

DAFTAR PUSTAKA

- Adamski, R. (2012). *Factors affecting the color of roasted cocoa bean* . 21–31.
- Anand, S. S., Philip, B. K., & Mehendale, H. M. (2014). Volatile Organic Compounds. In *Encyclopedia of Toxicology: Third Edition* (Third Edit, Vol. 4). Elsevier. <https://doi.org/10.1016/B978-0-12-386454-3.00358-4>
- Andruszkiewicz, J., Souza, R. N. D., Altun, I., Corno, M., & Kuhnert, N. (2019). *Thermally-induced formation of taste-active 2 , 5-diketopiperazines from short-chain peptide precursors in cocoa*. 121(March), 217–228. <https://doi.org/10.1016/j.foodres.2019.03.015>
- Andruszkiewicz, P. J., Souza, R. N. D., Corno, M., & Kuhnert, N. (2020). Novel Amadori and Heyns compounds derived from short peptides found in dried cocoa beans. *Food Research International*, 133(December 2019), 109164. <https://doi.org/10.1016/j.foodres.2020.109164>
- Anonim. (1998). *Basics of LC/MS*. Hewlett-Packard Company.
- anonymous. (2012). Carboxylic Acids. *Enological Chemistry*, 109–120. <https://doi.org/10.1016/b978-0-12-388438-1.00008-x>
- Api, A. M., Belsito, D., Botelho, D., Browne, D., Bruze, M., Burton, G. A., Buschmann, J., Calow, P., Dagli, M. L., Date, M., Dekant, W., Deodhar, C., Fryer, A. D., Joshi, K., La Cava, S., Lapczynski, A., Liebler, D. C., O'Brien, D., Parakhia, R., ... Wahler, J. (2017). RIFM fragrance ingredient safety assessment, isoamyl acetate, CAS Registry Number 123-92-2. *Food and Chemical Toxicology*, 110(123), S123–S132. <https://doi.org/10.1016/j.fct.2017.03.046>
- Appendix, S. (1976). *Chapter 4: Gas chromatography*. [https://doi.org/10.1016/s0301-4770\(08\)60731-9](https://doi.org/10.1016/s0301-4770(08)60731-9)
- Apriyanto, M., Sutardi, S., Supriyanto, S., & Harmayani, E. (2017). Amino acid analysis of cocoa fermented by high performance liquid chromatography (HPLC). *Asian Journal of Dairy and Food Research*, 36(02), 156–160. <https://doi.org/10.18805/ajdfr.v36i02.7962>
- Aprotosoai, A. C., Luca, S. V., & Miron, A. (2016). *Flavor Chemistry of Cocoa and Cocoa Products — An Overview*. 15, 73–91. <https://doi.org/10.1111/1541-4337.12180>

- Arihara, K., Yokoyama, I., & Ohata, M. (2019). DMHF (2,5-dimethyl-4-hydroxy-3(2H)-furanone), a volatile food component with attractive sensory properties, brings physiological functions through inhalation. In *Advances in Food and Nutrition Research* (1st ed., Vol. 89). Elsevier Inc. <https://doi.org/10.1016/bs.afnr.2019.05.001>
- Ascrizzi, R., Flamini, G., Tessieri, C., & Pistelli, L. (2017). From the raw seed to chocolate: Volatile profile of Blanco de Criollo in different phases of the processing chain. *Microchemical Journal*, 133, 474–479. <https://doi.org/10.1016/j.microc.2017.04.024>
- Barišić, V., Kopjar, M., Jozinović, A., Flanjak, I., Ačkar, Đ., Miličević, B., Šubarić, D., Jokić, S., & Babić, J. (2019). The chemistry behind chocolate production. *Molecules*, 24(17). <https://doi.org/10.3390/molecules24173163>
- Basile, F., Zhang, S., Kandar, S. K., & Lu, L. (2011). Mass spectrometry characterization of the thermal decomposition/digestion (TDD) at cysteine in peptides and proteins in the condensed phase. *Journal of the American Society for Mass Spectrometry*, 22(11), 1926–1940. <https://doi.org/10.1007/s13361-011-0222-9>
- BeMiller, J. N. (2019a). Carbohydrate Reactions. *Carbohydrate Chemistry for Food Scientists*, 25–48. <https://doi.org/10.1016/b978-0-12-812069-9.00002-9>
- BeMiller, J. N. (2019b). Monosaccharides. *Carbohydrate Chemistry for Food Scientists*, 1–23. <https://doi.org/10.1016/b978-0-12-812069-9.00001-7>
- BeMiller, J. N. (2019c). Nonenzymic Browning and Formation of Acrylamide and Caramel. *Carbohydrate Chemistry for Food Scientists*, 351–370. <https://doi.org/10.1016/b978-0-12-812069-9.00018-2>
- Berk, Z. (2018). Frying, baking, and roasting. *Food Process Engineering and Technology*, 583–590. <https://doi.org/10.1016/b978-0-12-812018-7.00024-5>
- Bikaki, M., Shah, R., Müller, A., & Kuhnert, N. (2021). Heat induced hydrolytic cleavage of the peptide bond in dietary peptides and proteins in food processing. *Food Chemistry*, 357(June 2020), 129621. <https://doi.org/10.1016/j.foodchem.2021.129621>
- Bonvehí, J. S. (2005). Investigation of aromatic compounds in roasted cocoa powder. *European Food Research and Technology*, 221(1–2), 19–29.

