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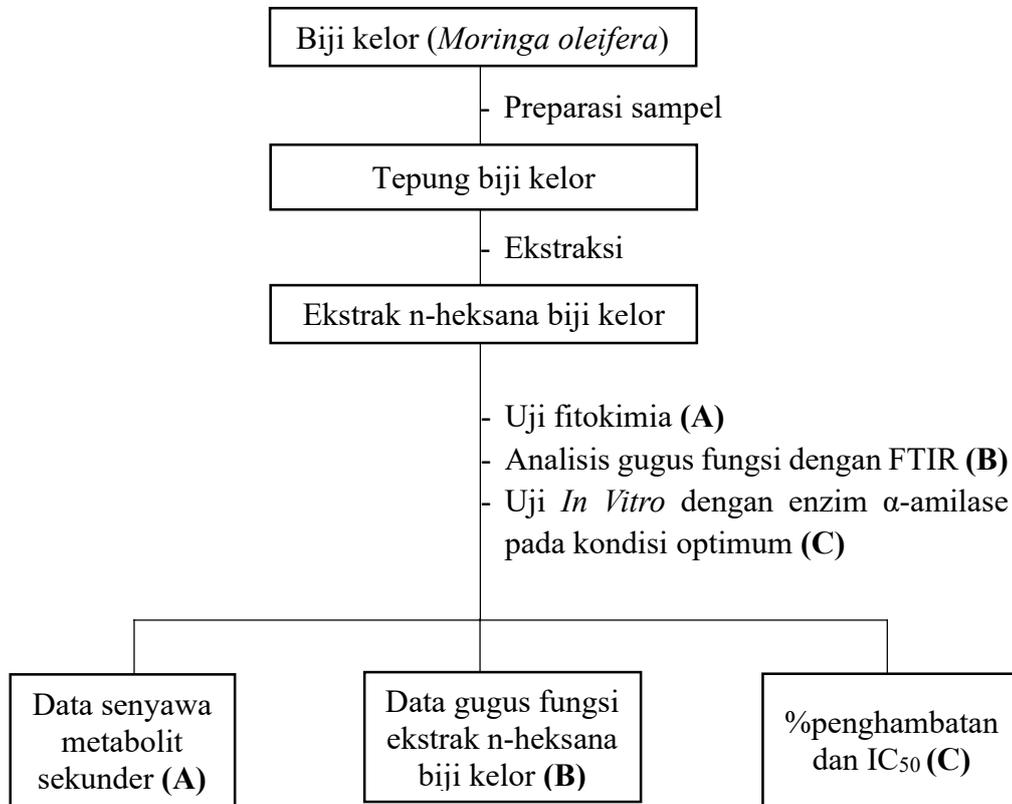
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