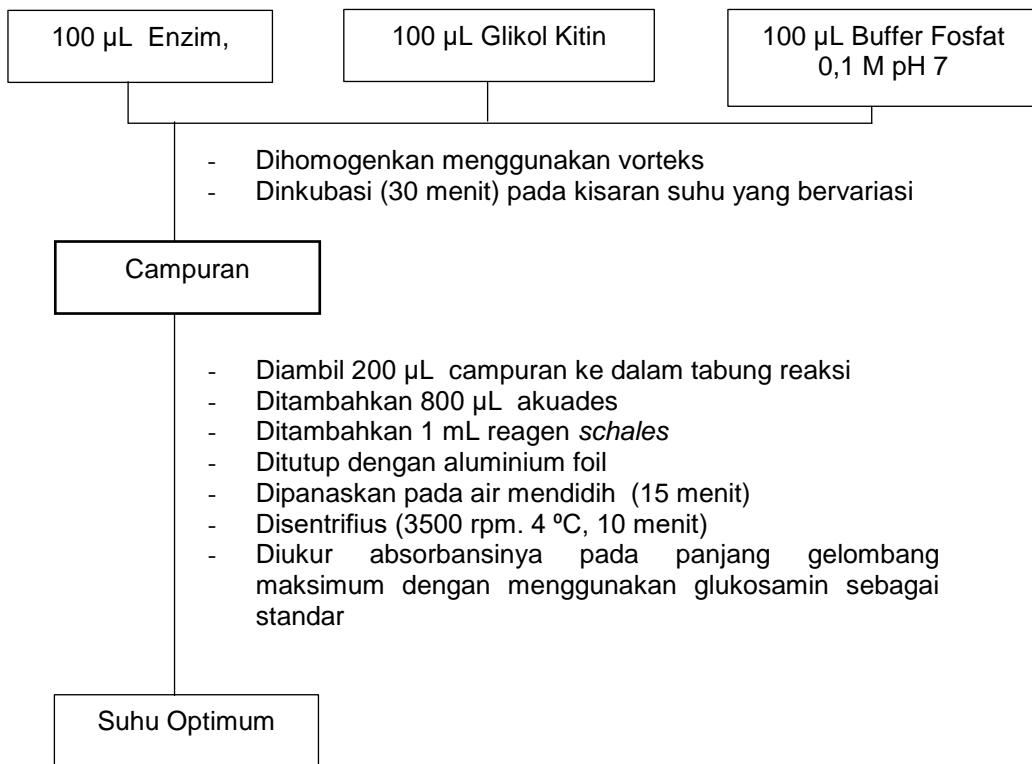
### Pengukuran Aktivitas Enzim Kitinase (Natsir et al., 2014)



### Penentuan Kadar Protein dengan Metode Lowry



## Karakterisasi Enzim Kitinase dari Mikroba Termofil Penentuan Suhu Optimum



**Lampiran 3.** Tabel Fraksinasi Ammonium Sulfat

Konsentrasi awal dari ammonium sulfat (% kejemuhan pada 0°C)	% Kejemuhan pada 0°C																
	20	25	30	35	40	45	50	55	60	65	70	75	80	85	90	95	100
<b>Penambahan ammonium sulfat kristal (gram) untuk pada 1 liter larutan</b>																	
0	106	134	164	194	226	258	291	326	361	398	436	476	516	559	603	650	697
5	79	108	137	166	197	229	262	296	331	368	405	444	484	526	570	615	662
10	53	81	109	139	169	200	233	266	301	337	374	412	452	493	536	581	627
15	26	54	82	111	141	172	204	237	271	306	343	381	420	460	503	547	592
20	0	27	55	83	113	143	175	207	241	276	312	349	387	427	469	512	557
25		0	27	56	84	115	146	179	211	245	280	317	355	395	436	478	522
30			0	28	56	86	117	148	181	214	249	285	323	362	402	445	488
35				0	28	57	87	118	151	184	218	254	291	329	369	410	453
40					0	29	58	89	120	153	187	222	258	296	335	376	418
45						0	29	59	90	123	156	190	226	263	302	342	383
50							0	30	60	92	125	159	194	230	268	308	348
55								0	30	61	93	127	161	197	235	273	313
60									0	31	62	95	129	164	201	239	279
65										0	31	63	97	132	168	205	244
70											0	32	65	99	134	171	209
75												0	32	66	101	137	174
80													0	33	67	103	139
85														0	34	68	105
90															0	34	70

**Lampiran 4.** Data

Hubungan antara OD dan aktivitas kitinase

<b>Waktu inkubasi (jam)</b>	<b>Absorbansi (660nm)</b>	<b>Aktivitas kitinase (U/mL)</b>
0	0.065	0.002
12	0.097	0.003
24	0.252	0.018
36	0.435	0.090
48	0.452	0.167
60	0.427	0.049
72	0.327	0.031
84	0.300	0.015
96	0.261	0.009
108	0.189	0.003
120	0.021	0.001

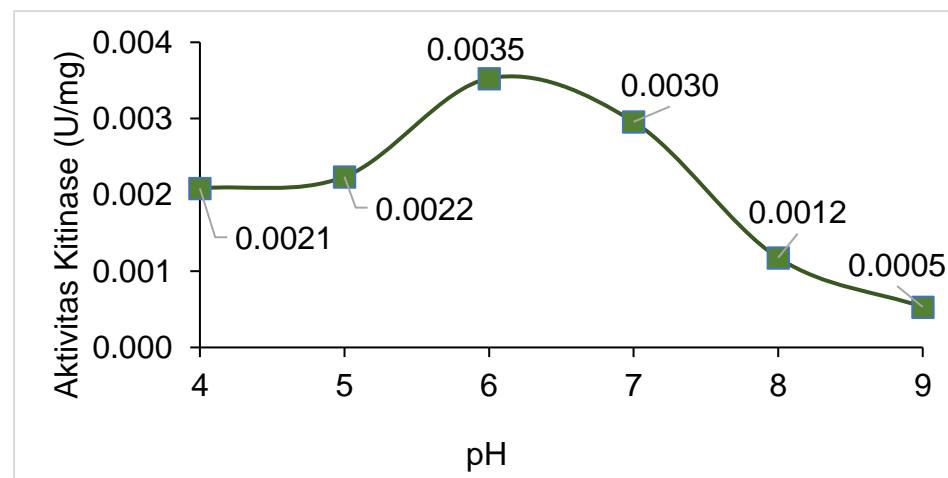
Hubungan antara kadar protein dan aktivitas spesifik

<b>Waktu inkubasi (jam)</b>	<b>Kadar protein (mg/mL)</b>	<b>Aktivitas spesifik (U/mg)</b>
0	0.357	0.006
12	0.406	0.008
24	0.629	0.029
36	0.748	0.121
48	0.937	0.178
60	1.282	0.038
72	1.251	0.025
84	0.885	0.017
96	0.866	0.010
108	0.677	0.005

## Karakterisasi pH

Konsentrasi pH	Abs Sampel	Abs Kontrol	Konsentrasi N-Ac-Glu Sampel	Konsentrasi N-Ac-Glu Kontrol	xs-xk	1000/200	600/400	1/30.	1/221.21	aktivitas
4	0.356	0.411	8.5738	6.7282	1.8456	5.0000	1.5000	0.0333	0.0045	0.0021
5	0.326	0.385	9.5805	7.6007	1.9799	5.0000	1.5000	0.0333	0.0045	0.0022
6	0.322	0.415	9.7148	6.5940	3.1208	5.0000	1.5000	0.0333	0.0045	0.0035
7	0.312	0.39	10.0503	7.4329	2.6174	5.0000	1.5000	0.0333	0.0045	0.0030
8	0.348	0.379	8.8423	7.8020	1.0403	5.0000	1.5000	0.0333	0.0045	0.0012
9	0.357	0.371	8.5403	8.0705	0.4698	5.0000	1.5000	0.0333	0.0045	0.0005

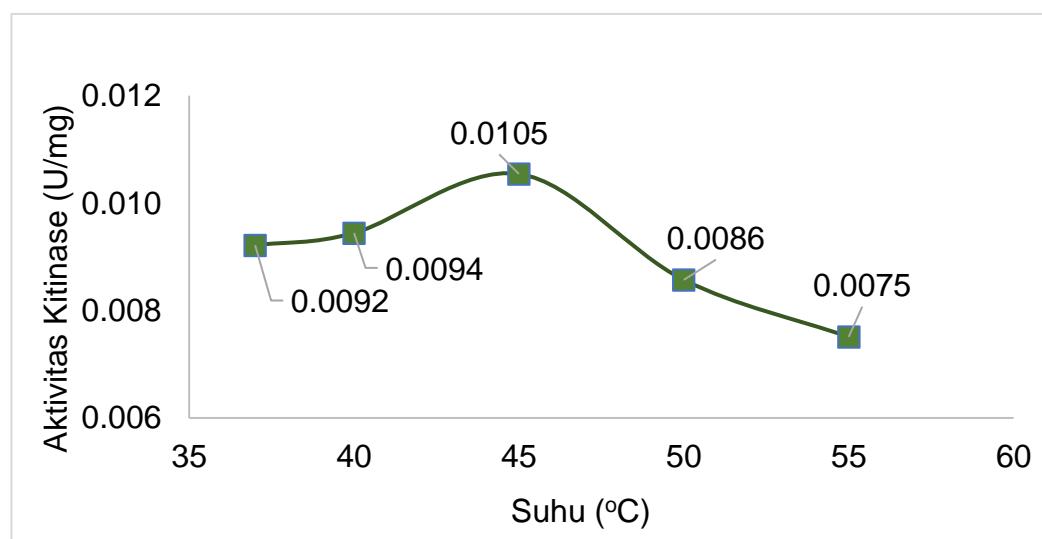
Konsentrasi pH	aktivitas
4	0.0021
5	0.0022
6	0.0035
7	0.0030
8	0.0012
9	0.0005