<https://doi.org/10.1007/s00217-005-1147-y>

- Caligiani, A., Marseglia, A., & Palla, G. (2015). Cocoa: Production, Chemistry, and Use. In *Encyclopedia of Food and Health* (1st ed.). Elsevier Ltd. <https://doi.org/10.1016/B978-0-12-384947-2.00177-X>
- Calvo, A. M., Botina, B. L., García, M. C., Cardona, W. A., Montenegro, A. C., & Criollo, J. (2021). Dynamics of cocoa fermentation and its effect on quality. *Scientific Reports*, *11*(1), 1–15. <https://doi.org/10.1038/s41598-021-95703-2>
- Castro-Alayo, E. M., Idrogo-Vásquez, G., Siche, R., & Cardenas-Toro, F. P. (2019). Formation of aromatic compounds precursors during fermentation of Criollo and Forastero cocoa. *Heliyon*, *5*(1). <https://doi.org/10.1016/j.heliyon.2019.e01157>
- Cetinkaya, S., Yenidunya, A. F., Aksu, A., & Celik, A. M. (2021). *Purification and characterization of 1-dodecanol from an isolate of Streptomyces viridodiataticus, Biocatalysis and Agricultural Biotechnology*. ISSN 1878-8181. <https://doi.org/https://doi.org/10.1016/j.bcab.2021.102013>
- Colin Slaughter, J. (2007). The naturally occurring furanones: formation and function from pheromone to food. *Biological Reviews of the Cambridge Philosophical Society*, *74*(3), 259–276. <https://doi.org/10.1017/S0006323199005332>
- Crawford Scientific. (2004). *Mass Spectrometry Fundamental LC-MS Introduction*. 1–24. www.chromacademy.com
- Crews, C., & Castle, L. (2007). A review of the occurrence, formation and analysis of furan in heat-processed foods. *Trends in Food Science and Technology*, *18*(7), 365–372. <https://doi.org/10.1016/j.tifs.2007.03.006>
- Dand, R. (2011). *Cocoa bean processing and the manufacture of chocolate*. 268–289. <https://doi.org/10.1016/B978-0-85709-125-3.50009-4>
- Diab, J., Hertz-Schünemann, R., Streibel, T., & Zimmermann, R. (2014). Online measurement of volatile organic compounds released during roasting of cocoa beans. *Food Research International*, *63*, 344–352. <https://doi.org/10.1016/j.foodres.2014.04.047>
- Dimian, A. C., Bildea, C. S., & Kiss, A. A. (2019). Acetic Acid. Applications in Design and Simulation of Sustainable Chemical Processes. *European Chemical News*, *74*(1940), 483–519.

<https://doi.org/10.1002/047084289x.ra007>

- Dotson GS, Maier A, Parker A, H. L. C. (2016). *Immediately dangerous to life or health (IDLH) value profile: furan* (Issue 110).
- Effenberger, I., Hoffmann, T., Jonczyk, R., & Schwab, W. (2019). Novel biotechnological glucosylation of high-impact aroma chemicals, 3(2H)- and 2(5H)-furanones. *Scientific Reports*, 9(1), 1–9.
<https://doi.org/10.1038/s41598-019-47514-9>
- Fabbri, A., Cevoli, C., Alessandrini, L., & Romani, S. (2011). Numerical modeling of heat and mass transfer during coffee roasting process. *Journal of Food Engineering*, 105(2), 264–269.
<https://doi.org/10.1016/j.jfoodeng.2011.02.030>
- Fadai, N. T., Melrose, J., Please, C. P., Schulman, A., & Van Gorder, R. A. (2017). A heat and mass transfer study of coffee bean roasting. *International Journal of Heat and Mass Transfer*, 104, 787–799.
<https://doi.org/10.1016/j.ijheatmasstransfer.2016.08.083>
- Fang, Y., Li, R., Chu, Z., Zhu, K., Gu, F., & Zhang, Y. (2020). Chemical and flavor profile changes of cocoa beans (*Theobroma cacao* L.) during primary fermentation. *Food Science and Nutrition*, 8(8), 4121–4133.
<https://doi.org/10.1002/fsn3.1701>
- Farah, D. M. H., Zaibunnisa, A. H., Misnawi, J., & Zainal, S. (2012). Effect of Roasting Process on the Concentration of Acrylamide and Pyrazines in Roasted Cocoa Beans from Different Origins. *APCBEE Procedia*, 4, 204–208. <https://doi.org/10.1016/j.apcbee.2012.11.034>
- Fischetti, F. (2010). Flavoring Materials. *Kirk-Othmer Encyclopedia of Chemical Technology*, 1.
<https://doi.org/10.1002/0471238961.0612012206091903.a01.pub2>
- Flanjak, I., & Bariši, V. (2019). The Chemistry behind Chocolate Production. *MDPI*. <https://doi.org/doi:10.3390/molecules24173163>
- Frauendorfer, F., & Schieberle, P. (2006). Identification of the key aroma compounds in cocoa powder based on molecular sensory correlations. *Journal of Agricultural and Food Chemistry*, 54(15), 5521–5529.
<https://doi.org/10.1021/jf060728k>
- Frauendorfer, F., & Schieberle, P. (2019). Key aroma compounds in fermented Forastero cocoa beans and changes induced by roasting.

European Food Research and Technology, 0123456789.
<https://doi.org/10.1007/s00217-019-03292-2>

Gibson, M., & Newsham, P. (2018). Chocolate/Cacao. *Food Science and the Culinary Arts*, 341–352. <https://doi.org/10.1016/b978-0-12-811816-0.00017-8>

Goldberg, I., & Stefan Rokem, J. (2019). Organic and fatty acid production, microbial. *Encyclopedia of Microbiology*, 358–382.
<https://doi.org/10.1016/B978-0-12-809633-8.13083-3>

Hamdouche, Y., Meile, J. C., Lebrun, M., Guehi, T., Boulanger, R., Teyssier, C., & Montet, D. (2019). Impact of turning, pod storage and fermentation time on microbial ecology and volatile composition of cocoa beans. *Food Research International*, 119(January), 477–491.
<https://doi.org/10.1016/j.foodres.2019.01.001>

Harreus, A. L. (2011). *1. Introduction : 2-Pyrrolidone*.

Hashim, P., Selamat, J., Muhammad, S. K. S., & Ali, A. (1998). Changes in free amino acid, peptide-N, sugar and pyrazine concentration during cocoa fermentation. *Journal of the Science of Food and Agriculture*, 78(4), 535–542. [https://doi.org/10.1002/\(SICI\)1097-0010\(199812\)78:4<535::AID-JSFA151>3.0.CO;2-6](https://doi.org/10.1002/(SICI)1097-0010(199812)78:4<535::AID-JSFA151>3.0.CO;2-6)

Heath, H. B. (1999). *Source Book Of Flavors* (G. Reineccius (ed.); second). Aspen Publishers.

