

DAFTAR PUSTAKA

- Afandhi, A., Choliq, F. A., Anggrilika, H. W. S., & Tarno, H. (2018). Distribution of the Endophytic Fungi in Apple Leaves. *Agrivita*, 40(1), 91–100.
- Akmalasari, I., Purwati, E. S., & Dewi, R. S. (2013). Isolasi dan identifikasi jamur endofit tanaman manggis (*Garcinia mangostana L.*). *Biosfera*, 30(2), 82–89.
- Alviolita, Z., Oramahi, H. A., & Diba, F. (2005). *Pengendali Jamur Penyebab Busuk Benih Tusam (Pinus merkusii Jungh et de Vriese) dengan Asap Cair Kayu Laban (Vitex pubescens Vahl)*. 263–268.
- Andriani, D., & Heriansyah, P. (2021). Identifikasi Jamur Kontaminan pada Berbagai Eksplan Kultur Jaringan Anggrek Alam (*Bromheadia finlaysoniana* (Lind.) Miq.). *Agro Bali : Agricultural Journal*, 4(2), 192–199.
<https://doi.org/10.37637/ab.v4i2.723>
- Andriani, S., Aini, F., & Ihsan, M. (2019). Isolasi dan Identifikasi Jamur Patogen pada Tanaman Nanas Ananas comosus (L). Merr. var. Tangkit. *Jurnal Bio-Site*, 4(1), 13–20. <https://doi.org/10.22437/bs.v5i01.6579>
- Ariani, E., & Rifin, A. (2017). Analisis Usahatani Kakao Pada Dua Pola Tanam Polikultur. *Forum Agribisnis*, 7(2), 173–190.
<https://doi.org/10.29244/fagb.7.2.173-190>
- Astuti, cindy cahyaning. (2017). Analisis Korelasi untuk Mengetahui Keeratan Hubungan antara Keaktifan Mahasiswa dengan Hasil Belajar Akhirn. *Journal of Information and Computer Technology Education*, 1(1), 1–7.
- Baderan, D. W. K., Rahim, S., Angio, M., & Salim, A. I. Bin. (2021). Keanekaragaman, Kemerataan, Dan Kekayaan Spesies Tumbuhan Dari Geosite Potensial Benteng Otanaha Sebagai Rintisan Pengembangan

- Geopark Provinsi Gorontalo. *AL-KAUNIYAH: Jurnal Biologi*, 14(2), 264–274.
<http://dx.doi.org/10.15408/kauniyah.v14i2.16746>
- Benjamin, C. R., Haynes, W. C., & Hassalline, C. \v. (1964). Micro-Organisms. *Agricultural Research Service*, 955.
<https://naldc.nal.usda.gov/download/32293/PDF>
- Bierza, W., Woźniak, G., Kompała-Bąba, A., Magurno, F., Malicka, M., Chmura, D., Błońska, A., Jagodziński, A. M., & Piotrowska-Seget, Z. (2023). The Effect of Plant Diversity and Soil Properties on Soil Microbial Biomass and Activity in a Novel Ecosystem. *Sustainability*, 15(6), 4880.
<https://doi.org/10.3390/su15064880>
- Botek, M., Jabuddin, L. O., Purwanti, R. E., Syarni, P., Mallarangeng, R., & Syair. (2020). Pengaruh Paecilomyces sp. Pada Berbagai Bahan Organik Terhadap Ketahanan dan Produksi Padi Gogo. *Agercolere*, 2(2), 30–36.
<https://doi.org/10.37195/jac.v2i2.91>
- Damanik, S. (2012). Pengembangan Karet Alam (*Hevea brasiliensis*) Berkelanjutan Di Indonesia. *Perspektif*, 11(1), 79–90.
- Gusnawati, H., Taufik, M., Triana, L., & Asniah. (2014). Karakteristik Morfologi Trichoderma spp . Indigenus Sulawesi Tenggara. *Agroteknos*, 4(2), 88–94.
- Herlina, L. (2013). Uji Potensi Gliocladium sp Terhadap Pertumbuhan dan Produksi Tanaman Tomat. *Biosaintifika*, 5(2), 88–93.
- Hilarino, M. P. A., e Silveira, F. A. de O., Oki, Y., Rodrigues, L., Santos, J. C., Corrêa, A., Fernandes, G. W., & Rosa, C. A. (2011). Distribuição da comunidade de fungos endofíticos em folhas de *Bauhinia brevipes* (Fabaceae). *Acta Botanica Brasilica*, 25(4), 815–821.