### Karakterisasi suhu

Suhu	Abs Sampel	Abs Kontrol	Konsentrasi N-Ac-Glu Sampel	Konsentrasi N-Ac-Glu Kontrol	xs-xk	1000/200	600/400	1/30.	1/221.21	aktivitas
37	0.235	0.478	12.6342	4.4799	8.1544	5.0000	1.5000	0.0333	0.0045	0.0092
40	0.223	0.472	13.0369	4.6812	8.3557	5.0000	1.5000	0.0333	0.0045	0.0094
45	0.213	0.491	13.3725	4.0436	9.3289	5.0000	1.5000	0.0333	0.0045	0.0105
50	0.226	0.452	12.9362	5.3523	7.5839	5.0000	1.5000	0.0333	0.0045	0.0086
55	0.223	0.421	13.0369	6.3926	6.6443	5.0000	1.5000	0.0333	0.0045	0.0075

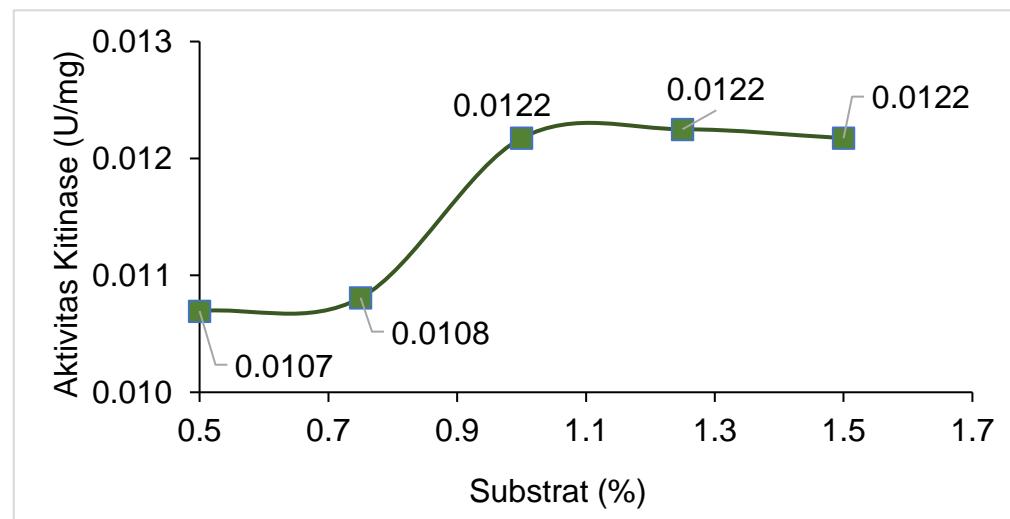
Suhu	aktivitas
37	0.0092
40	0.0094
45	0.0105
50	0.0086
55	0.0075



### Penentuan substrat optimum

Konsentrasi Substrat (%)	Abs Sampel	Abs Kontrol	Konsentrasi N-Ac-Glu Sampel	Konsentrasi N-Ac-Glu Kontrol	xs-xk	1000/200	600/400	1/30.	1/221.21	aktivitas
0.5	0.239	0.521	12.5000	3.0369	9.4631	5.0000	1.5000	0.0333	0.0045	0.0107
0.75	0.242	0.527	12.3993	2.8356	9.5638	5.0000	1.5000	0.0333	0.0045	0.0108
1	0.262	0.583	11.7282	0.9564	10.7718	5.0000	1.5000	0.0333	0.0045	0.0122
1.25	0.244	0.567	12.3322	1.4933	10.8389	5.0000	1.5000	0.0333	0.0045	0.0122
1.5	0.233	0.554	12.7013	1.9295	10.7718	5.0000	1.5000	0.0333	0.0045	0.0122

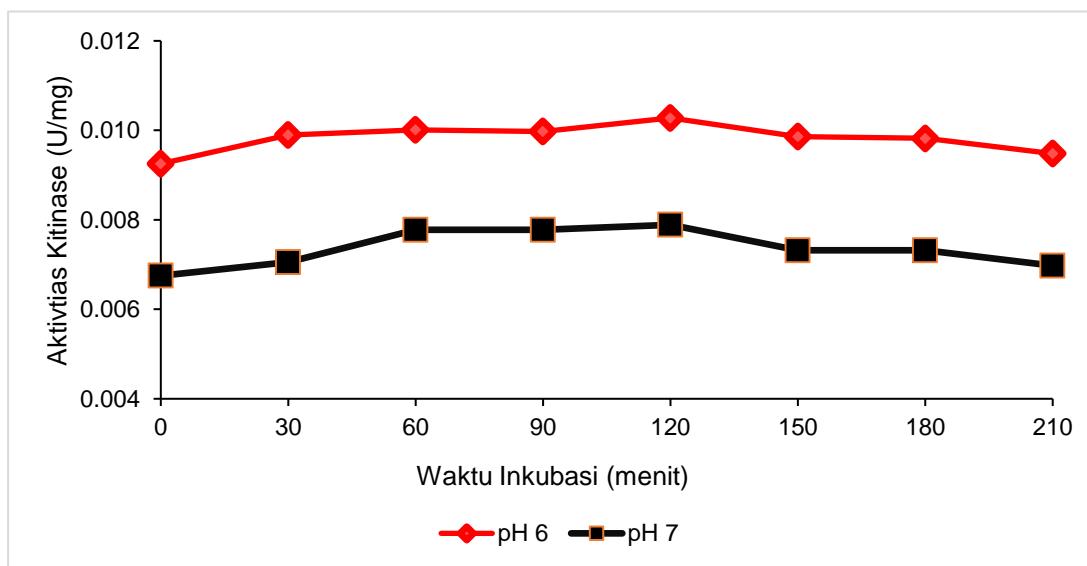
Konsentrasi Substrat (%)	aktivitas
0.5	0.0107
0.75	0.0108
1	0.0122
1.25	0.0122
1.5	0.0122



## Stabilitas pH

Stabilitas kitinase pH 6											
Waktu inkubasi(menit)	Absorbansi Sampel	Absorbansi Kontrol	Konsentrasi N-Ac-Glu Sampel	Konsentrasi N-Ac-Glu Kontrol	xs-xk	1000/200	600/400	1/30.	1/221.21	aktivitas	
0	0.311	0.555	10.0839	1.8960	8.1879	5.0000	1.5000	0.0333	0.0045	0.0093	
30	0.324	0.585	9.6477	0.8893	8.7584	5.0000	1.5000	0.0333	0.0045	0.0099	
60	0.323	0.587	9.6812	0.8221	8.8591	5.0000	1.5000	0.0333	0.0045	0.0100	
90	0.328	0.591	9.5134	0.6879	8.8255	5.0000	1.5000	0.0333	0.0045	0.0100	
120	0.346	0.617	8.9094	-0.1846	9.0940	5.0000	1.5000	0.0333	0.0045	0.0103	
150	0.325	0.585	9.6141	0.8893	8.7248	5.0000	1.5000	0.0333	0.0045	0.0099	
180	0.326	0.585	9.5805	0.8893	8.6913	5.0000	1.5000	0.0333	0.0045	0.0098	
210	0.322	0.572	9.7148	1.3255	8.3893	5.0000	1.5000	0.0333	0.0045	0.0095	
Stabilitas kitinase pH 7											
Waktu inkubasi(menit)	Absorbansi Sampel	Absorbansi Kontrol	Konsentrasi N-Ac-Glu Sampel	Konsentrasi N-Ac-Glu Kontrol	xs-xk	1000/200	600/400	1/30.	1/221.21	aktivitas	
0	0.283	0.461	11.0235	5.0503	5.9732	5.0000	1.5000	0.0333	0.0045	0.0068	
30	0.279	0.465	11.1577	4.9161	6.2416	5.0000	1.5000	0.0333	0.0045	0.0071	
60	0.298	0.503	10.5201	3.6409	6.8792	5.0000	1.5000	0.0333	0.0045	0.0078	
90	0.294	0.499	10.6544	3.7752	6.8792	5.0000	1.5000	0.0333	0.0045	0.0078	
120	0.317	0.525	9.8826	2.9027	6.9799	5.0000	1.5000	0.0333	0.0045	0.0079	
150	0.292	0.485	10.7215	4.2450	6.4765	5.0000	1.5000	0.0333	0.0045	0.0073	
180	0.303	0.496	10.3523	3.8758	6.4765	5.0000	1.5000	0.0333	0.0045	0.0073	
210	0.279	0.463	11.1577	4.9832	6.1745	5.0000	1.5000	0.0333	0.0045	0.0070	