Hidalgo, F. J., Alcón, E., & Zamora, R. (2013). Cysteine- and serine-thermal degradation products promote the formation of Strecker aldehydes in amino acid reaction mixtures. *Food Research International*, 54(2), 1394–1399. <https://doi.org/10.1016/j.foodres.2013.09.006>

Ho, V. T. T., Zhao, J., & Fleet, G. (2014). Yeasts are essential for cocoa bean fermentation. *International Journal of Food Microbiology*, 174, 72–87.
<https://doi.org/10.1016/j.ijfoodmicro.2013.12.014>

Ho, V. T. T., Zhao, J., & Fleet, G. (2015). The effect of lactic acid bacteria on cocoa bean fermentation. *International Journal of Food Microbiology*, 205, 54–67. <https://doi.org/10.1016/j.ijfoodmicro.2015.03.031>

Hustiany, R. (2016). *REAKSI MAILLARD*. LMU Press. <https://repositorien.ulm.ac.id/handle/123456789/17832>

- Illegghems, K., Pelicaen, R., Vuyst, L. De, & Weckx, S. (2016). Assessment of the contribution of cocoa-derived strains of *Acetobacter ghanensis* and *Acetobacter senegalensis* to the cocoa bean fermentation process through a genomic approach. *Food Microbiology*, *58*, 68–78.
<https://doi.org/10.1016/j.fm.2016.03.013>
- Ismail, B., & Nielsen, S. S. (2010). Basic Principle of Chromatography. In S. S. Nielsen (Ed.), *Food Analysis*. Springer, New York.
<https://doi.org/10.1007/978-1-4419-1478-1>
- Jaeger, H., Janositz, A., & Knorr, D. (2010). La réaction de Maillard et son contrôle pendant la fabrication des aliments. Le potentiel des nouvelles technologies. *Pathologie Biologie*, *58*(3), 207–213.
<https://doi.org/10.1016/j.patbio.2009.09.016>
- Jati, M., & Budi Tunjung Sari, A. (2011). Analysis of Pyrazine and Volatile Compounds in Cocoa Beans Using Solid Phase Microextraction. *Pelita Perkebunan (a Coffee and Cocoa Research Journal)*, *27*(1), 24–35.
<https://doi.org/10.22302/icri.jur.pelitaperkebunan.v27i1.143>
- Jiang, X., Peng, D., Zhang, W., Duan, M., Ruan, Z., Huang, S., Zhou, S., & Fang, Q. (2021). Effect of aroma-producing yeasts in high-salt liquid-state fermentation soy sauce and the biosynthesis pathways of the dominant esters. *Food Chemistry*, *344*, 128681.
<https://doi.org/10.1016/j.foodchem.2020.128681>
- John, W. A., Böttcher, N. L., Aßkamp, M., Bergounhou, A., Kumari, N., Ho, P. W., D'Souza, R. N., Nevoigt, E., & Ullrich, M. S. (2019). Forcing fermentation: Profiling proteins, peptides and polyphenols in lab-scale cocoa bean fermentation. *Food Chemistry*, *278*, 786–794.
<https://doi.org/10.1016/j.foodchem.2018.11.108>
- Karimi, G., & Vahabzadeh, M. (2014). Butyric Acid. In *Encyclopedia of Toxicology: Third Edition* (Third Edit, Vol. 1). Elsevier.
<https://doi.org/10.1016/B978-0-12-386454-3.00591-1>
- Kirchhoff, P., Biehl, B., Hammor, M., & Lieberei, R. (1989). *Kinetics of the Formation of Free Amino Acids in Cocoa Seeds during Fermentation*. 1, 161–179.
- Koné, M. K., Guéhi, S. T., Durand, N., Ban-Koffi, L., Berthiot, L., Tachon, A. F., Brou, K., Boulanger, R., & Montet, D. (2016). Contribution of predominant yeasts to the occurrence of aroma compounds during cocoa bean fermentation. *Food Research International*, *89*, 910–917.

<https://doi.org/10.1016/j.foodres.2016.04.010>

- Kruis, A. J., Bohnenkamp, A. C., Patinios, C., van Nuland, Y. M., Levisson, M., Mars, A. E., van den Berg, C., Kengen, S. W. M., & Weusthuis, R. A. (2019). Microbial production of short and medium chain esters: Enzymes, pathways, and applications. *Biotechnology Advances*, 37(7), 107407. <https://doi.org/10.1016/j.biotechadv.2019.06.006>
- Krysiak, W. (2006). Influence of roasting conditions on coloration of roasted cocoa beans. *Journal of Food Engineering*, 77(3), 449–453. <https://doi.org/10.1016/j.jfoodeng.2005.07.013>
- Lefeber, T., Janssens, M., Moens, F., Gobert, W., & De Vuyst, L. (2011). Interesting starter culture strains for controlled cocoa bean fermentation revealed by simulated cocoa pulp fermentations of cocoa-specific lactic acid bacteria. *Applied and Environmental Microbiology*, 77(18), 6694–6698. <https://doi.org/10.1128/AEM.00594-11>
- Lemarcq, V., Tuenter, E., Bondarenko, A., De, D. Van, Vuyst, L. De, Pieters, L., Sioriki, E., & Dewettinck, K. (2020). Roasting-induced changes in cocoa beans with respect to the mood pyramid. *Food Chemistry*, 127467. <https://doi.org/10.1016/j.foodchem.2020.127467>
- López-Pedrouso, M., Lorenzo, J. M., Zapata, C., & Franco, D. (2019). Proteins and amino acids. In *Innovative Thermal and Non-Thermal Processing, Bioaccessibility and Bioavailability of Nutrients and Bioactive Compounds*. Elsevier Inc. <https://doi.org/10.1016/B978-0-12-814174-8.00005-6>
- M. J. Lane, & H. E. Nursten. (1983). The Maillard Reaction in Foods and Nutrition. In *Maillard Reaction in Chemistry, Food, and Health* (Vol. 215). Woodhead Publishing Ltd. <https://doi.org/10.1016/B978-1-85573-792-1.50007-2>
- Marczenko, Z., & Balcerzak, M. (. (2000). Principles of Spectrophotometry. Separation, Preconcentration and Spectrophotometry in Inorganic Analysis Chapter 2. *Analytical Spectroscopy Library*, 10(C), 26–38. [https://doi.org/10.1016/S0926-4345\(00\)80066-8](https://doi.org/10.1016/S0926-4345(00)80066-8)
- Marseglia, A., Musci, M., Rinaldi, M., Palla, G., & Caligiani, A. (2020). Volatile fingerprint of unroasted and roasted cocoa beans (*Theobroma cacao* L.) from different geographical origins. *Food Research International*, 132, 109101. <https://doi.org/10.1016/j.foodres.2020.109101>