<https://doi.org/10.1590/S0102-33062011000400008>

Hutapea, S., Siregar, T. H., & Astuti, R. (2017). Iklim dan Perkebunan Karet : Suatu Tinjauan Dalam Kaitannya pada Budidaya Tumpang Sari. *Drat Monograph*, 1–18.

Ismail, M., & Supijatno. (2016). Tapping of Rubber (*Hevea brasiliensis* Muell Arg.) at Sumber Tengah Resources, Jember, East Java. *Agrohorti*, 4(3), 257–265.

Izzati, I., Lubis, L., & Hasanuddin. (2019). Eksplorasi Cendawan Endofit pada Akar Tanaman Karet (*Hevea brasiliensis* Muell.Arg.) sebagai Agens Hayati Jamur Akar Putih (*Rigidoporus microporus* (Swartz; Fr)) di Kabupaten Asahan. *Agroekoteknologi FP USU*, 7(2), 347–355.

Janudianto, Prahmono, A., Napitupulu, H., & Rahayu, S. (2013). Panduan Budidaya Karet untuk Petani Skala Kecil. *World Agroforestry Centre (ICRAF) Southeast Asia Regional Program*, 1–16.

Kartika, T., Suciatmih, Tarmadi, D., Guswenriyo, I., Prianto, A. H., & Yusuf, S. (2006). Rayap Tanah *Coptotermes* sp . Pathogenic Ability of *Cunninghamella* sp . against Subterranean Termites *Coptotermes* sp . *Tropical Wood Science and Technology*, 4(1), 24–27.

Kasongat, H., Gafur, M. A., & Ponisri, P. (2019). Identifikasi Dan Keanekaragaman Jenis Jamur Ektomikoriza Pada Hutan Jati Di Seram Bagian Timur. *Median : Jurnal Ilmu Ilmu Eksakta*, 11(1), 39–46.

<https://doi.org/10.33506/md.v11i1.461>

Meyer, K. M., & Leveau, J. H. J. (2012). Microbiology of the phyllosphere: A playground for testing ecological concepts. *Oecologia*, 168(3), 621–629.

<https://doi.org/10.1007/s00442-011-2138-2>

Mukrimin, Musdalifah, N., Larekeng, S. H., Sultan, & Christita, M. (2021a). Fungal diversity inhabiting tissues of ebony (*Diospyros celebica* Bakh .) in urban forest. *2nd Biennial Conference of Tropical Biodiversity*, 0–16.

<https://doi.org/10.1088/1755-1315/886/1/012031>

Mukrimin, Musdalifah, N., Larekeng, S. H., Sultan, & Christita, M. (2021b). Fungal diversity inhabiting tissues of ebony (*Diospyros celebica* Bakh .) in urban forest Fungal diversity inhabiting tissues of ebony (*Diospyros celebica* Bakh .) in urban forest. *2nd Biennial Conference of Tropical Biodiversity*.

<https://doi.org/10.1088/1755-1315/886/1/012031>

Munanda, R., Yusran, Wardah, & Rahmawati. (2022). Populasi Mikroba Tanah pada Lahan Agroforestri dan Monokultur Kakao Di Cagar Alam Pangi Binangga Kabupaten Parigi Moutong. *Wata Rimba : Jurnal Ilmiah Kehutanan*, 10(3), 161–168.

Nganji, M. U., & Simanjuntak, B. H. (2020). Penentuan Pola Tanam Tanaman Pangan Berdasarkan Neraca Keseimbangan Air di Kecamatan Umbu Ratu Nggay Barat, Kabupaten Sumba Tengah, Provinsi Nusa Tenggara Timur.

Jurnal Ilmiah Teknologi Pertanian Agrotechno, 5(2), 67.

<https://doi.org/10.24843/jitpa.2020.v05.i02.p04>

Nugroho, P. A. (2012). The Potency of Natural Rubber Development Under Industrial Plant Forest Scheme. *Warta Perkaretan*, 31(2), 95–102.