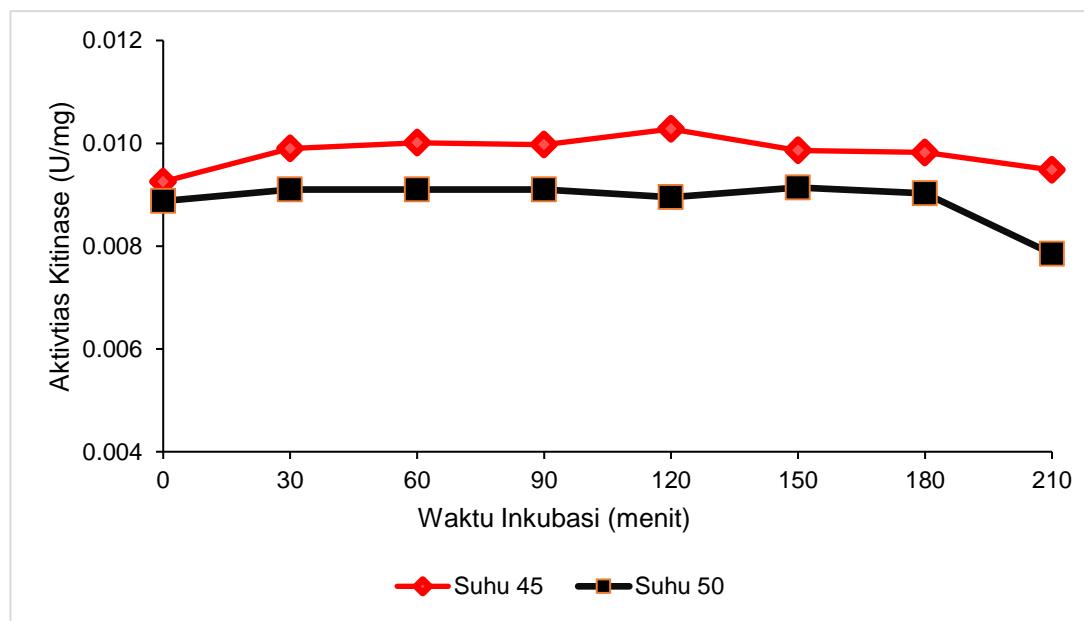
Waktu inkubasi(menit)	pH 6	pH 7
0	0.0093	0.0068
30	0.0099	0.0071
60	0.0100	0.0078
90	0.0100	0.0078
120	0.0103	0.0079
150	0.0099	0.0073
180	0.0098	0.0073
210	0.0095	0.0070



## Stabilitas suhu

Suhu 45										
Absorbansi Sampel	Absorbansi Kontrol	Konsentrasi N-Ac-Glu Sampel	Konsentrasi N-Ac-Glu Kontrol	xs-xk	1000/200	600/400	1/30.	1/221.21	aktivitas	
0.311	0.555	10.0839	1.8960	8.1879	5.0000	1.5000	0.0333	0.0045	0.0093	
0.324	0.585	9.6477	0.8893	8.7584	5.0000	1.5000	0.0333	0.0045	0.0099	
0.323	0.587	9.6812	0.8221	8.8591	5.0000	1.5000	0.0333	0.0045	0.0100	
0.328	0.591	9.5134	0.6879	8.8255	5.0000	1.5000	0.0333	0.0045	0.0100	
0.346	0.617	8.9094	-0.1846	9.0940	5.0000	1.5000	0.0333	0.0045	0.0103	
0.325	0.585	9.6141	0.8893	8.7248	5.0000	1.5000	0.0333	0.0045	0.0099	
0.326	0.585	9.5805	0.8893	8.6913	5.0000	1.5000	0.0333	0.0045	0.0098	
0.322	0.572	9.7148	1.3255	8.3893	5.0000	1.5000	0.0333	0.0045	0.0095	
Suhu 50										
Absorbansi Sampel	Absorbansi Kontrol	Konsentrasi N-Ac-Glu Sampel	Konsentrasi N-Ac-Glu Kontrol	xs-xk	1000/200	600/400	1/30.	1/221.21	aktivitas	
0.321	0.555	9.7483	1.8960	7.8523	5.0000	1.5000	0.0333	0.0045	0.0089	
0.324	0.564	9.6477	1.5940	8.0537	5.0000	1.5000	0.0333	0.0045	0.0091	
0.324	0.564	9.6477	1.5940	8.0537	5.0000	1.5000	0.0333	0.0045	0.0091	
0.326	0.566	9.5805	1.5268	8.0537	5.0000	1.5000	0.0333	0.0045	0.0091	
0.322	0.558	9.7148	1.7953	7.9195	5.0000	1.5000	0.0333	0.0045	0.0090	
0.327	0.568	9.5470	1.4597	8.0872	5.0000	1.5000	0.0333	0.0045	0.0091	
0.33	0.568	9.4463	1.4597	7.9866	5.0000	1.5000	0.0333	0.0045	0.0090	
0.355	0.562	8.6074	1.6611	6.9463	5.0000	1.5000	0.0333	0.0045	0.0079	

Suhu 45	Suhu 50
0.0093	0.0089
0.0099	0.0091
0.0100	0.0091
0.0100	0.0091
0.0103	0.0090
0.0099	0.0091
0.0098	0.0090
0.0095	0.0079



**Lampiran 5. rRNA\_typestrains/16S\_ribosomal\_RNA 16S ribosomal RNA (Bacteria and Archaea type strains)**

Job Title:4318396\_K.W.R.4.1

Program: BLASTN

Query: 4318396\_K.W.R ID: Icl|Query\_152129(dna) Length: 702

Database: rRNA\_typestrains/16S\_ribosomal\_RNA 16S ribosomal RNA (Bacteria and Archaea type strains)

Sequences producing significant alignments:

Description	Scientific Name	Common Name
Aneurinibacillus thermoerophilus strain DSM 18154 16S ribosomal RNA, partial...	Aneurinibaci...	NA
Aneurinibacillus thermoerophilus strain L420-91 16S ribosomal...	Aneurinibaci...	NA
Aneurinibacillus sediminis strain 1-18M-8-7-50 16S ribosomal...	Aneurinibaci...	NA
Aneurinibacillus danicus strain DB4 16S ribosomal RNA, partial...	Aneurinibaci...	NA
Aneurinibacillus danicus strain NBRC 102444 16S ribosomal RNA,...	Aneurinibaci...	NA
Aneurinibacillus humi strain U33 16S ribosomal RNA, partial...	Aneurinibaci...	NA
Aneurinibacillus aneurinilyticus ATCC 12856 strain DSM 5562 16...	Aneurinibaci...	NA
Aneurinibacillus aneurinilyticus ATCC 12856 strain DSM 5562 16...	Aneurinibaci...	NA
Aneurinibacillus migulanus strain BB278 16S ribosomal RNA,...	Aneurinibaci...	NA
Aneurinibacillus aneurinilyticus strain Murayama 16S ribosomal...	Aneurinibaci...	NA

Taxid	Max Score	Total Score	Query cover	E Value	Per. Ident	Acc. Len	Accession
143495	1234	1234	99%	0.0	98.57	1491	NR_112216.1
143495	1229	1229	99%	0.0	98.43	1485	NR_029383.1
1776164	1099	1099	99%	0.0	95.26	1414	NR_157740.1
267746	1070	1070	97%	0.0	94.90	1417	NR_028657.1
267746	1066	1066	97%	0.0	94.75	1501	NR_114088.1
1515627	1066	1066	99%	0.0	94.40	1451	NR_169383.1
649747	1050	1050	96%	0.0	94.57	1489	NR_112215.1
649747	1050	1050	96%	0.0	94.57	1489	NR_112283.1
47500	1050	1050	96%	0.0	94.57	1483	NR_036799.1
1391	1050	1050	96%	0.0	94.57	1482	NR_036798.1

>Aneurinibacillus thermoerophilus strain DSM 10154 16S ribosomal RNA, partial sequence

Sequence ID: NR\_112216.1 Length: 1491

Range 1: 47 to 743

Score:1234 bits(668), Expect:0.0,

Identities:690/700(99%), Gaps:4/700(0%), Strand: Plus/Plus

Query 4	GGA-TGCTTGGCCTTCTGAGGTTAGCGGGGGACGGGTGAGTAACACGTAGGCAGCTGC	62
Sbjct 47	GGAGTGCTT-GCATTCCTGAGGTTAGCGGGGGACGGGTGAGTAACACGTAGGCAGCTGC	105
Query 63	CTGTACGACCGGGATAACTCCGGAAACCGGAGGCTAATACCGGATAGGATGCCGAACCGC	122
Sbjct 106	CTGTACGACCGGGATAACTCCGGAAACCGGAGGCTAATACCGGATAGGATGCCGAACCGC	165
Query 123	ATGGTTCGGCATGGAAAGGGCTTTGAGCCCGTACAGATGGGCTGCGGCCATTAGCTG	182
Sbjct 166	ATGGTTCGGCATGGAAAGGGCTTTGAGCCCGTACAGATGGGCTGCGGCCATTAGCTA	225
Query 183	GTTGGTGGGTAACGGCTTACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATACG	242
Sbjct 226	GTTGGTGGGTAACGGCTTACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATACG	284
Query 243	GCC4CACTGGGACTGAGACACGGCTCCAGACTCTACGGGAGGCAGCAGTAGGGAATCTT	302
Sbjct 285	GCC4CACTGGGACTGAGACACGGC-CCAGACTCTACGGGAGGCAGCAGTAGGGAATCTT	343
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Sbjct 344	CCGCAATGGACGAAAGTCTGACGGAGCAACGCCCGTGAAGTGAGGAAGGTTCTCGGATCG	403
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Sbjct 404	TAAAAACTCTGTTGTCAGGGAAAGAACCGCCGGATGACCTCCGGTCTGACGGTACCTGAC	463
Query 423	GAGAAAGCCCGGCTAACTACGTGCCAGCACGCCGGTAATACGTAGGGGCAAGCGTTG	482
Sbjct 464	GAGAAAGCCCGGCTAACTACGTGCCAGCACGCCGGTAATACGTAGGGGCAAGCGTTG	523
Query 483	TCCGGAAATTATGGCGTAAAGCGCGCGCAGCGGGCTTCTTAAGTCAGGTGTGAAAGCCC	542
Sbjct 524	TCCGGAAATTATGGCGTAAAGCGCGCGCAGCGGGCTTCTTAAGTCAGGTGTGAAAGCCC	583
Query 543	ACGGCTCAACCGTGGAGGGCCATCTGAAACTGGGAGCTTGAAGTCAGGATAGGAGAGCG	602
Sbjct 584	ACGGCTCAACCGTGGAGGGCCATCTGAAACTGGGAGCTTGAAGTCAGGAGGGAGAGCG	643
Query 603	GAATTCCACGTGTAAGGGTGAAATGCGTAAAGATGTTGGAGGAAACCCAGTGGCGAAAGCG	662
Sbjct 644	GAATTCCACGTGTAAGGGTGAAATGCGTAAAGATGTTGGAGGAAACCCAGTGGCGAAAGCG	703
Query 663	GCTCTCTGGCCTGTAACTGACGCTGAGGCCCGAAAAGCGTG	702
Sbjct 704	GCTCTCTGGCCTGTAACTGACGCTGAGGCCCGAAAAGCGTG	743