- Mensah-brown, H., & Afoakwa, E. O. (2013). *Changes in Nib Acidity, Proteolysis and Sugar Concentration as Influenced by Pod Storage and Roasting Conditions of Fermented Cocoa (Theobroma cacao) Beans*. December 2014.
- Moro, S., Chipman, J. K., Wegener, J. W., Hamberger, C., Dekant, W., & Mally, A. (2012). Furan in heat-treated foods: Formation, exposure, toxicity, and aspects of risk assessment. *Molecular Nutrition and Food Research*, 56(8), 1197–1211. <https://doi.org/10.1002/mnfr.201200093>
- Morya, R., Salvachúa, D., & Thakur, I. S. (2020). Burkholderia: An Untapped but Promising Bacterial Genus for the Conversion of Aromatic Compounds. *Trends in Biotechnology*, 38(9), 963–975. <https://doi.org/10.1016/j.tibtech.2020.02.008>
- Mühlbauer, W., & Müller, J. (2020). Cocoa (*Theobroma cacao* L.). *Drying Atlas*, 239–245. <https://doi.org/10.1016/b978-0-12-818162-1.00028-6>
- Nashalian, O., & Yaylayan, V. A. (2016). In situ formation of the amino sugars 1-amino-1-deoxy-fructose and 2-amino-2-deoxy-glucose under Maillard reaction conditions in the absence of ammonia. *Food Chemistry*, 197, 489–495. <https://doi.org/10.1016/j.foodchem.2015.10.140>
- Nemr, K., Müller, J. E. N., Joo, J. C., Gawand, P., Choudhary, R., Mendonca, B., Lu, S., Yu, X., Yakunin, A. F., & Mahadevan, R. (2018). Engineering a short, aldolase-based pathway for (R)-1,3-butanediol production in *Escherichia coli*. *Metabolic Engineering*, 48, 13–24. <https://doi.org/10.1016/j.ymben.2018.04.013>
- Orozco, W., Clavijo, S., Delgado, Y., Ayala, C., & Zambrano, A. (2020). *The effect of fermentation and roasting on free amino acids profile in Criollo cocoa (Theobroma cacao L .) grown in Venezuela*. 1–12.
- Ouellette, R. J., & Rawn, J. D. (2015). Carboxylic Acids and Esters. *Principles of Organic Chemistry*, 287–314. <https://doi.org/10.1016/b978-0-12-802444-7.00011-2>
- Páramo, D., García-alamilla, P., Salgado-cervantes, M. A., Robles-olvera, V. J., & Rodríguez-jimenes, G. C. (2010). Mass transfer of water and volatile fatty acids in cocoa beans during drying. *Journal of Food Engineering*, 99(3), 276–283. <https://doi.org/10.1016/j.jfoodeng.2010.02.028>
- Parker, J. K. (2015a). Introduction to aroma compounds in foods. In *Flavour*

- Development, Analysis and Perception in Food and Beverages*. Elsevier Ltd. <https://doi.org/10.1016/b978-1-78242-103-0.00001-1>
- Parker, J. K. (2015b). Thermal generation of aroma. In *Flavour Development, Analysis and Perception in Food and Beverages*. Elsevier Ltd. <https://doi.org/10.1016/b978-1-78242-103-0.00008-4>
- Peterson, D. G., & Reineccius, G. A. (2017). *Gas Chromatography*. 227–253. <https://doi.org/10.1007/978-3-319-45776-5>
- Qian, M. C., Peterson, D. G., & Reineccius, G. A. (2010). Gas Chromatography. In S. S. Nielsen (Ed.), *Food Analysis*. Springer, New York. https://doi.org/10.1007/978-1-4419-1478-1_29
- Ramli, N., Hassan, O., Said, M., Samsudin, W., & Idris, N. A. (2006). Influence of roasting conditions on volatile flavor of roasted Malaysian cocoa beans. *Journal of Food Processing and Preservation*, 30(3), 280–298. <https://doi.org/10.1111/j.1745-4549.2006.00065.x>
- Rawel, H. M., Huscheck, G., Sagu, S. T., & Homan, T. (2019). *Cocoa Bean Proteins—Characterization, Changes and Modifications due to Ripening and Post-Harvest Processing*. <https://doi.org/10.3390/nu11020428>
- Rocha, I. S., De Santana, L. R. R., Soares, S. E., & Bispo, E. da S. (2017). Effect of the roasting temperature and time of cocoa beans on the sensory characteristics and acceptability of chocolate. *Food Science and Technology*, 37(4). <https://doi.org/10.1590/1678-457x.16416>
- Rocha, I. S., Regina, L., Santana, R. De, Soares, S. E., & Bispo, S. (2017). *Effect of the roasting temperature and time of cocoa beans on the sensory characteristics and acceptability of chocolate*. 37(4), 522–530.
- Rodriguez-Campos, J., Escalona-Buendía, H. B., Contreras-Ramos, S. M., Orozco-Avila, I., Jaramillo-Flores, E., & Lugo-Cervantes, E. (2012). Effect of fermentation time and drying temperature on volatile compounds in cocoa. *Food Chemistry*, 132(1), 277–288. <https://doi.org/10.1016/J.FOODCHEM.2011.10.078>
- Rojas S, M., Chejne, F., Ciro, H., & Montoya, J. (2020). Roasting impact on the chemical and physical structure of Criollo cocoa variety (*Theobroma cacao* L). *Journal of Food Process Engineering*, 43(6). <https://doi.org/10.1111/jfpe.13400>
- Rorrer, J., He, Y., Toste, F. D., & Bell, A. T. (2017). Mechanism and kinetics

- of 1-dodecanol etherification over tungstated zirconia. *Journal of Catalysis*, 354, 13–23. <https://doi.org/10.1016/j.jcat.2017.08.001>
- Sánchez-Castañeda, A. K., Athès, V., Moussa, M., López-Miranda, J., Páez-Lerma, J. B., Soto-Cruz, N. Ó., & Trelea, I. C. (2018). Modeling of isoamyl acetate production by fermentation with *Pichia fermentans* in an aerated system coupled to in situ extraction. *Process Biochemistry*, 65, 11–20. <https://doi.org/10.1016/j.procbio.2017.10.010>
- Sarbu, I., & Csutak, O. (2019). The microbiology of cocoa fermentation. In *Caffeinated and Cocoa Based Beverages: Volume 8. The Science of Beverages*. Elsevier Inc. <https://doi.org/10.1016/B978-0-12-815864-7.00013-1>
- Shimadzu, C. (2020). Gas Chromatography- mass spectrometry (gc-ms). *SpringerReference*. https://doi.org/10.1007/springerreference_30045
- Shirey, R. E. (2012). SPME Commercial Devices and Fibre Coatings. In *Handbook of Solid Phase Microextraction*. Elsevier Inc. <https://doi.org/10.1016/B978-0-12-416017-0.00004-8>
- Shugar, G. J., & Ballinger, J. T. (1996). *Technicians' Ready Reference Handbook, fourth edition* (L. M. Dawkins (ed.); fourth edi). McGraw-Hill, Inc, USA.
- Sirbu, D., Corno, M., Ullrich, M. S., & Kuhnert, N. (2018). Characterization of triacylglycerols in unfermented cocoa beans by HPLC- ESI mass spectrometry. *Food Chemistry*, 254(February), 232–240. <https://doi.org/10.1016/j.foodchem.2018.01.194>
- Sirbu, D., Grimbs, A., Corno, M., Ullrich, M. S., & Kuhnert, N. (2018). *Variation of triacylglycerol profiles in unfermented and dried fermented cocoa beans of different origins*. 111(May), 361–370. <https://doi.org/10.1016/j.foodres.2018.05.025>
- Spietelun, A., Pilarczyk, M., Kloskowski, A., & Namieśnik, J. (2010). Current trends in solid-phase microextraction (SPME) fibre coatings. *Chemical Society Reviews*, 39(11), 4524–4537. <https://doi.org/10.1039/c003335a>
- Stadler, R. H., Hughes, G., & Guillaume-Gentil, O. (2014). Safety of Food and Beverages: Coffee, Tea and Herbals, Cocoa and Derived Products. In *Encyclopedia of Food Safety* (Vol. 3). Elsevier Ltd. <https://doi.org/10.1016/B978-0-12-378612-8.00288-2>