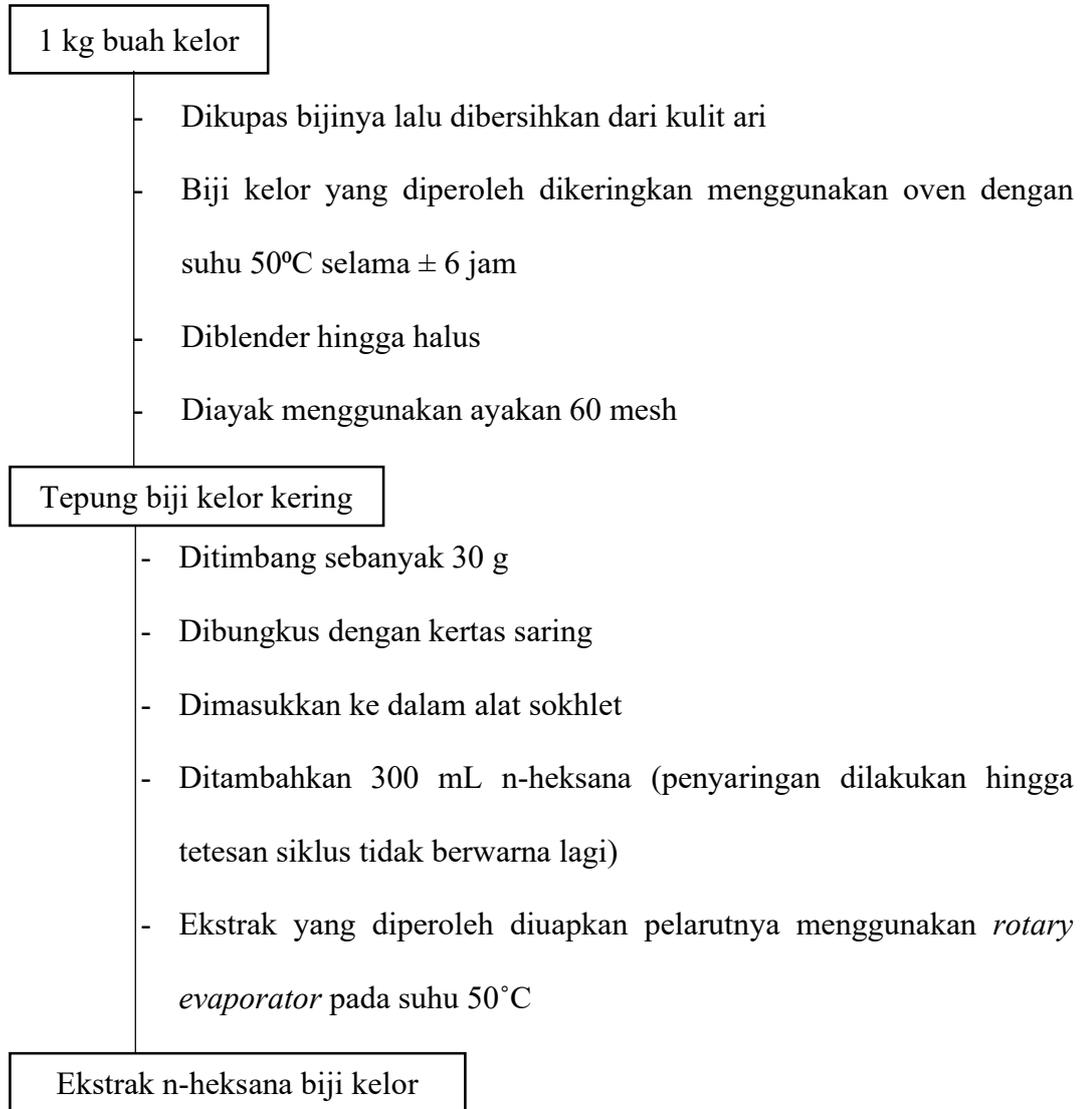
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**Lampiran 1.** Diagram Alir Penelitian