>Aneurinibacillus thermoerophilus strain L420-91 16S ribosomal RNA, partial sequence

Sequence ID: NR\_029303.1 Length: 1485

Range 1: 51 to 747

Score:1229 bits(665), Expect:0.0,

Identities:689/700(98%), Gaps:4/700(0%), Strand: Plus/Plus

Query	4	GGA-TGCTTGGCCTTCCTGAGGTTAGCGGCAGACGGGTGAGTAACACGTAGCGAACCTGC	62
Sbjct	51	GGAGTGCTT-GCATTCCTGAGGTTAGCGGCAGACGGGTGAGTAACACGTAGCGAACCTGC	109
Query	63	CTGTACGACCGGGATAACTCCGGAAAACCGGAGCTAATACCGGATAGGATGCCAACCGC	122
Sbjct	110	CTGTACGACCGGGATAACTCCGGAAAACCGGAGCTAATACCGGATAGGATGCCAACCGC	169
Query	123	ATGGTTCCGCAATGGAAAGGGCTTTGAGCCGCGTACAGATGGCCTGCCGCATTAGCTG	182
Sbjct	170	ATGGTTCCGCAATGGAAAGGGCTTTGAGCCGCGTACAGATGGCCTGCCGCATTAGCTA	229
Query	183	GTTGGTGGGTAACGGCCTACCAAGGGGACGATGCGTAGCCGACCTGAGAGGGTGATAAG	242
Sbjct	230	GTTGGTGGGTAACGGCCTACCAAGGGGACGATGCGTAGCCGACCTGAGAGGGTGAG-ACG	288
Query	243	GCCACACTGGGACTGAGACACGGCTCCAGACTCCTACGGGAGGCAGCAGTAGGGAAATCTT	302
Sbjct	289	GCCACACTGGGACTGAGACACGTC-CCAGACTCCTACGGGAGGCAGCAGTAGGGAAATCTT	347
Query	303	CCGCAATGGACGAAAGTCTGACGGAGCAACGGCGCGTGAAGTGAGGAAGGTCTCGGATCG	362
Sbjct	348	CCGCAATGGACGAAAGTCTGACGGAGCAACGGCGCGTGAAGTGAGGAAGGTCTCGGATCG	407
Query	363	TAAAACTCTGTTGTCAGGGAAAGAACCGGCCGGATGACCTCCGGCTGACGGTACCTGAC	422
Sbjct	408	TAAAACTCTGTTGTCAGGGAAAGAACCGGCCGGATGACCTCCGGCTGACGGTACCTGAC	467
Query	423	GAGAAAGCCCCGGCTAACTACGTGCCAGCACGGCGCGTAATAACGTAGGGGGCAAGCGTTG	482
Sbjct	468	GAGAAAGCCCCGGCTAACTACGTGCCAGCACGGCGCGTAATAACGTAGGGGGCAAGCGTTG	527
Query	483	TCCGGATTATTGGCGTAAAGCGCGCGCAGGGGGCTTCTTAAGTCAGGTGTGAAAGCCC	542
Sbjct	528	TCCGGATTATTGGCGTAAAGCGCGCGCAGGGGGCTTCTTAAGTCAGGTGTGAAAGCCC	587
Query	543	ACGGCTAACCGTGGAGGGCCATCTGAAACTGGGGAGCTTGAGTCAGGATAGGAGGCG	602
Sbjct	588	ACGGCTAACCGTGGAGGGCCATCTGAAACTGGGGAGCTTGAGTCAGGAGGGAGGCG	647
Query	603	GAATTCCACGTGAGCGGTGAAATCGTAAAGATGTGGAGGAACACCGTGGCGAAGGGCG	662
Sbjct	648	GAATTCCACGTGAGCGGTGAAATCGTAAAGATGTGGAGGAACACCGTGGCGAAGGGCG	707
Query	663	GCTCTCTGGCCTGTAACGTGACGCTGAGGGCGCAAAGCGTG	702
Sbjct	708	GCTCTCTGGCCTGTAACGTGACGCTGAGGGCGCAAAGCGTG	747

>Aneurinibacillus sediminis strain 1-10M-8-7-50 16S ribosomal RNA, partial sequence

Sequence ID: NR\_157740.1 Length: 1414

Range 1: 25 to 716

Score:1099 bits(595), Expect:0.0,

Identities:663/696(95%), Gaps:4/696(0%), Strand: Plus/Plus

Query	7	TGCTTGGCCTTCCTGAGGTTAGCGCGGACGGGTGAGTAAACACGTAGGCACCTGCCGT	66
Sbjct	25	TGCTTGCACCTT-CGGAGGTTAGCGCGGACGGGTGAGTAAACACGTAGGCACCTGCCGT	83
Query	67	ACGACCGGGATAACTCCGGAAAACCGGAGCTAATACCGGATAAGGATGCCAACCGCATGG	126
Sbjct	84	ACGACCGGGATAACTCCGGAAAACCGGAGCTAATACCGGATAACGATGCCAACCGCATGG	143
Query	127	TTCGGCATGGAAAGGGCTTGAAGCCCGTACAGATGGGCTGCCGCATTAGCTGGTTG	186
Sbjct	144	TTTGCATGGAAA-GACCATGAGTCACGTACAGATGGGCTGCCGCATTAGCTAGTTG	202
Query	187	GTGGGGTAACCGGCCTACCAAGGGGACGATGCGTAGCCGACCTGAGAGGGTGATACGGCCA	246
Sbjct	203	GTAGGGTAACCGGCCTACCAAGGGGACGATGCGTAGCCGACCTGAGAGGGTGAT-CGGCCA	261
Query	247	CACTGGACTGAGACACGGCTCCAGACTCTACGGGAGGCAGCAGTAGGGAAATCTTCCGC	306
Sbjct	262	CACTGGACTGAGACACGGC-CCAGACTCTACGGGAGGCAGCAGTAGGGAAATCTTCCGC	320
Query	307	AATGGACGAAAGTCTGACGGAGCAACGCCCGGTGAGTGAAGGAAGGTCTTCGGATCGTAAA	366
Sbjct	321	AATGGACGAAAGTCTGACGGAGCAACGCCCGGTGAAACGATGAAGGTTTCGGATCGTAAA	380
Query	367	ACTCTGTTGTCAGGGAAAGAACGCCGGGATGACCTCCGGTCTGACGGTACCTGACGAGA	426
Sbjct	381	GTTCTGTTGTTAGGGAAAGAACGCCGGGATGACCTCCGGTCTGACGGTACCTAACGAGA	440
Query	427	AAGCCCCGGCTAAACTACGTGCCAGCAGCCGGTAATACGTAGGGGGCAAAGCGTTGTCCG	486
Sbjct	441	AAGCCCCGGCTAAACTACGTGCCAGCAGCCGGTAATACGTAGGGGGCAAAGCGTTGTCCG	500
Query	487	GAATTATTGGCGTAAAGCGCGCGCAAGCGCGCTCTTAAGTCAGGTGTGAAAGCCCACGG	546
Sbjct	501	GAATTATTGGCGTAAAGCGCGCGCAAGCGCGCTCTTAAGTCAGGTGTGAAAGCCCACGG	560
Query	547	CTCAACCGTGGAGGGCCATCTGAAAATGGGAGCTTGAGTGCAGGATAGGGAGAGCGGAAT	606
Sbjct	561	CTCAACCGTGGAGGGCCATCTGAAAATGGGAGCTTGAGTGCAGGAGAGGGAGAGCGGAAT	620
Query	607	TCCACGTGTAGCGGTGAAATGCGTAAAGATGTGGAGGAACACCCAGTGCGAACGGCGCTC	666
Sbjct	621	TCCACGTGTAGCGGTGAAATGCGTAAAGATGTGGAGGAACACCCAGTGCGAACGGCGCTC	680
Query	667	TCTGGCCTGTAACGTGACGCTGAGGCACGAAAGCGTG	702
Sbjct	681	TCTGGCCTGTAACGTGACGCTGAGGCACGAAAGCGTG	716

>Aneurinibacillus danicus strain DB4 16S ribosomal RNA, partial sequence  
 Sequence ID: NR\_028657.1 Length: 1417  
 Range 1: 60 to 742  
 Score:1070 bits(579), Expect:0.0,  
 Identities:651/686(95%), Gaps:3/686(0%), Strand: Plus/Plus