- Taeho Kim, Peter J. Stogios, Anna N. Khusnutdinova, Kayla Nemr, Tatiana Skarina, Robert Flick, Jeong Chan Joo, Radhakrishnan Mahadevan, Alexei Savchenko, A. F. Y. (2020). Rational engineering of 2-deoxyribose-5-phosphate aldolases for the biosynthesis of (R)-1,3-butanediol. *Journal of Biological Chemistry*, Volume 295(2), 597–609. <https://doi.org/https://doi.org/10.1074/jbc.RA119.011363>
- Theopold, P. F., Klaus, R. L. et al. (2020). *Collision Theory*. <https://chem.libretexts.org/@go/page/191379>
- Torres-moreno, M., Tarrega, A., Blanch, C., & Torres-moreno, M. (2021). Effect of cocoa roasting time on volatile composition of dark chocolates from different origins determined by HS-SPME / GC-MS different origins determined by HS-SPME / GC-MS. *CyTA - Journal of Food*, 19(1), 81–95. <https://doi.org/10.1080/19476337.2020.1860137>
- Tran, Q. N. M., Mimoto, H., Koyama, M., & Nakasaki, K. (2019). Lactic acid bacteria modulate organic acid production during early stages of food waste composting. *Science of the Total Environment*, 687, 341–347. <https://doi.org/10.1016/j.scitotenv.2019.06.113>
- UNDP. (2018). Annual Report 2017. In *NTT Docomo* (Vol. 21, Issue 5). [file:///C:/Users/green/Downloads/Yemen HC Annual report 2017.pdf](file:///C:/Users/green/Downloads/Yemen%20HC%20Annual%20report%202017.pdf)
- Utrilla-Vázquez, M., Rodríguez-Campos, J., Avendani-Arazate, C. H., Gschaedler, A., & Lugo-Cervantes, E. (2020). Analysis of volatile compounds of five varieties of Maya cocoa during fermentation and drying processes by Venn diagram and PCA. *Food Research International*, 129, 108834. <https://doi.org/10.1016/j.foodres.2019.108834>
- van Rijswijck, I. M. H., Kruis, A. J., Wolkers – Rooijackers, J. C. M., Abee, T., & Smid, E. J. (2019). Acetate-ester hydrolase activity for screening of the variation in acetate ester yield of *Cyberlindnera fabianii*, *Pichia kudriavzevii* and *Saccharomyces cerevisiae*. *Lwt*, 104(December 2018), 8–15. <https://doi.org/10.1016/j.lwt.2019.01.019>
- Viesser, J. A., de Melo Pereira, G. V., de Carvalho Neto, D. P., Rogez, H., Góes-Neto, A., Azevedo, V., Brenig, B., Aburjaile, F., & Soccol, C. R. (2021). Co-culturing fructophilic lactic acid bacteria and yeast enhanced sugar metabolism and aroma formation during cocoa beans fermentation. *International Journal of Food Microbiology*, 339(September 2020). <https://doi.org/10.1016/j.ijfoodmicro.2020.109015>
- Wardiyah. (2016). *Kimia Organik* (edisi pert). KEMENTRIAN KESEHATAN

REPUBLIK INDONESIA, Pusat Pendidikan Sumber Daya Manusia Kesehatan.

- Yang, C., Wang, R., & Song, H. (2012). The mechanism of peptide bonds cleavage and volatile compounds generated from pentapeptide to heptapeptide via Maillard reaction. *Food Chemistry*, 133(2), 373–382. <https://doi.org/10.1016/j.foodchem.2012.01.044>
- Yi, C., Zhu, H., Yang, R., Bao, J., He, H., & Niu, M. (2020). Links between microbial compositions and volatile profiles of rice noodle fermentation liquid evaluated by 16S rRNA sequencing and GC-MS. *Lwt*, 118, 108774. <https://doi.org/10.1016/j.lwt.2019.108774>
- Youngquist, J. T., Schumacher, M. H., Rose, J. P., Raines, T. C., Politz, M. C., Copeland, M. F., & Pflieger, B. F. (2013). Production of medium chain length fatty alcohols from glucose in *Escherichia coli*. *Metabolic Engineering*, 20, 177–186. <https://doi.org/10.1016/j.ymben.2013.10.006>
- Yu, H., Zhang, R., Yang, F., Xie, Y., Guo, Y., Yao, W., & Zhou, W. (2021). Control strategies of pyrazines generation from Maillard reaction. In *Trends in Food Science and Technology* (Vol. 112, pp. 795–807). Elsevier. <https://doi.org/10.1016/j.tifs.2021.04.028>
- Zackiyah. (2016). Spektrometri Ultra Violet/Sinar Tampak (UV-Vis). *Kimia Analitik Instrumen*, 1–46.
- Zhodu, Y. Y., Li, Y., & Yu, A. N. (2016). The effects of reactants ratios, reaction temperatures and times on Maillard reaction products of the L-ascorbic acid/L-glutamic acid system. *Food Science and Technology*, 36(2), 268–274. <https://doi.org/10.1590/1678-457X.02415>
- Zyzelewicz, D., Krysiak, W., Oracz, J., Sosnowska, D., Budryn, G., & Nebesny, E. (2016). The influence of the roasting process conditions on the polyphenol content in cocoa beans, nibs and chocolates. *Food Research International*, 89, 918–929. <https://doi.org/10.1016/j.foodres.2016.03.026>

LAMPIRAN

Lampiran 1 : Data Selama Proses Penyangraian

Tanggal : : 17 Maret 2021
 Kapasitas mesin penyangrai : 2 kg
 Varietas Biji kakao : Forestero
 Suhu awal penyangraian : 135 °C
 Kadar air biji sebelum penyangraian : 14 %
 Berat biji awal (gram) : 1000 gram