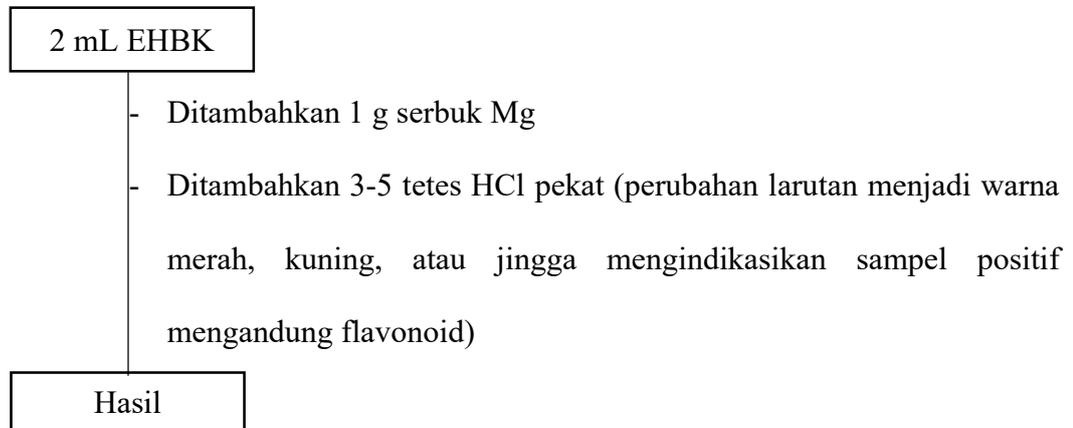
## Lampiran 2. Bagan Kerja

### 1. Preparasi dan Ekstraksi Biji Kelor

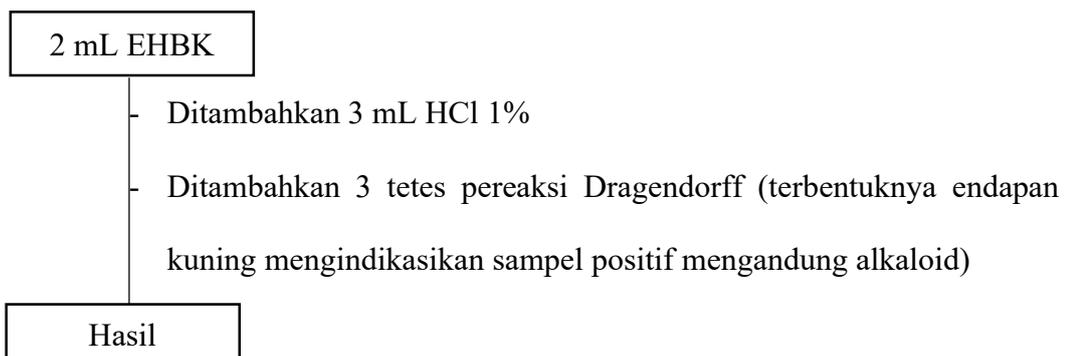


## 2. Uji Fitokimia

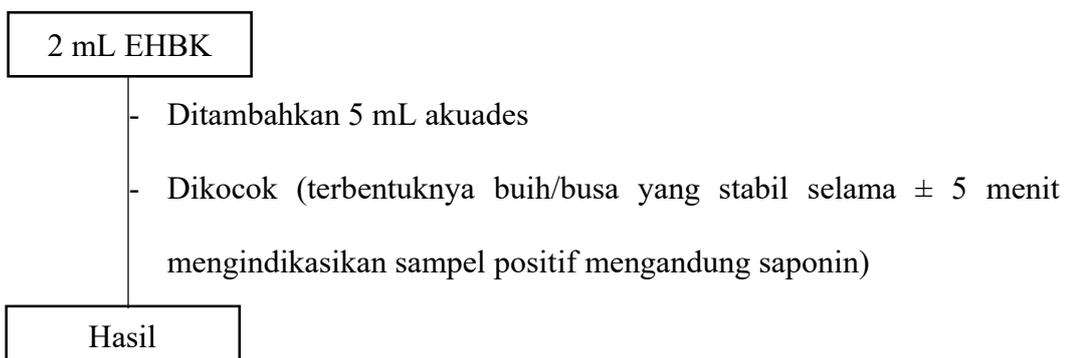
### a) Flavonoid



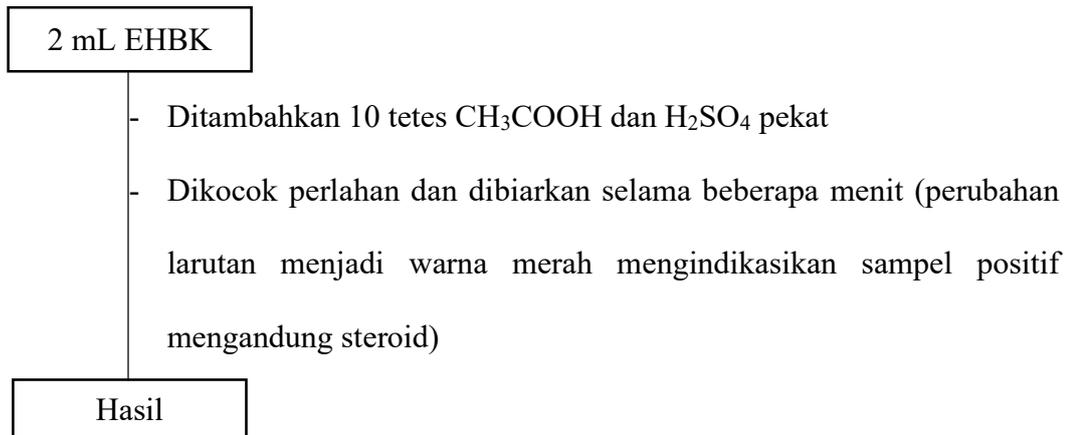
### b) Uji Alkaloid



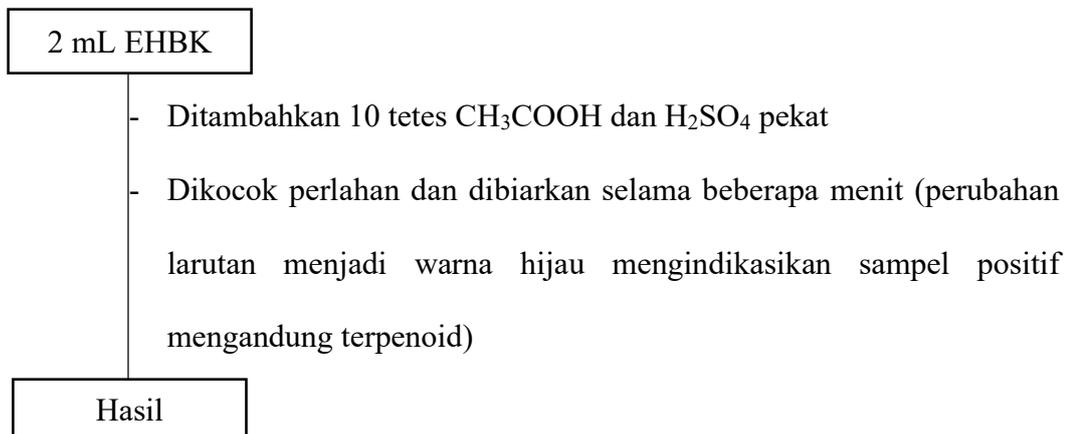
### c) Uji Saponin



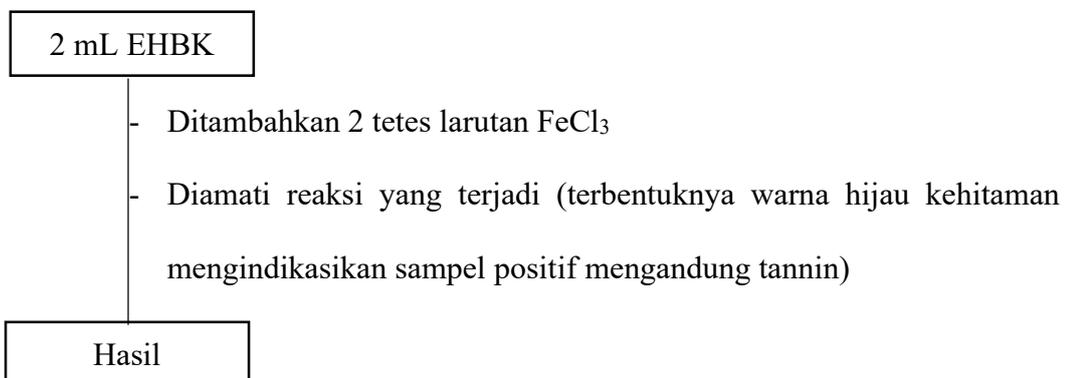
d) Uji Steroid



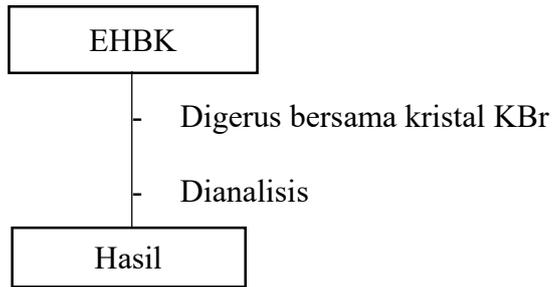
e) Uji Terpenoid