Pham, N. Q., Barnes, I., Chen, S. F., Pham, T. Q., Lombard, L., Crous, P. W., & Wingfield, M. J. (2018). New species of Cylindrocladiella from plantation soils in South-East Asia. *MycoKeys*, 32, 1–24.

<https://doi.org/10.3897/mycokeys.32.23754>

Prasetyo, A. D., Indriyanto, & Riniarti, M. (2019). Jenis-jenis Tanaman Di Lahan

- Garapan Petani KPPH Wana Makmur dalam Tahura Wan Abdul Rachman. *EnviroScienteae*, 15(2), 154–165.
- Prayudyaningsih, R., Nursyamsi, & Sari, R. (2015). Mikroorganisme tanah bermanfaat pada rhizosfer tanaman umbi di bawah tegakan hutan rakyat Sulawesi Selatan. *Pros Sem Nas Masy Biodiv Indon*, 1(4), 954–959.
<https://doi.org/10.13057/psnmbi/m010453>
- Rahma, Y. A., & Karimah, I. (2021). Eksplorasi dan Identifikasi Agen Hayati *Gliocladium* sp. dalam Menghambat Pertumbuhan Cendawan Patogen *Collectotrichum* sp. *Prosiding Seminar Nasional BIO*, 1, 432–440.
- Rauf, C. A., Ahmad, I., & Ashraf, M. (2007). Anastomosis groups of *Rhizoctonia solani* Kühn isolates from potato in Pakistan. *Pakistan Journal of Botany*, 39(4), 1335–1340.
- Ristiari, N. P. N., Julyasih, K. S. M., & Suryanti, I. A. P. (2018). Isolasi dan Identifikasi Jamur Mikroskopis pada Rizosfer Tanaman Jeruk Siam (*Citrus nobilis* Lour .) Di Kecamatan Kintamani, Bali. *Jurnal Pendidikan Biologi Undiksha*, 6(1), 10–19.
- Robianto, & Supijatno. (2017). Sistem Penyadapan Karet (*Hevea brasiliensis* Muell. Arg.) di Tulung Gelam Estate, Sumatera Selatan. *Agrohorti*, 5(2), 274–282.
- Saif, F. A., Yaseen, S., Alameen, A., Mane, S., & Undre, P. (2020). Identification of *Penicillium* Species of Fruits Using Morphology and Spectroscopic Methods. *Journal of Physics : Conference Series*, 1–10.
<https://doi.org/10.1088/1742-6596/1644/1/012019>
- Sarah, Asrul, & Lakani, I. (2018). Uji Antagonis Jamur *Aspergillus niger* terhadap

- Perkembangan Jamur Patogenik Fusarium oxysporum pada Bawang Merah (Allium cepa aggregatum L . aggregatum group) Secara In Vitro. *Agrotekbis*, 6(2), 266–273.
- Sardjono, M. A., Djogo, T., Arifin, H. S., & Wijayanto, N. (2003). Klasifikasi dan pola kombinasi komponen agroforestri. *World Agroforestry Centre (ICRAF)*, *Bagian 1*, 25.
- Sofiani, I. H., Ulfiah, K., & Fitriyanie, L. (2018). Rubber Tree (*Hevea brasiliensis*) Cultivation In Indonesia and Its Economic Study. *Munich Personal RePEc Archive*, 90336.
- Soldati, G. T., & Albuquerque, U. P. de. (2008). Non-Timber Forest Products in Ontario: An Overview. *Functional Ecosystems and Communities*, 2(Special Issue 1), 21–31.
<http://www.mnr.gov.on.ca/stdprodconsume/groups/lr/@mnr/@ofri/documents/documents/279238.pdf>
- Solihin, M. A., & Fitriatin, B. N. (2017). Sebaran Mikroba Tanah pada Berbagai Jenis Penggunaan Lahan Di Kawasan Bandung Utara. *SoilREns*, 15(1), 38–45. <https://doi.org/10.24198/soilreens.v15i1.13345>
- Suryani, Y., Taupiqurrahman, O., & Kulsum, Y. (2020). Mikologi. In *PT. Freeline Cipta Granesia*.
- Syahputra, N., Mawardati, & Suryadi. (2017). Analisis Faktor yang Mempengaruhi Petani Memilih Pola Tanam pada Tanaman Perkebunan Di Desa Paya Palas Kecamatan Ranto Peureulak Kabupaten Aceh Timur. *Agrifo*, 2(1), 41–50.
- Usuman, I., & Fitriyaningsih. (2012). Penerapan Sistem Integrasi Elektronik dan