Parameter pengamatan	Waktu penyangraian (menit)											
	0			10			14			18		
	A	B	C	A	B	C	A	B	C	A	B	C
Berat biji setelah di roasting (gram)	Kontrol			858	903	893	852	865	868	835	865	
Suhu biji setelah diroasting (*C)				164	149	145	180	150	162	164	162	
Suhu akhir <i>Roaster</i> (*C)								183	176		188	186

Keterangan :

Tidak dilakukan penyangraian
Suhu tidak dicatat
Penyangraian dianggap <i>over cook</i>

Kode (A) : pengulangan I
 Kode (B) : pengulangan II
 Kode (C) : pengulangan III

Data pada kode (C) untuk penyangraian 10 dan 14 menit, akan digunakan untuk menggantikan data pada kode (A) pada penyangraian 10 dan 14 menit

Lampiran 2 : Keterangan Performa Alat GC-MS yang digunakan

```

===== Analytical Line 1 =====

[GC-2010]
Column Oven Temp.      :60.0 °C
Injection Temp.        :260.00 °C
Injection Mode         :Splitless
Sampling Time          :1.00 min
Flow Control Mode     :Pressure
Pressure               :38.9 kPa
Total Flow             :37.5 mL/min
Column Flow           :0.78 mL/min
Linear Velocity        :32.2 cm/sec
Purge Flow             :3.0 mL/min
Split Ratio            :-1.0
High Pressure Injection :OFF
Carrier Gas Saver      :OFF
Oven Temp. Program
Rate                   Temperature(°C)      Hold Time(min)
-                       60.0                          3.00
5.00                    220.0                         5.00

< Ready Check Heat Unit >
  Column Oven          : Yes
  SPL1                 : Yes
  MS                   : Yes

< Ready Check Detector(FTD) >
< Ready Check Baseline Drift >
< Ready Check Injection Flow >
  SPL1 Carrier         : Yes
  SPL1 Purge           : Yes

< Ready Check APC Flow >
< Ready Check Detector APC Flow >
External Wait          :No
Equilibrium Time      :3.0 min

[GC Program]

[GCMS-QP2010 Plus]
IonSourceTemp         :200.00 °C
Interface Temp.       :200.00 °C
Solvent Cut Time      :1.00 min
Detector Gain Mode    :Relative
Detector Gain         :0.00 kV
Threshold              :1000

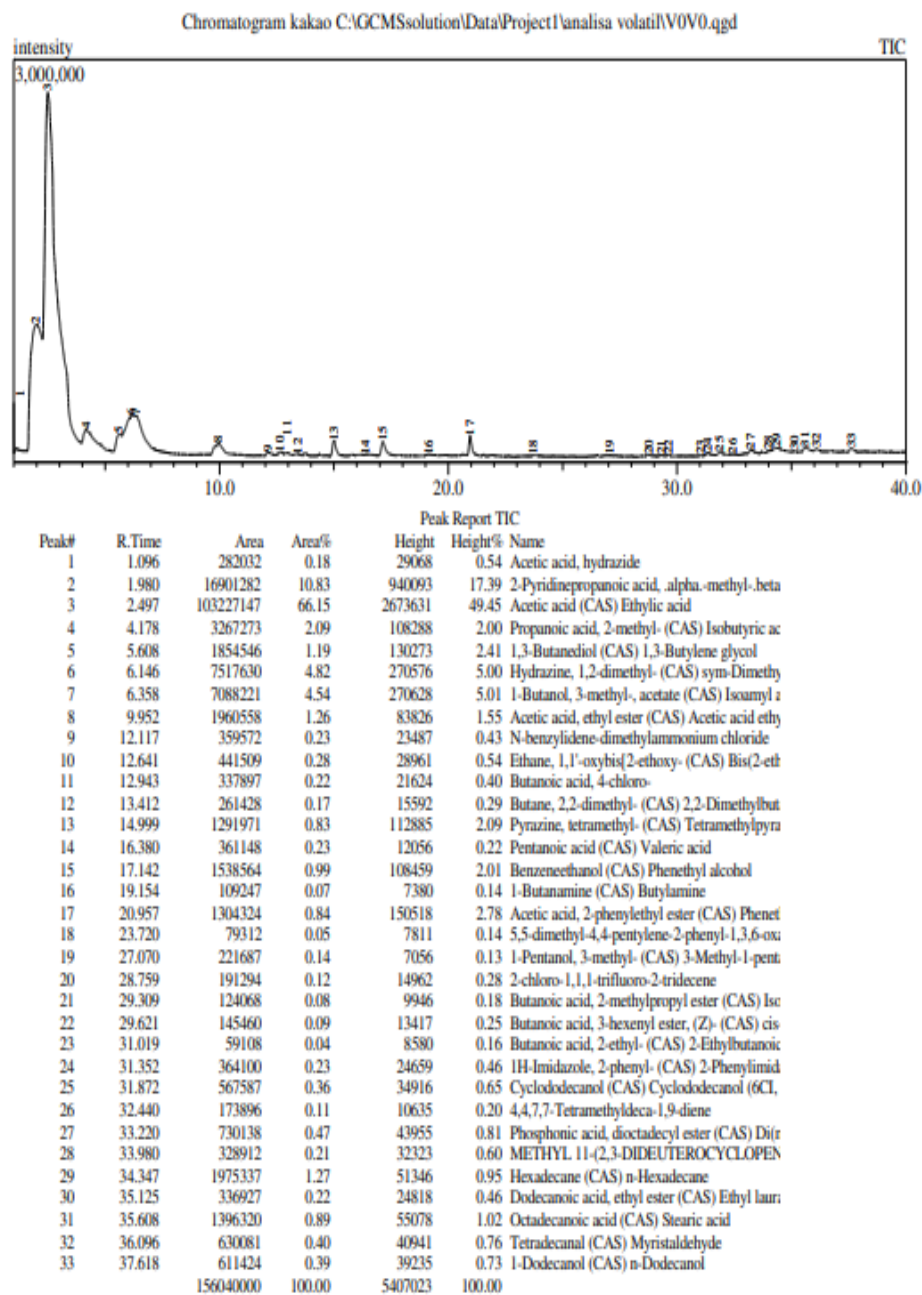
[MS Table]
--Group 1 - Event 1--
Start Time            :1.00min
End Time              :40.00min
ACQ Mode              :Scan
Event Time            :0.60sec
Scan Speed            :555
Start m/z             :30.00
End m/z              :350.00

Sample Inlet Unit     :GC

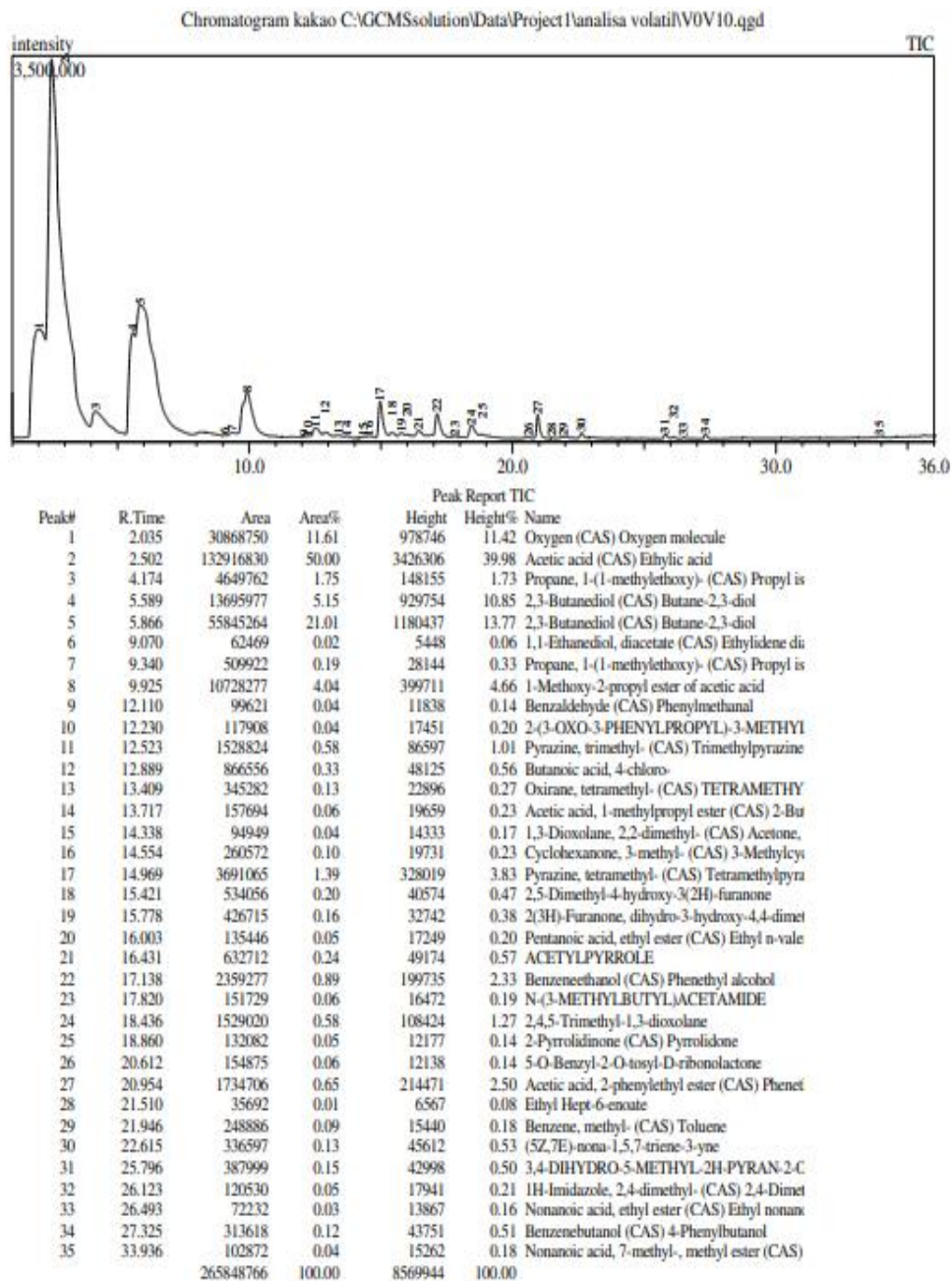
[MS Program]
Use MS Program        :OFF