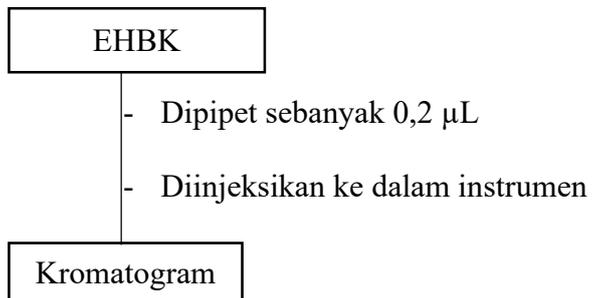
f) Uji Tanin



### 3. Uji FTIR

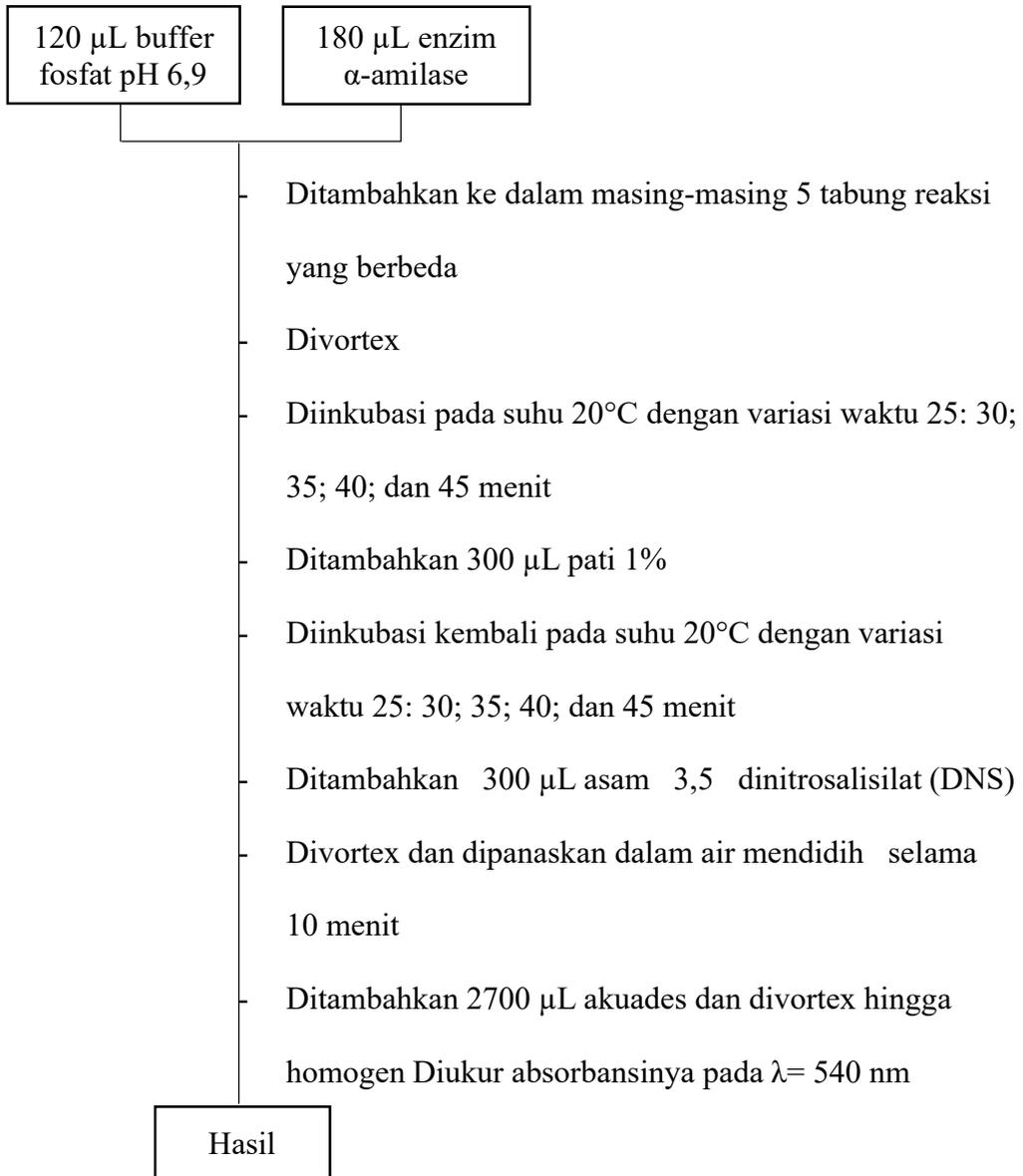


### 4. Uji GC-MS



## 5. Uji Aktivitas Enzim

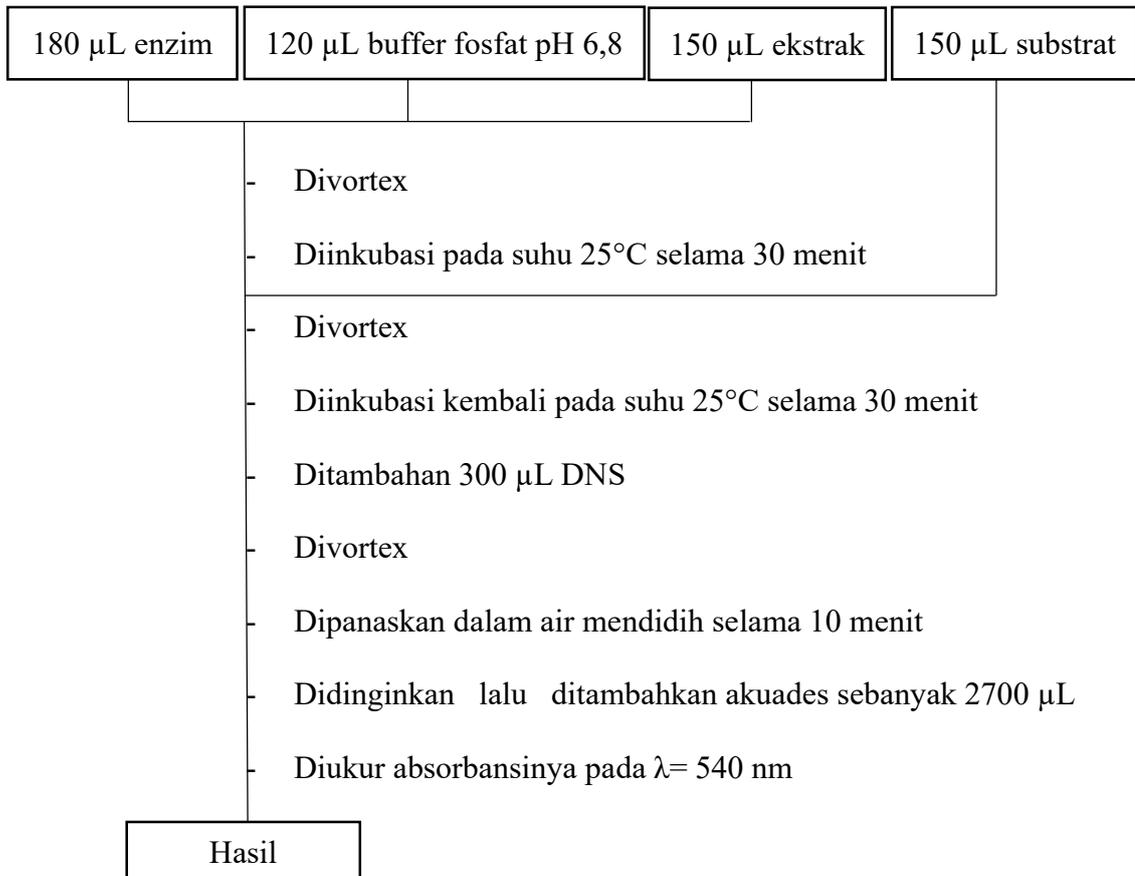
- Penentuan Waktu Optimum Enzim



### Catatan:

- Dilakukan prosedur yang sama pada penentuan pH optimum (variasi pH pH 6,4; 6,8; 6,9; 7; dan 7,4) dan suhu optimum (variasi suhu 20°C, 25°C, 30°C, 37°C, dan 40°C).
- Blanko dibuat tanpa penambahan enzim

6. Uji Aktivitas Penghambatan Enzim  $\alpha$ -amilase secara *In Vitro*