- Pengamatan Perlakuan Sifat Jamur Berdasarkan Suhu dan Kelembaban Pada Ruang Tumbuh Jamur ikasi RFID untuk Sistem Kuping (Auricularia Sp.). *IJEIS*, 1(2), 11–20.
- Utama, A. P., Ristiati, N. P., & Suryanti, I. A. P. (2018). Jumlah Total Koloni Jamur Endofti pada Tanaman Anggur Bali (*Vitis vinifera L . var Alphonso Lavalle*) Di Desa Banjar, Kecamatan Banjar, Buleleng Bali. *Pendidikan Biologi Undiksha*, 5(3), 166–175.
- Waren, L., Freitas, S. De, José, R., Oliveira, V. De, Rhafhaella, T., & Leite, C. (2021). *Gongronella pedratalhadensis* , a new species of Mucorales (Mucromycota) isolated from the Brazilian. *Sydowia, January*.
<https://doi.org/10.12905/0380.sydowia72-2020-0061>
- Widyati, E. (2016). Efektivitas Rhizosfir pada Sistem Agroforestri: Sebuah Pemahaman dari Sudut Pandang Biologi Tanah. *Bpp/hhb*, 2(1), 1–11.
- Widyati, E. (2017). Memahami Komunikasi Tumbuhan-Tanah dalam Areal Rhizosfir untuk Optimasi Pengelolaan Lahan. *Jurnal Sumberdaya Lahan*, 11(1), 33–42.
- Wulandari, N. L. D., Proborini, M. W., & Sundra, I. K. (2013). Spasial Exploration of Fungi in Rhizosphere A Cashew Plantations (*Anacardium occidentale L.*) At Karangasem and Buleleng-Bali. *Simbiosis*, 1(2), 85–101.
- Yulia, E., Rahayu, A., & Suganda, T. (2022). Antagonisme Jamur Rizosfer Tanaman Karet terhadap *Rigidoporus microporus* secara In Vitro dan In Planta. *Jurnal Agro*, 9(1), 64–79.
- Zhang, Z., Han, Y., Chen, W., & Liang, Z. (2019). *Gongronella sichuanensis* (Cunninghamellaceae, Mucorales), a new species isolated from soil in

China. *Phytotaxa*, 416(2), 167–174.

LAMPIRAN

Lampiran 1. Proses Pengambilan Sampel

Pengambilan sampel kulit pada tanaman karet yang sedang disadap



Pengambilan sampel kulit tanah pada pola tanam monokultur



Pengambilan sampel daun pada tanaman karet yang sedang disadap



Pengukuran tinggi pohon pada pola tanam Agroforestri

Lampiran 2. Proses Penggeraan di Laboratorium

Menimbang Agar-Agar dan Glukosa untuk pembuatan media *Potato Dextrose Agar*



Menuang larutan media *Potato Dextrose Agar* pada cawan petri



Tahap isolasi Cendawan (metode pengenceran bertingkat)



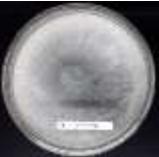
Identifikasi Mikroskopik cendawan menggunakan mikroskop

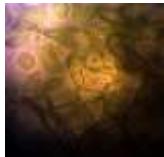
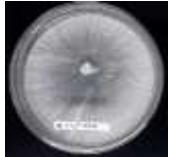
Lampiran 3. Deskripsi Plot