Query	17	TCCTGAGGTTAGCGGCGGA CGGGT GAGTAAACACGTAGGCACCTGCCGTGACGACCGGGAA	76
Sbjct	68	TCCTGCGGTCA CGGGCGGA CGGGT GAGTAAACACGTAGGCACCTGCCGTGACGACCGGGAA	119
Query	77	TAACTCCGGAAACCGGAGCTAATAACCGGATAGGATGCCAACCGCATGGTTCGGCATGG	136
Sbjct	120	TAACTCCGGAAACCGGAGCTAATAACCGGATAGGATTTCAAGACCGCATGGTTGGAAATGG	179
Query	137	AAAGGGCTTGAGCCCGTACAGATGGGCCTGCCGCATTAGCTGGTGGTGGGTAAC	196
Sbjct	180	AAA-GACCCCTGTGTCACGTACAGATGGGCCTGCCGCATTAGCTAGTTGGTGGGTAAC	238
Query	197	GCGCTACCAAAGCGACCGATGCGTAGCCGACCTGAGAGGGGTGATACGGCCACACTGGGACT	256
Sbjct	239	GCGCTACCAAAGCGACCGATGCGTAGCCGACCTGAGAGGGGTGA-ACGGCCACACTGGGACT	297
Query	257	GAGACACGGCTCCAGACTCCTACGGGAGGCAGCAGTAGGGAAATCTCCGCAATGGACGAA	316
Sbjct	298	GAGACACGGC-CCAGACTCCTACGGGAGGCAGCAGTAGGGAAATCTCCGCAATGGACGAA	356
Query	317	AGTCTGACGGAGCAACGCCGCGTGAGTGAAGGAAGGTTCTCGATCGTAAACCTCTGTTGT	376
Sbjct	357	AGTCTGACGGAGCAACGCCGCGTGAAACGAAGAAGGTTCTCGATCGTAAAGTTCTGTTGT	416
Query	377	CAGGGAAAGAACCGCCGGGATGACCTCCCGGTCTGACGGTACCTGACGAGAAAGCCCCGGC	436
Sbjct	417	TAGGGAAAGAACCGCCGGGATAACCTCCCGGTCTGACGGTACCTAACGAGAAAGCCCCGGC	476
Query	437	TAACTACGTGCCAGCAAGCGCGGTTAAACGTAGGGGCAAGCGTTGTCCGGAAATTATTGG	496
Sbjct	477	TAACTACGTGCCAGCAAGCGCGGTTAAACGTAGGGGCAAGCGTTGTCCGGAAATTATTGG	536
Query	497	GCGTAAAGCGCGCGCAAGCGGGTTCTTAAGTCAGGTGAAAGGCCAACGGCTCAACCGTG	556
Sbjct	537	GCGTAAAGCGCGCGCAAGCGGGTTCTTAAGTCAGGTGAAAGGCCAACGGCTCAACCGTG	596
Query	557	GAGGGCCATCTGAAACTGGGGAGCTTGAGTGCAAGGATAGGGAGAGCGGAATTCCACGTGTA	616
Sbjct	597	GAGGGCCACTTGAAGACTGGGAGGGCTTGAGTGCAAGGAGAGGGAGAGCGGAATTCCACGTGTA	656
Query	617	GCGGTGAAATGCGTAAAGATGTGGAGGAACACCCAGTGCGGAAGGCCGGCTCTGGCCGT	676
Sbjct	657	GCGGTGAAATGCGTAGAGATGTGGAGGAACACCTGTGGCGGAAGGCCGGCTCTGGCCGT	716
Query	677	AACTGACGCTGAGGGCGGAAAAGCGTG	782
Sbjct	717	AACTGACGCTGAGGGCGGAAAAGCGTG	742

>Aneurinibacillus danicus strain NBRC 102444 16S ribosomal RNA, partial sequence

Sequence ID: NR\_114088.1 Length: 1501

Range 1: 62 to 744

Score:1066 bits(577), Expect:0.0,

Identities:650/686(95%), Gaps:3/686(0%), Strand: Plus/Plus

Query	17	TCCTGAGGTTAGCGGCAGGGTAGGTAAACACGTAGGGCACCTGCCGTACGACCGGGAA	76
Sbjct	62	TCCTGCGGTCAAGCGGCAGGGTAGGTAAACACGTAGGGCACCTGCCGTACGACCGGGAA	121
Query	77	TAACTCCGGAAACCGGAGCTAATACCGGATAGGATGCCGAACCGCATGGTCGGCATGG	136
Sbjct	122	TAACTCCGGAAACCGGAGCTAATACCGGATAGGATTCAGACCCCATGGTTGGAAATGG	181
Query	137	AAAAGGGCTTGAGCCCGTACAGATGGGCTGCCGCGATTAGCTGGTTGGGGTAAC	196
Sbjct	182	AAA-GACCCCTGTGTCAGGTACAGATGGGCTGCCGCGATTAGCTAGTTGGTGGGGTAAC	240
Query	197	GGCCTACCAAGGGGACGGATGCCGTAGCCGACCTGAGAGGGGTGATAACGGCCACACTGGGACT	256
Sbjct	241	GGCCTACCAAGGGGACGGATGCCGTAGCCGACCTGAGAGGGGTGAA-ACGGCCACACTGGGACT	299
Query	257	GAGACACGGCTCCAGACTCCTACGGGAGGGCAGCAGTAGGGAAATCTCCGCAATGGACGAA	316
Sbjct	300	GAGACACGGC-CCAGACTCCTACGGGAGGGCAGCAGTAGGGAAATCTCCGCAATGGACGAA	358
Query	317	AGTCTGACGGGAGCAACGCCGCGTGAGTAGGGAGGTCTTCGGATCGTAAAAACTCTGTTGT	376
Sbjct	359	AGTCTGACGGGAGCAACGCCGCGTGAAACGAAAGAAGGTTTCGGATCGTAAAAGTTCTGTTGT	418
Query	377	CAGGGAAAGAACGCCGGGATGACCTCCCGGCTGACGGTACCTGACGAGAAAAGCCCCGGC	436
Sbjct	419	TAGGGAAAGAACGCCGGGATAACCTCCCGGCTGACGGTACCTAACGAGAAAAGCCCCGGC	478
Query	437	TAACTACGTGCCAGCAAGCGCGGTAATACGTAGGGGGCAAGCGTTGTCCGGATTATTGG	496
Sbjct	479	TAACTACGTGCCAGCAAGCGCGGTAATACGTAGGGGGCAAGCGTTGTCCGGATTATTGG	538
Query	497	GCGTAAAGCGCGCGCAAGCGCGCTTCTTAAGTCAGGTGTGAAAGCCCCACGGCTCAACCGTG	556
Sbjct	539	GCGTAAAGCGCGCGCAAGCGCGCTTCTTAAGTCAGGTGTGAAAGCCCCACGGCTCAACCGTG	598
Query	557	GAGGGCCATCTGAAACTGGGGAGCTTGAGTGCAGGATAAGGAGAGGGAAATTCCACGTGTA	616
Sbjct	599	GAGGGCCACTTGAAACTGGGGAGCTTGAGTGCAGGAGAGGGAGAGNGGAATTCCACGTGTA	658
Query	617	GCGGTGAAATGCGTAAAGATGTGGAGGAACACCAAGTGGCGAAAGGGGGCTCTCTGGCCTGT	676
Sbjct	659	GCGGTGAAATGCGTAAAGATGTGGAGGAACACCAAGTGGCGAAAGGGGGCTCTCTGGCCTGT	718
Query	677	AACTGACGCTGAGGCAGGAAAGCGTG	702
Sbjct	719	AACTGACGCTGAGGCAGGAAAGCGTG	744

>Aneurinibacillus humi strain U33 16S ribosomal RNA, partial sequence  
 Sequence ID: NR\_169383.1 Length: 1451  
 Range 1: 54 to 745  
 Score:1066 bits(577), Expect:0.0,  
 Identities:657/696(94%), Gaps:4/696(0%), Strand: Plus/Plus

Query 7	TGCTTGGCCTTCCTGAGGTAGCGCGGACGGGTGAGTAACACGTAGGCAGACCTGCCGT	66
Sbjct 54	TGCTTGCACCT-CGGCGGTTAGCGCGGACGGGTGAGTAACACGTAGGCAGACCTGCCGT	112
Query 67	ACGACCGGGATAACTCGGGAAACCGGAGCTAACACCGGATAGGAAGCCGAAACCGCATGG	126
Sbjct 113	ACGACTGGGATAACTTCGGGGAAACCGAAGCTAACACCGGATAGGTTAACGAAACCGCATGG	172
Query 127	TTCGGCATGGAAAGGGCTTGAGCGCGTACAGATGGGCTGCGCGCATTAGCTGGTTG	186
Sbjct 173	TTCGTTAAAGAAA-GACCTTGCGTCACGTACAGATGGGCTGCGCGCATTAGCTAGTTG	231
Query 187	GTGGGGTAACGGCCTACCAAGCGACGATGCGTAGCCGACCTGAGAGGGTGTACGGCCA	246
Sbjct 232	GTAGGGTAACGGCCTACCAAGCGACGATGCGTAGCCGACCTGAGAGGGTGTACGGCCA	290
Query 247	CACTGGGACTGAGACACGGCTCCAGACTCTACGGGAGGCGACGAGTAGGGAAATCTCCGC	306
Sbjct 291	CACTGGGACTGAGACACGGC-CCAGACTCTACGGGAGGCGACGAGTAGGGAAATCTCCGC	349
Query 307	AATGGACGAAAGTCTGACGGAGCAACGCCCGTGAAGTGAAGGAAGGTCTCGGATCGTAAA	366
Sbjct 350	AATGGACGAAAGTCTGACGGAGCAACGCCCGTGAACGATGAAGGTTTCGGATCGTAAA	409
Query 367	ACTCTGTTGCAGGGAAGAACCGCGGGATGACCTCCCGCTCTGACGGTACCTGACGAGA	426
Sbjct 410	GTTCTGTTGTAAGGGAAAGAACCGCGGGATAACCTCCCGCTCTGACGGTACCTAACGAGA	469
Query 427	AAGCCCCGGCTAACTACGTGCCAGCAGCCCGGTAATACGTAGGGGGCAAGCGTTGTCG	486
Sbjct 470	AAGCCCCGGCTAACTACGTGCCAGCAGCCCGGTAATACGTAGGGGGCAAGCGTTGTCG	529
Query 487	GAATTATTGGCGTAAAGCGCGCGCAGGCGGCTCTTAAGTCAGGTGTGAAGGCCACGG	546
Sbjct 530	GAATTATTGGCGTAAAGCGCGCGCAGGCGGCTCTTAAGTCAGGTGTGAAGGCCACGG	589
Query 547	CTCAACCGTGGAGGGCCATCTGAAACTGGGGAGCTTGAGTGCAGGAAGGGAGGGAAAT	606
Sbjct 590	CTCAACCGTGGAGGGCCATCTGAAACTGGGAAGCTTGAGTGCAGGAAGGGAGGGAAAT	649
Query 607	TCCACGTGTAGCGGTGAAATGCGTAAAGATGTGGAGGAACACCAAGTGGCGAAGGCCGCTC	666
Sbjct 650	TCCACGTGTAGCGGTGAAATGCGTAAAGATGTGGAGGAACACCCGTGGCGAAGGCCGCTC	709
Query 667	TCTGGCCTGTAACGTACCGCTGAGGCCGAAAGCGTG	702
Sbjct 710	TCTGGCCTGTAACGTACCGCTGAGGCCGAAAGCGTG	745