```

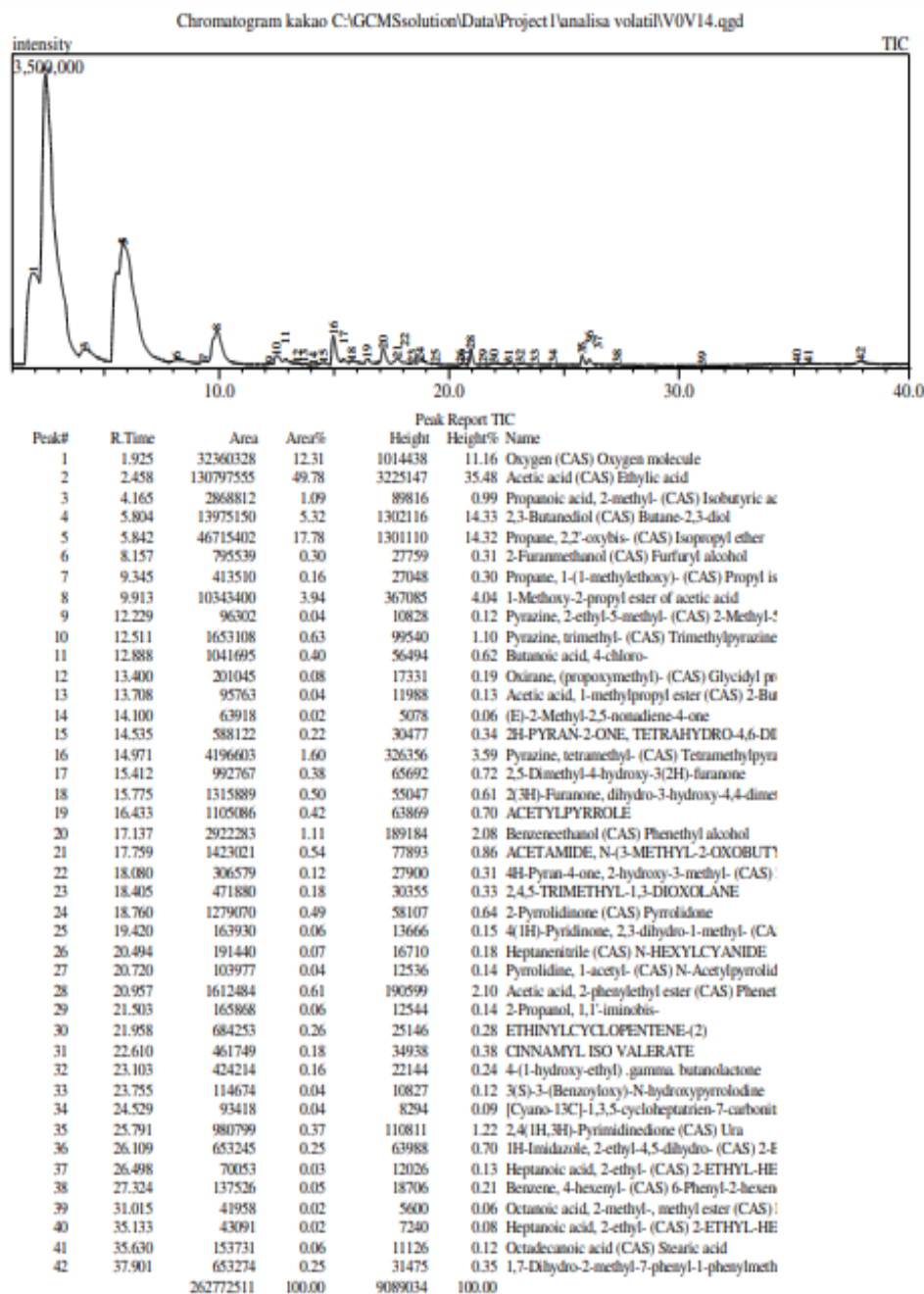
Lampiran 3. Analisa Senyawa Volatil Biji Kakao Fermentasi pada perlakuan penyangraian 0 menit menggunakan metode ekstraksi HS-SPME/GC-MS