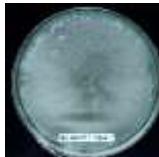
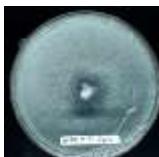
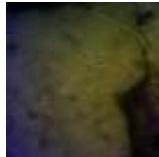
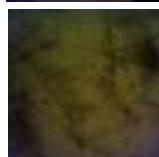
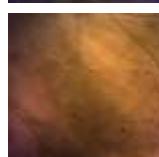
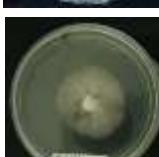
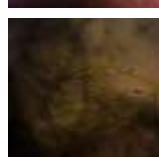
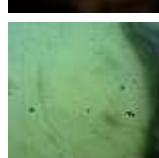
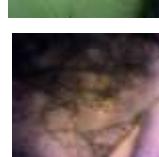
	Elevasi (mdpl)	Kelembaban (%)	Suhu (°C)	Jarak Tanam (m)	Umur (Tahun)
Monokultur (Sadap)					
Plot 1	306	76	22	6 x 3.5	20
Plot 2	293	73	25	4 x 7	10
Plot 3	285	57	31	4 x 7	8
Monokultur (Belum Sadap)					
Plot 4	310	63	32	6 x 4	3
Plot 5	303	73	29	3 x 3	3
Plot 6	296	81	23	5 x 3	3
Agroforestri (Sadap)					
Plot 7	286	78	25	4 x 3	15
Plot 8	345	70	26	3 x 3.5	12
Plot 9	118	74	26	4 x 3	11
Agroforestri (Belum Sadap)					
Plot 10	324	71	29	4 x 4.5	5
Plot 11	110	82	24	4 x 3	5
Plot 12	151	70	27	3 x 6	7

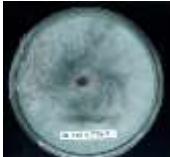
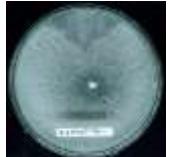
Lampiran 4. Identifikasi Makroskopis dan Mikroskopis Isolat Cendawan

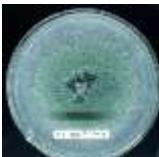
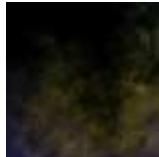
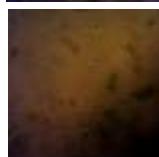
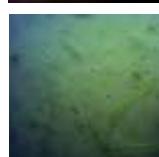
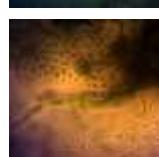
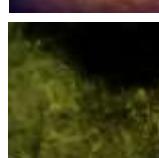
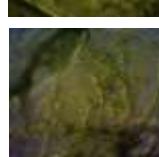
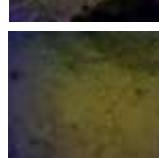
pada Tanaman Karet

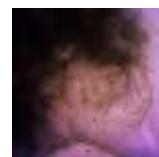
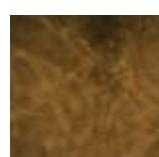
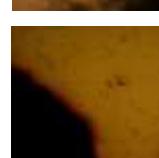
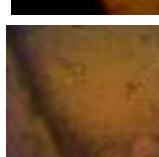
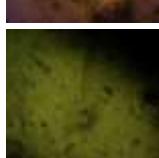
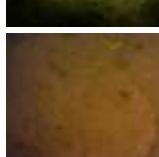
No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
1	HB 1 (2) ^I			<i>Trichoderma</i>
2	HB 1 (3) ^I			<i>Aspergillus</i>
3	HB 1 (3) ^{II}			<i>Trichoderma</i>
4	HB 1 (4)(10 ⁻²) ^I			<i>Trichoderma</i>
5	HB 1 (4)(10 ⁻²) ^{II}			<i>Trichoderma</i>
6	HB 1 (4)(10 ⁻²) ^{III}			<i>Aspergillus</i>
7	HB 1 (4)(10 ⁻³) ^I			<i>Trichoderma</i>
8	HB 1 (4)(10 ⁻³) ^{II}			X

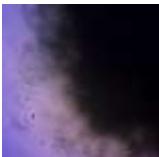
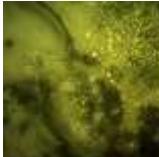
No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
9	HB 2 (1) ^I			X
10	HB 2 (2) ^I			X
11	HB 2 (2) ^{II}			<i>Rhizoctonia</i>
12	HB 2 (3) ^I			<i>Trichoderma</i>
13	HB 2 (4)(10 ⁻²) ^I			X
14	HB 2 (4)(10 ⁻²) ^{II}			<i>Penicillium</i>
15	HB 2 (4)(10 ⁻³) ^I			<i>Aspergillus</i>
16	HB 3 (2) ^I			<i>Cylindrocladiella</i>
17	HB 3 (3) ^I			X