>Aneurinibacillus aneurinilyticus ATCC 12856 strain DSM 5562 16S ribosomal RNA, partial sequence

Sequence ID: NR\_112215.1 Length: 1489

Range 1: 66 to 743

Score:1050 bits(568), Expect:0.0,

Identities:645/682(95%), Gaps:6/682(0%), Strand: Plus/Plus

Query	23	GTTAGCGGGGAGCGGGTGAGTAAACACGTAGCGACCTGCCTGTACGACCGGGATAACTC	82
Sbjct	66	GTTAGCGGGGAGCGGGTGAGTAAACACGTAGCGACCTGCCTGTACGACCGGGATAACTC	125
Query	83	CGGGAAACCGGGAGCTAATACCGGATA-GGATGCCGAACCGCATGGTCGGCATG-GAAAG	140
Sbjct	126	CGGGAAACCGGGAGCTAATACCGGATAACGTTTCAGACCGCATGG-TCTGAAAGAGAAAAG	184
Query	141	GGCTTGTAGCCCGTAGAGATGGCCTGCGCGCATTAGCTGGTGGTGGGTAACGGCC	200
Sbjct	185	ACCTTG-GTCACGTACAGATGGCCTGCGCGCATTAGCTAGTTGGTGGGTAACGGCC	243
Query	201	TACCAAGCGGAGCGATGCGTAGCCGACCTGAGAGGGTGATACGGCCACACTGGGACTGAGA	260
Sbjct	244	TACCAAGCGGAGCGATGCGTAGCCGACCTGAGAGGGTGAT-CGGCCACACTGGGACTGAGA	302
Query	261	CACGGCTCCAAGACTCCTACGGGAGGCAGCAGTAGGGGAATCTTCGCAATGGACGAAAGTC	320
Sbjct	303	CACGGC-CCAGACTCCTACGGGAGGCAGCAGTAGGGGAATCTTCGCAATGGACGAAAGTC	361
Query	321	TGACGGAGCAACCGCCCGTGAAGTGAGGAAAGGTCTTCGGATCGTAAAGCTCTGGTCAGG	380
Sbjct	362	TGACGGAGCAACCGCCCGTGAACGATGAAGGTTTCGGATCGTAAAGTTCTGGTGTAGG	421
Query	381	GAAGAACCGCCGGATGACCTCCGGCTGACGGTACCTGACGAAGAACCCCCGGCTAAC	440
Sbjct	422	GAAGAACCGCCGGATGACCTCCGGCTGACGGTACCTAACGAGAACCCCCGGCTAAC	481
Query	441	TACGTGCCAGCGAGCCGGTAATACGTAGGGGCAAGCGTTCTGGAAATTATTGGCGT	500
Sbjct	482	TACGTGCCAGCGAGCCGGTAATACGTAGGGGCAAGCGTTCTGGAAATTATTGGCGT	541
Query	501	AAAGCGCGCGCAAGCGGCTTCTTAAGTCAGGTGTAAAGGCCAACGGCTAACCGTGGAGG	560
Sbjct	542	AAAGCGCGCGCAAGCGGCTTCTTAAGTCAGGTGTAAAGGCCAACGGCTAACCGTGGAGG	601
Query	561	GCCATCTGAAACTGGGAGCTTGAGTGCAGGATAGGGAGAGCGGAATTCCACGTGTAGCGG	620
Sbjct	602	GCCACTTGAAACTGGGAGCTTGAGTGCAGGAGAGGGAGAGCGGAATTCCACGTGTAGCGG	661
Query	621	TGAAATCGTAAAGATGTGGAGGAACACCGTAGGGCGAAGGCCTCTCTGGCTGTAACT	680
Sbjct	662	TGAAATCGTAAAGATGTGGAGGAACACCGTAGGGCGAAGGCCTCTCTGGCTGTAACT	721
Query	681	GACGCTGAGGCAGCGAAAGCGTG	702
Sbjct	722	GACGCTGAGGCAGCGAAAGCGTG	743

>Aneurinibacillus aneurinilyticus ATCC 12856 strain DSM 5562 16S ribosomal RNA, partial sequence

Sequence ID: NR\_112203.1 Length: 1489

Range 1: 66 to 743

Score:1050 bits(568), Expect:0.0,

Identities:645/682(95%), Gaps:6/682(0%), Strand: Plus/Plus

Query	23	GTTAGCGGGGAGCGGGTGAGTAACACGTAGGCCAACCTGCCGTACGACCGGGATAACTC	82
Sbjct	66	GTTAGCGGGGAGCGGGTGAGTAACACGTAGGCCAACCTGCCGTACGACCGGGATAACTC	125
Query	83	CGGGAAACCGGGAGCTAAACCGGATA-GGATGCCAACCGCATGGTTCCGCATG-GAAAAG	148
Sbjct	126	CGGGAAACCGGGAGCTAAACCGGATAACCGGATACGTTTTCAGACCGCATGG-TCTGAAAGAGAAAAG	184
Query	141	GGCTTTGAGCCCGTACAGATGGCCTGCCGCCATTAGCTGGTTGGTGGGTAACGCC	200
Sbjct	185	ACCTTTG-GTCACGTACAGATGGCCTGCCGCCATTAGCTAGTTGGTGGGTAACGCC	243
Query	201	TACCAAGGCAGCATGCGTAGCCGACCTGAGAGGGTGATAACGCCACACTGGACTGAGA	260
Sbjct	244	TACCAAGGCAGCATGCGTAGCCGACCTGAGAGGGTGATAACGCCACACTGGACTGAGA	302
Query	261	CACGGCTCCAGACTCCTACGGGAGGCAGCAGTAGGGAACTTCCGCAATGGACGAAAGTC	320
Sbjct	303	CACGGC-CCAGACTCCTACGGGAGGCAGCAGTAGGGAACTTCCGCAATGGACGAAAGTC	361
Query	321	TGACGGAGCAAAGCCCGGTGAGTGAGGAAGGTCTCGGATCGAAAAACTCTGTTGTCAGG	380
Sbjct	362	TGACGGAGCAAAGCCCGGTGAAACGATGAAGGTTTCGGATCGAAAGTTCTGTTGTTAGG	421
Query	381	GAAGAAACGCCGGGATGACCTCCGGCTGACGGTACCTGACGAGAAAGCCCCGGCTAAC	440
Sbjct	422	GAAGAAACGCCGGGATGACCTCCGGCTGACGGTACCTAACGAGAAAGCCCCGGCTAAC	481
Query	441	TACGTGCCAGCACCGCGGTAAACGTAGGGGCAAGCGTTGCCGAAATTATTGGCGT	500
Sbjct	482	TACGTGCCAGCACCGCGGTAAACGTAGGGGCAAGCGTTGCCGAAATTATTGGCGT	541
Query	501	AAAGCGCGCGCAGGCCGGCTTCTTAAGTCAGGTGTGAAAGCCCACGGCTAACCGTGGAGG	560
Sbjct	542	AAAGCGCGCGCAGGCCGGCTTCTTAAGTCAGGTGTGAAAGCCCACGGCTAACCGTGGAGG	601
Query	561	GCCATCTGAAACTGGGGAGCTTGAGTGCAGGATAGGGAGAGCGGAATTCCACGTGTAGCGG	620
Sbjct	602	GCCACTTGAAACTGGGGAGCTTGAGTGCAGGAGAGGGAGAGCGGAATTCCACGTGTAGCGG	661
Query	621	TGAAATGCGTAAAGATGTGGAGGAACACCAAGTGGCGAAAGGCCGCTCTCTGCCGTAACT	680
Sbjct	662	TGAAATGCGTAAAGATGTGGAGGAACACCCGTGGCGAAAGGCCGCTCTCTGCCGTAACT	721
Query	681	GACGCTGAGGCCCGAAAAGCGTG	782
Sbjct	722	GACGCTGAGGCCCGAAAAGCGTG	743