### Lampiran 3. Tempat Pengambilan Sampel



## Lampiran 4. Perhitungan Data Penelitian

### 1. Perhitungan Rendemen Ekstrak n-Heksana Biji Kelor

$$\begin{aligned}\% \text{ rendemen} &= \frac{\text{berat ekstrak (g)}}{\text{berat sampel (g)}} \times 100\% \\ &= \frac{5,09 \text{ g}}{30,01 \text{ g}} \times 100\% \\ &= 16,96\%\end{aligned}$$

### 2. Pembuatan Larutan

#### a. Buffer Fosfat

##### 1) Larutan stok

- Ditimbang  $\text{NaH}_2\text{PO}_4$  sebanyak 0,68995 g lalu dilarutkan ke dalam 25 mL akuades (A)
- Ditimbang  $\text{Na}_2\text{HPO}_4$  sebanyak 0,44497 g lalu dilarutkan ke dalam 25 mL akuades (B)

x mL larutan stok A + y mL larutan stok B, lalu dicukupkan volumenya hingga 10 mL

x (mL)	y (mL)	pH
3,675	1,325	6,4
2,55	2,75	6,8
2,25	1,325	6,9
1,95	3,05	7
0,95	4,05	7,4

b. Larutan NaOH 2 M dalam 50 mL

$$M = \frac{\text{massa}}{\text{MR}} \times \frac{1000}{V}$$

$$2 \text{ M} = \frac{\text{massa}}{40 \text{ g/mol}} \times \frac{1000}{50 \text{ mL}}$$