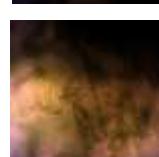
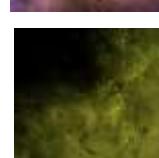
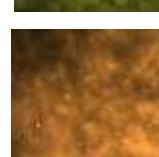
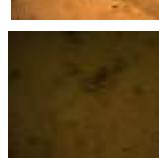
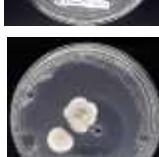
No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
18	HB 3 (4)(10 ⁻²) ^I			<i>Trichoderma</i>
19	HB 3 (4)(10 ⁻²) ^{II}			<i>Trichoderma</i>
20	HB 3 (10 ⁻³)			<i>Rhizoctonia</i>
21	HB 3(4) (10 ⁻³) ^{II}			X
22	HB 4 (1) ^I			X
23	HB 4 (1) ^{II}			X
24	HB 4 (2) ^I			<i>Cylindrocladiella</i>
25	HB 4 (2) ^{II}			<i>Cylindrocladiella</i>
26	HB 4 (3) ^I			<i>Trichoderma</i>

No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
27	HB 4 (4)(10 ⁻²) ^I			<i>Rhizoctonia</i>
28	HB 4 (4)(10 ⁻²) ^{II}			<i>Penicillium</i>
29	HB 4 (4)(10 ⁻³) ^I			<i>Trichoderma</i>
30	HB 4 (4)(10 ⁻³) ^{II}			<i>Trichoderma</i>
31	HB 5 (1) ^I			<i>Cylindrocladiella</i>
32	HB 5 (2) ^I			<i>Cylindrocladiella</i>
33	HB 5 (3) ^I			<i>Trichoderma</i>
34	HB 5 (4)(10 ⁻²) ^I			<i>Trichoderma</i>
35	HB 5 (4)(10 ⁻²) ^{II}			<i>Penicillium</i>

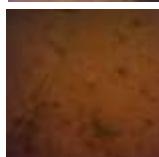
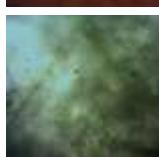
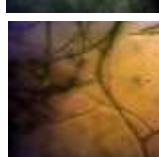
No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
36	HB 5 (4)(10 ⁻²) ^{III}			<i>Trichoderma</i>
37	HB 5 (4)(10 ⁻³) ^I			<i>Trichoderma</i>
38	HB 5 (4)(10 ⁻³) ^{II}			<i>Penicillium</i>
39	HB 6 (1) ^I			X
40	HB 6 (2) ^I			<i>Cylindrocladiella</i>
41	HB 6 (3) ^I			<i>Trichoderma</i>
42	HB 6 (4)(10 ⁻²) ^I			<i>Trichoderma</i>
43	HB 6 (4)(10 ⁻³) ^I			<i>Rhizoctonia</i>
44	HB 7 (1) ^I			<i>Cunningmahella</i>

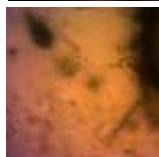
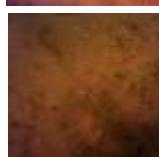
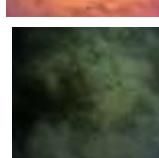
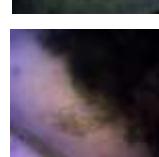
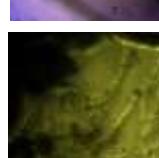
No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
45	HB 7 (1) ^{II}			<i>Cylindrocladiella</i>
46	HB 7 (2) ^I			<i>Cylindrocladiella</i>
47	HB 7 (3) ^I			<i>Cunningmahela</i>
48	HB 7 (3) ^{II}			<i>Trichoderma</i>
49	HB 7 (3) ^{III}			<i>Paecilomyces</i>
50	HB 7 (4)(10 ⁻²) ^I			<i>Penicillium</i>
51	HB 7 (4)(10 ⁻²) ^{II}			<i>Paecilomyces</i>
52	HB 7 (4)(10 ⁻²) ^{III}			<i>Penicillium</i>
53	HB 7 (4)(10 ⁻²) ^{IV}			X