&gt;Aneurinibacillus migulanus strain B0270 16S ribosomal RNA, partial sequence

Sequence ID: NR\_036799.1 Length: 1483

Range 1: 70 to 747

Score:1050 bits(568), Expect:0.0,

Identities:645/682(95%), Gaps:6/682(0%), Strand: Plus/Plus

Query	23	GGTTAGCGCGGACCGGTGAGTAACACGTAAGCGAACCTGCCTGTACGACCGGATAACTC	82
Sbjct	70	GGTTAGCGCGGACCGGTGAGTAACACGTAAGCGAACCTGCCTGTACGACCGGATAACTC	129
Query	83	CGGGAAAACCGGAGCTAATACCGGATA-GGATGCCAACCGCATGGTCGGCATG-GAAAG	148
Sbjct	130	CGGGAAAACCGGAGCTAATACCGGATACTTCTTCAGACCGCATGG-TCTGAAAGAGAAAG	188
Query	141	GGCTTGAGCCCGGTACAGATGGGCTGCGGCGCATTAGCTGGTGGTGGGTAACGGCC	200
Sbjct	189	ACCTTTG-GTCACTGACAGATGGGCTGCGGCGCATTAGCTAGTTGGTGGGTAACGGCC	247
Query	281	TACCAAGGCACGATGCGTAGCCGACCTGAGAGGGTGATAACGGCACACTGGGACTGAGA	260
Sbjct	248	TACCAAGGCACGATGCGTAGCCGACCTGAGAGGGTGATAACGGCACACTGGGACTGAGA	306
Query	261	CACGGCTCCAGACTCCTACCGGAGGCAGCAGTAGGGAACTCTCCGCAATGGACGAAAGTC	320
Sbjct	307	CACGGC-CCAGACTCCTACCGGAGGCAGCAGTAGGGAACTCTCCGCAATGGACGAAAGTC	365
Query	321	TGACGGAGCAACGCCCGTGAAGTGAAGGAGGTCTCGGATCGTAAACTCTGTTGTCAGG	380
Sbjct	366	TGACGGAGCAACGCCCGTGAACGATGAAGGTTTCGGATCGTAAAGTTCTGTTGTTAGG	425
Query	381	GAAGAAACGCCGGGATGACCTCCGGTCTGACGGTACCTGACGAGAAAGCCCCGGCTAAC	440
Sbjct	426	GAAGAAACGCCGGGATGACCTCCGGTCTGACGGTACCTAACGAGAAAGCCCCGGCTAAC	485
Query	441	TACGTGCCAGCGCCGGTAATACGTAGGGGGCAAGCGTTGTCGGATTATTGGCGT	500
Sbjct	486	TACGTGCCAGCGCCGGTAATACGTAGGGGGCAAGCGTTGTCGGATTATTGGCGT	545
Query	501	AAAGCGCGCGCAGGGGGCTTCTTAAGTCAGGTGTGAAAGCCCACGGCTAACCGTGGAGG	560
Sbjct	546	AAAGCGCGCGCAGGGGGCTTCTTAAGTCAGGTGTGAAAGCCCACGGCTAACCGTGGAGG	605
Query	561	GCCATCTGAAACTGGGGAGCTTGAGTCAGGATAGGAGAGGGAAATTCCACGTGTAGCGG	620
Sbjct	606	GCCACTTGAAACTGGGAAGCTTGAGTCAGGAGGGAGGGGAATTCCACGTGTAGCGG	665
Query	621	TGAAATGCGTAAAGATGTGGAGGAACACCGATGGCGAAGGGGGCTCTCTGCGCTGTAACT	680
Sbjct	666	TGAAATGCGTAAAGATGTGGAGGAACACCGATGGCGAAGGGGGCTCTCTGCGCTGTAACT	725
Query	681	GACGCTGAGGGCGCGAAAGCGTG 702	
Sbjct	726	GACGCTGAGGGCGCGAAAGCGTG 747	

>Aneurinibacillus aneurinilyticus strain Murayama 16S ribosomal RNA, partial sequence