$$\text{massa} = 4 \text{ g}$$

$$w = \frac{2,5\%}{100\%} \times 10 \text{ mL}$$
$$= 0,25 \text{ g}$$

c. Larutan enzim  $\alpha$ -amilase

- Larutan induk

$$V_1 \times M_1 = V_2 \times M_2$$

$$V_1 \times 100,28 \text{ U/mL} = 10 \text{ mL} \times 20 \text{ U/mL}$$

$$V_1 = 1,994 \text{ mL}$$

$$= 1994 \mu\text{L}$$

- Larutan enzim 0,5 U/mL

$$V_1 \times M_1 = V_2 \times M_2$$

$$V_1 \times 20 \text{ U/mL} = 10 \text{ mL} \times 0,5 \text{ U/mL}$$

$$V_1 = 0,4 \text{ mL}$$

$$= 400 \mu\text{L}$$

### 3. Perhitungan %inhibisi

a. Ekstrak n-heksana biji kelor 5%

$$\%inhibisi = \frac{\text{Absorbansi Kontrol}-\text{Absorbansi Sampel}}{\text{Absorbansi Kontrol}} \times 100\%$$

$$\%inhibisi = \frac{0,340-0,167}{0,340} \times 100\%$$

$$= 55,4\%$$

b. Ekstrak n-heksana biji kelor 10%

$$\%inhibisi = \frac{\text{Absorbansi Kontrol}-\text{Absorbansi Sampel}}{\text{Absorbansi Kontrol}} \times 100\%$$

$$\%inhibisi = \frac{0,340-0,154}{0,340} \times 100\%$$

$$= 58,4\%$$

c. Ekstrak n-heksana biji kelor 15%

$$\%inhibisi = \frac{\text{Absorbansi Kontrol}-\text{Absorbansi Sampel}}{\text{Absorbansi Kontrol}} \times 100\%$$

$$\%inhibisi = \frac{0,340-0,121}{0,121} \times 100\%$$

$$= 67,6\%$$

d. *Acarbose* 5%

$$\%inhibisi = \frac{\text{Absorbansi Kontrol}-\text{Absorbansi Sampel}}{\text{Absorbansi Kontrol}} \times 100\%$$

$$\%inhibisi = \frac{0,340-0,293}{0,340} \times 100\%$$

$$= 13,82\%$$

e. *Acarbose* 10%

$$\%inhibisi = \frac{\text{Absorbansi Kontrol}-\text{Absorbansi Sampel}}{\text{Absorbansi Kontrol}} \times 100\%$$

$$\%inhibisi = \frac{0,340-0,251}{0,340} \times 100\%$$

$$= 26,17\%$$

f. *Acarbose* 15%

$$\%inhibisi = \frac{\text{Absorbansi Kontrol}-\text{Absorbansi Sampel}}{\text{Absorbansi Kontrol}} \times 100\%$$

$$\%inhibisi = \frac{0,340-0,198}{0,340} \times 100\%$$

$$= 41,76\%$$

## Lampiran 5. Dokumentasi Penelitian



Sampel biji kelor kering



Proses penghalusan sampel



Biji kelor yang telah halus



Penimbangan biji kelor



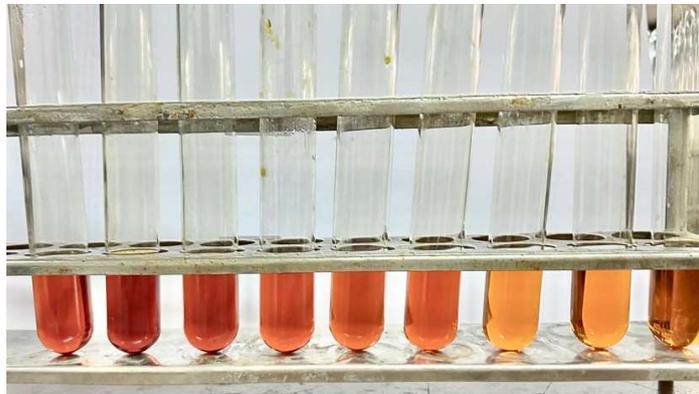
Proses sokhletasi



Proses evaporasi



Deret standar glukosa



Uji aktivitas inhibisi enzim  $\alpha$ -amilase dengan ekstrak n-heksana biji kelor