No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
54	HB 7 (4)(10 ⁻³) ^I			<i>Trichoderma</i>
55	HB 7 (4)(10 ⁻³) ^{II}			<i>Trichoderma</i>
56	HB 8 (1) ^I			<i>Cylindrocladiella</i>
57	HB 8 (2) ^I			<i>Cylindrocladiella</i>
58	HB 8 (2) ^{II}			X
59	HB 8 (3) ^I			<i>Trichoderma</i>
60	HB 8 (4)(10 ⁻²) ^I			<i>Trichoderma</i>
61	HB 8 (4)(10 ⁻²) ^{II}			<i>Trichoderma</i>
62	HB 8 (4)(10 ⁻²) ^{III}			<i>Aspergillus</i>

No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
63	HB 8 (4)(10 ⁻³) ^I			X
64	HB 8 (4)(10 ⁻³) ^{II}			<i>Mucor</i>
65	HB 9 (1) ^I			<i>Cylindrocladiella</i>
66	HB 9 (1) ^{II}			X
67	HB 9 (2) ^I			<i>Cylindrocladiella</i>
68	HB 9 (3) ^I			<i>Trichoderma</i>
69	HB 9 (3) ^{II}			<i>Trichoderma</i>
70	HB 9 (4)(10 ⁻²) ^I			<i>Penicillium</i>
71	HB 9 (4)(10 ⁻²) ^{II}			<i>Gongronella</i>

No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
72	HB 9 (4)(10 ⁻³) ^I			<i>Penicillium</i>
73	HB 10 (1) ^I			<i>Aspergillus</i>
74	HB 10 (1) ^{II}			X
75	HB 10 (2) ^I			<i>Mucor</i>
76	HB 10 (2) ^{II}			X
77	HB 10 (3) ^I			X
78	HB 10 (3) ^{II}			X
79	HB 10 (4)(10 ⁻²) ^I			<i>Aspergillus</i>
80	HB 10 (4)(10 ⁻²) ^{II}			X

No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
81	HB 10 (4)(10 ⁻²) ^{III}			<i>Aspergillus</i>
82	HB 10 (4)(10 ⁻³) ^I			X
83	HB 10 (4)(10 ⁻³) ^{II}			<i>Penicillium</i>
84	HB 11 (1) ^I			<i>Cylindrocladiella</i>
85	HB 11 (2) ^I			<i>Rhizoctonia</i>
86	HB 11 (3) ^I			<i>Trichoderma</i>
87	HB 11 (3) ^{II}			<i>Penicillium</i>
88	HB 11 (3) ^{III}			<i>Aspergillus</i>
89	HB 11 (3) ^{IV}			<i>Penicillium</i>

No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
90	HB 11 (4)(10 ⁻²) ^I			<i>Gongronella</i>
91	HB 11 (4)(10 ⁻²) ^{II}			<i>Penicillium</i>
92	HB 11 (4)(10 ⁻²) ^{III}			<i>Penicillium</i>
93	HB 11 (4)(10 ⁻³) ^I			<i>Trichoderma</i>
94	HB 11 (4)(10 ⁻³) ^{II}			<i>Gliocladium</i>
95	HB 12 (1) ^I			<i>Cylindrocladiella</i>
96	HB 12 (2) ^I			<i>Cylindrocladiella</i>
97	HB 12 (3) ^I			<i>Trichoderma</i>
98	HB 12 (3) ^{II}			<i>Trichoderma</i>

No	Kode Isolat	Gambar		Genus
		Makroskopis	Mikroskopis	
99	HB 12 (4)(10 ⁻²) ^I			X
100	HB 12 (4)(10 ⁻²) ^{II}			<i>Trichoderma</i>
101	HB 12 (4)(10 ⁻³) ^I			<i>Penicillium</i>