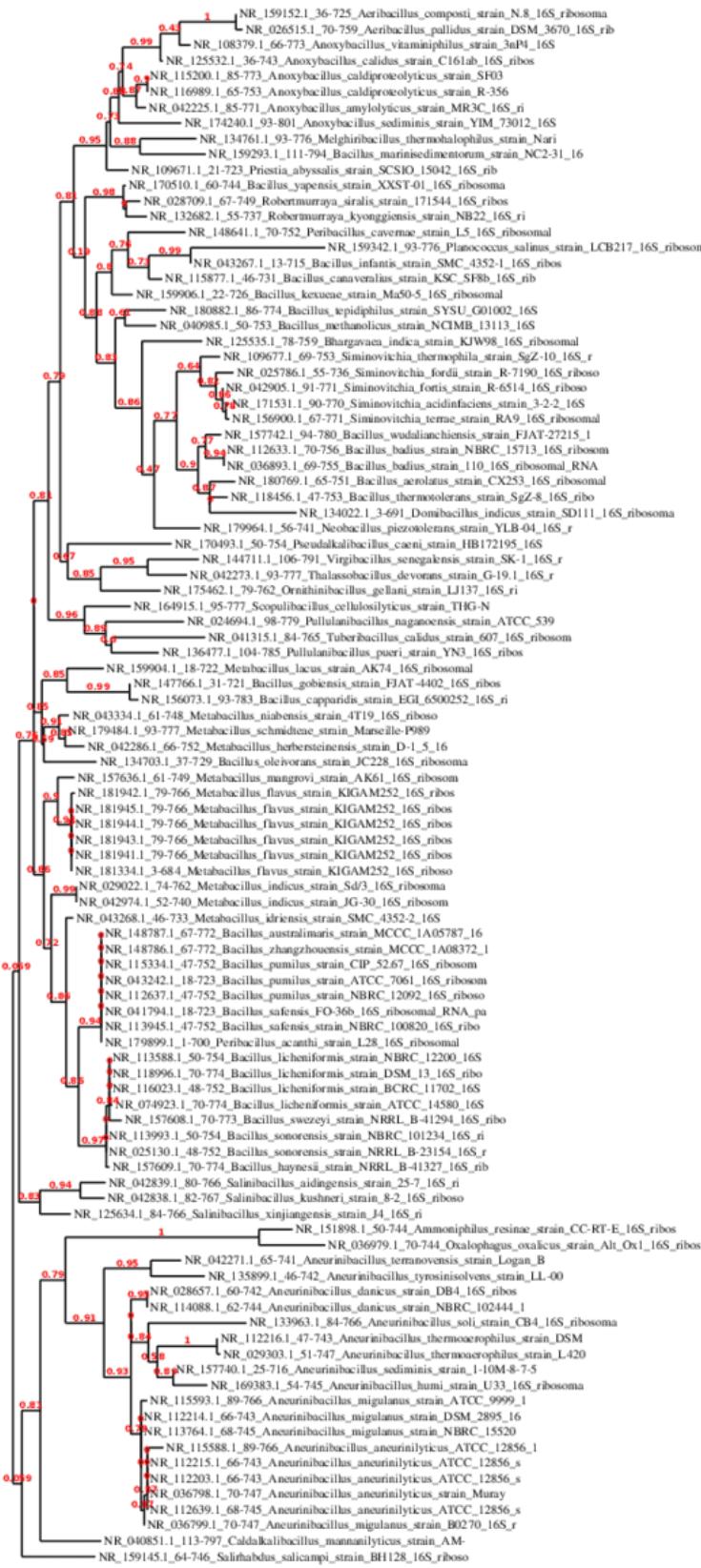
Sequence ID: NR\_036798.1 Length: 1482

Range 1: 70 to 747

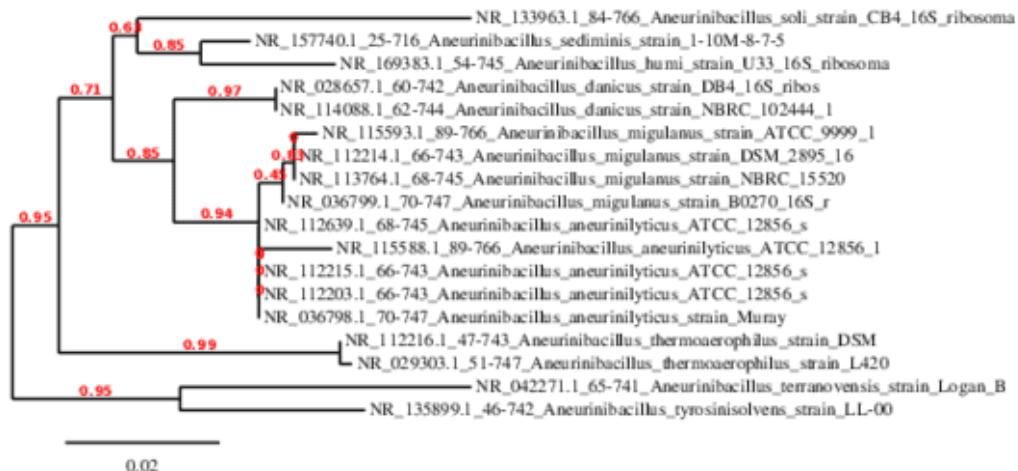
Score:1050 bits(568), Expect:0.0,

Identities:645/682(95%), Gaps:6/682(0%), Strand: Plus/Plus

Query	23	GGTTAGCGGCCGACGGGTGAAGTAAACACGTAAGCGAACCTGCCTGTACGACCCGGATAACTC	82
Sbjct	70	GGTTAGCGGCCGACGGGTGAAGTAAACACGTAAGCGAACCTGCCTGTACGACCCGGATAACTC	129
Query	83	CGGGAAAACCGGAGCTAATACCGGATA-GGATGCCGAACCGCATGGTCGGCATG-GAAAG	148
Sbjct	130	CGGGAAAACCGGAGCTAATACCGGATAACGTTTCAGACCGCATGG-TCTGAAAGAGAAAAG	188
Query	141	GGCTTTGAGCCCGTACAGATGGGCCTGCGCGCATTAGCTGGTTGGTGGGTAACGGCC	200
Sbjct	189	ACCTTTG-GTCACGTACAGATGGGCCTGCGCGCATTAGCTAGTTGGTGGGTAACGGCC	247
Query	201	TACCAAGGCACGATGCGTAAGCGAACCTGAAGAGGGTGATAACGCCACACTGGGACTGAGA	260
Sbjct	248	TACCAAGGCACGATGCGTAAGCGAACCTGAAGAGGGTGATAACGCCACACTGGGACTGAGA	306
Query	261	CACGGCTCCAGACTCCTACGGGAGGCAGCAGTAGGGGAAATCTTCCGCAATGGACGAAAGTC	320
Sbjct	307	CACGGC-CCAGACTCCTACGGGAGGCAGCAGTAGGGGAAATCTTCCGCAATGGACGAAAGTC	365
Query	321	TGACGGAGCAACGCCCGCGTGAAGTAGGGAAAGGTCTTCGGATCGTAAACTCTGTTGTCAGG	380
Sbjct	366	TGACGGAGCAACGCCCGCGTGAACGATGAAGGTTTCGGATCGTAAAGTTCTGTTGTCAGG	425
Query	381	GAAGAAACGCCGGGATGACCTCCCGGCTGACCGTACCTGACCGAAAAGCCCGGCTAAC	440
Sbjct	426	GAAGAAACGCCGGGATGACCTCCCGGCTGACCGTACCTAACGAGAAAAGCCCGGCTAAC	485
Query	441	TACGTGCCAGCAGCCCGCGTAATAACGTAGGGGGCAAGCGTTGTCGGAAATTATTGGCGT	500
Sbjct	486	TACGTGCCAGCAGCCCGCGTAATAACGTAGGGGGCAAGCGTTGTCGGAAATTATTGGCGT	545
Query	501	AAAGCGCGCGCAGCGCGCTTCTTAAGTCAGGTGTGAAGAGCCCACGGCTAACCGTGGAGG	560
Sbjct	546	AAAGCGCGCGCAGCGCGCTTCTTAAGTCAGGTGTGAAGAGCCCACGGCTAACCGTGGAGG	605
Query	561	GCCATCTGAAACTGGGGAGCTTGAGTCAGGATAAGGAGAGCGGAATTCCACCGTGTAGCGG	620
Sbjct	606	GCCATCTGAAACTGGGGAGCTTGAGTCAGGATAAGGAGAGCGGAATTCCACCGTGTAGCGG	665
Query	621	TGAAAATGCGTAAAGATGTGGAGGAAACACCAAGTGGCGAAGGGCGCTCTCTGGCTGTAACT	680
Sbjct	666	TGAAAATGCGTAAAGATGTGGAGGAAACACCCGTGGCGAAGGGCGCTCTCTGGCTGTAACT	725
Query	681	GACGCTGAGGGCGCGAAAGCGTG 782	
Sbjct	726	GACGCTGAGGGCGCGAAAGCGTG 747	



## Pohon filogenetik kekerabatan bakteri hasil seq 16s RNA



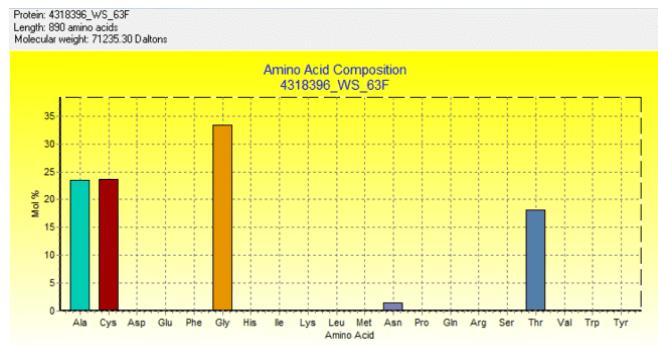
## Protein Penyusun

Protein: Isolt Baktei K.W.R-4.1

Length = 890 amino acids

Molecular Weight = 71235.30 Daltons

Amino Acid	Number	Mol%
Ala	A	209
Cys	C	210
Asp	D	0
Glu	E	0
Phe	F	0
Gly	G	297
His	H	0
Ile	I	0
Lys	K	0
Leu	L	0
Met	M	0
Asn	N	13
Pro	P	0
Gln	Q	0
Arg	R	0
Ser	S	0
Thr	T	161
Val	V	0
Trp	W	0
Tyr	Y	0



**Nucleotide Composition**

DNA molecule: 4318396\_WS\_63F

Length = 890 base pairs

Molecular Weight = 269976.00 Daltons, single stranded

Molecular Weight = 542232.00 Daltons, double stranded

G+C content = 56.97%

A+T content = 41.57%

Nucleotide	Number	Mol%
A	209	23.48
C	210	23.60
G	297	33.37
T	161	18.09
N	13	1.46

**Lampiran 6.** Dokumentasi penelitian

Proses pengambilan sampel dan penentuan kondisi lingkungan sampel



Proses pengambilan sampel



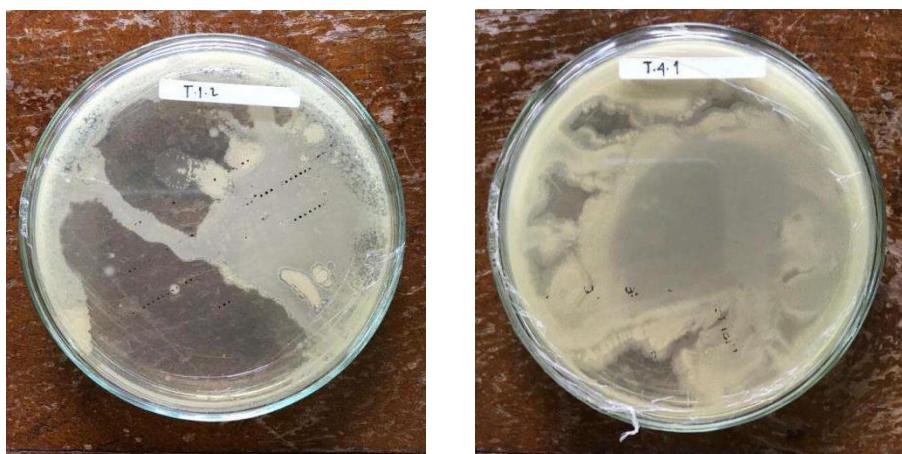
Pengukuran pH



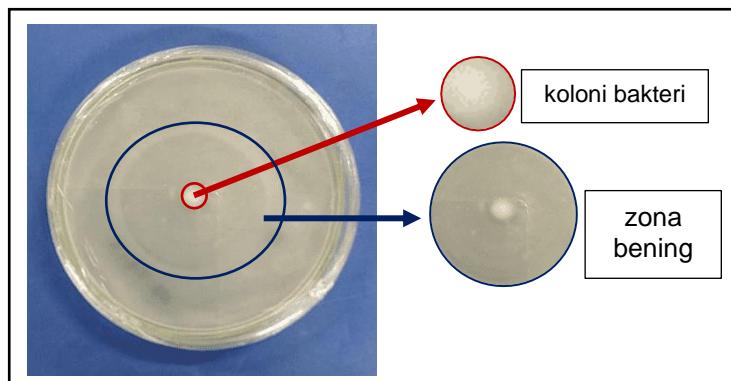
Pengukuran suhu



Proses pembuatan media



Isolasi bakteri



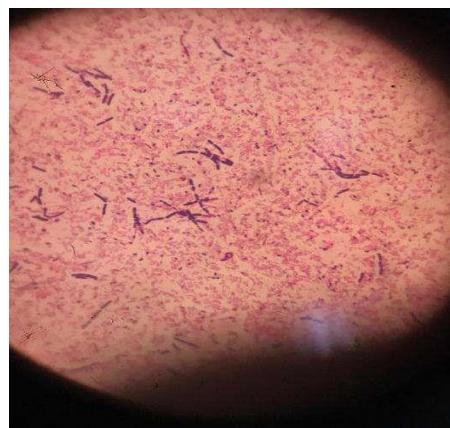
Hasil seleksi bakteri termofilik penghasil kitinase



Penyiapan produksi enzim kitinase (media selektif)



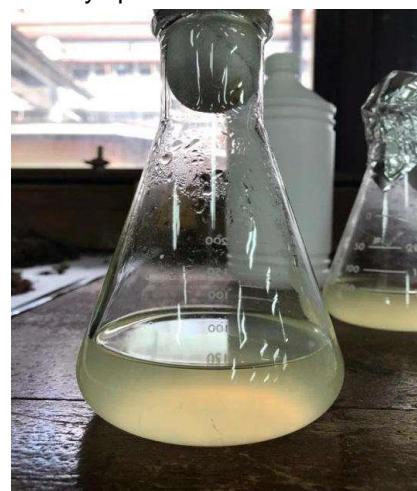
Uji biokimia isolate bakteri termofilik K.W.R 4-1



Pengamatan secara makroskopis



Penyiapan media inoculum



Inokulum aktif



Standar N-asetil glukosamin



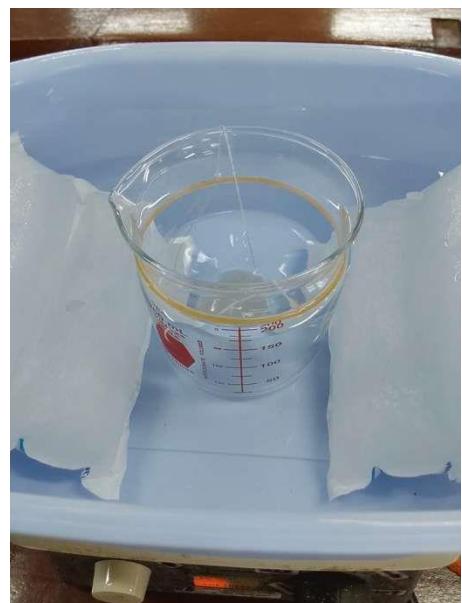
## Pengukuran aktivitas kitinase



## Pengukuran kadar protein



#### Karakterisasi enzim



Tahapan dialisis