Lampiran 5. Uji T-Test

Group Statistics

	Pola Tanam	N	Mean	Std. Deviation	Std. Error Mean
Jumlah Isolat	Monokultur	6	7.1667	1.47196	.60093
	Agroforestri	6	9.6667	1.96638	.80277

Independent Samples Test

	Levene's Test for Equality of Variances			t-test for Equality of Means					
	F	Sig.	t	df	Sig. (2-tailed)	Mean Difference	Std. Error Difference	95% Confidence Interval of the Difference	
								Lower	Upper
Jumlah Isolat	Equal variances assumed	1.406	.263	-2.493	10	.032	-2.50000	1.00277	-4.73432
	Equal variances not assumed			-2.493	9.264	.034	-2.50000	1.00277	-4.75860
									.26568
									.24140

Jika, nilai Sig. < 0,05 maka terdapat perbedaan signifikan antara pola tanam, dan nilai Sig > 0,05 tidak terdapat perbedaan signifikan antara pola tanam

Lampiran 6. Anova Pola Tanam

Jumlah Isolat Berdasarkan Pola Tanam	
Monokultur	Agroforestri
8	12
7	9
6	8
9	11
8	11
5	7

Pola Tanam	Rata-Rata	Simpangan Baku	Banyak Data	Standar Error
Agroforestri	9.666666667	1.966384161	6	0.802772972
Monokultur	7.166666667	1.471960144	6	0.600925213

ANOVA

Jumlah Isolat

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	18.750	1	18.750	6.215	.032
Within Groups	30.167	10	3.017		
Total	48.917	11			

Jika Sig > 0.05 maka H0 diterima, Sig <0.05 maka H0 ditolak

Lampiran 7. Anova Status Penyadapan

Jumlah Isolat	
Sadap	Belum Sadap
8	9
7	8
6	5
12	11
9	11
8	7
50	51

Status Sadap	Rata-Rata	Simpangan Baku	Banyak Data	Standar error
Sadap	14.28571	15.86100339	6	6.475227519
Belum Sadap	14.57143	16.20552522	6	6.615877967

ANOVA

Jumlah Isolat

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	.083	1	.083	.017	.899
Within Groups	48.833	10	4.883		
Total	48.917	11			

Jika Sig > 0.05 maka H0 diterima, Sig <0.05 maka H0 ditolak

Lampiran 8. Anova Jaringan dan Rhizosfer

Jumlah Isolat			
Daun	Kulit	Akar	Tanah
5	8	7	23
9	8	14	27

Jaringan	Rata-Rata	Simpangan Baku	Banyak Data	Standart Error
Daun	7	2.828427125	2	2
Kulit	8	0	2	0
Akar	10.5	4.949747468	2	3.5
Tanah	25	2.828427125	2	2

ANOVA

Jumlah Isolat

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	70.229	3	23.410	30.074	.000
Within Groups	34.250	44	.778		
Total	104.479	47			

Jika Sig > 0.05 maka H0 diterima, Sig <0.05 maka H0 ditolak

Lampiran 9. Uji Tukey Jaringan dan Rhizosfer

Jumlah Isolat

Tukey HSD^a

Jaringan dan Rhizosfer	N	Subset for alpha = 0.05	
		1	2
Daun	12	1.1667	
Kulit	12	1.3333	
Akar	12	1.7500	
Tanah	12		4.1667
Sig.		.378	1.000

Means for groups in homogeneous subsets are displayed.

a. Uses Harmonic Mean Sample Size = 12.000.

Lampiran 10. Analisis Korelasi

		Correlations		
		Jumlah Isolat	Jumlah Genus	Jumlah Jenis Tanaman Campuran
Jumlah Isolat	Pearson Correlation	1	.577*	.511
	Sig. (2-tailed)		.049	.090
	N	12	12	12
Jumlah Genus	Pearson Correlation	.577*	1	.480
	Sig. (2-tailed)	.049		.114
	N	12	12	12
Jumlah Jenis Tanaman Campuran	Pearson Correlation	.511	.480	1
	Sig. (2-tailed)	.090	.114	
	N	12	12	12

*. Correlation is significant at the 0.05 level (2-tailed).

Jika nilai Sig < 0,05, maka berkorelasi, nilai Sig > 0,05 tidak berkorelasi