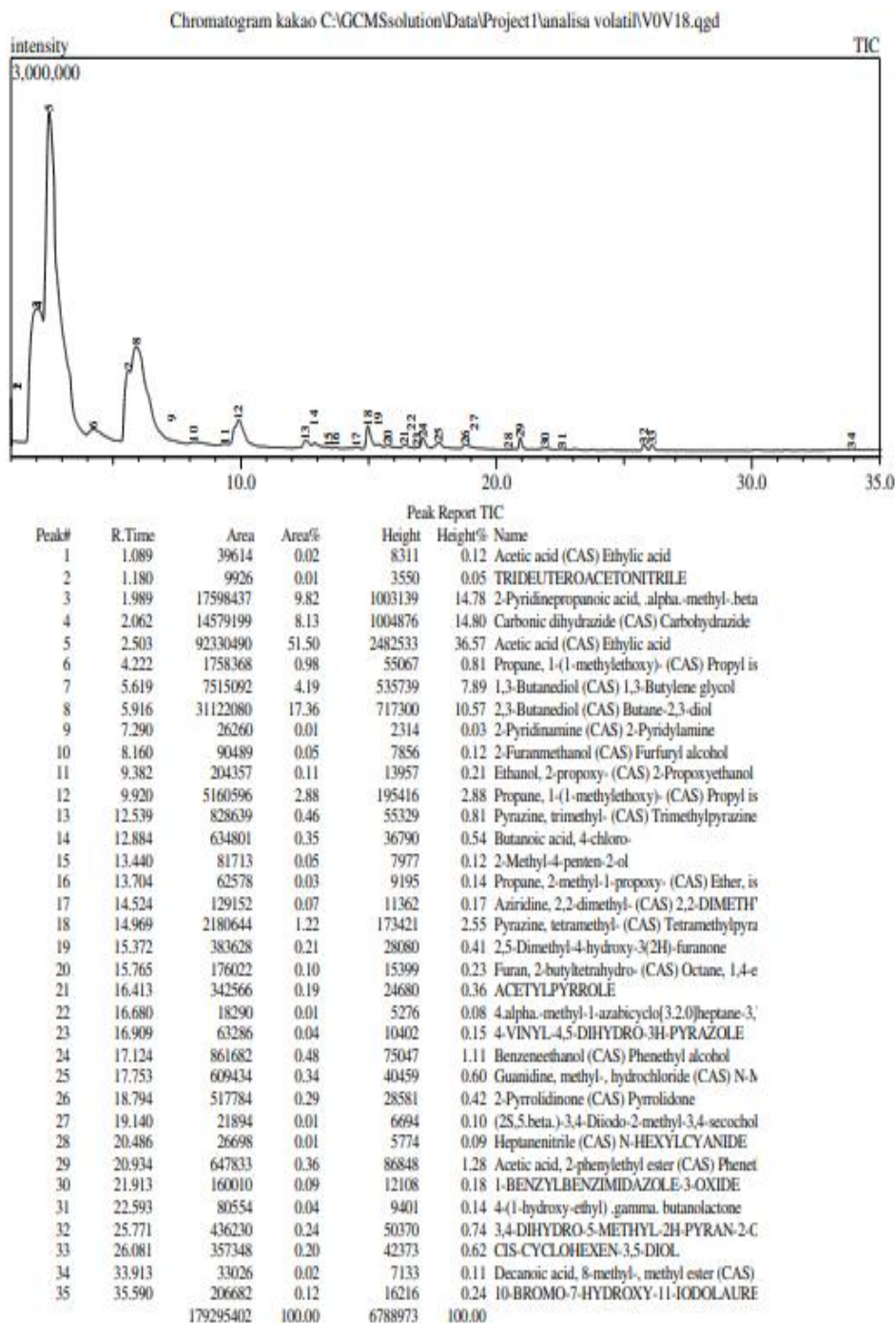
Lampiran 4. Analisa Senyawa Volatil Biji Kakao Fermentasi pada perlakuan penyangraian 10 menit menggunakan metode ekstraksi HS-SPME/GC-MS



Lampiran 5. Analisa Senyawa Volatil Biji Kakao Fermentasi pada perlakuan penyangraian 14 menit menggunakan metode ekstraksi HS-SPME/GC-MS



Lampiran 6. Analisa Senyawa Volatil Biji Kakao Fermentasi pada perlakuan penyangraian 18 menit menggunakan metode ekstraksi HS-SPME/GC-MS.



Lampiran 7 : Hasil uji kadar air biji kakao selama perlakuan penyangraian menggunakan metode oven

Perlakuan	Berat cawan petri kosong (w2)	Berat cawan petri + sampel awal	Berat sampel awal (W)	Berat cawan petri + sampel kering (W1)	$W-(W1-W2)$	$W1-W2$	Kadar air berat basah	Kadar air berat kering	Nilai rata-rata (%)	Keterangan
K ₀ K ₀	83.5973	85.5982	2.0009	85.483	0.1152	1.8857	5.757	6.109	6.109	0 menit penyangraian
K ₁ K ₁₀	77.1357	79.137	2.0013	79.11	0.027	1.9743	1.349	1.368	1.378	10 menit penyangraian
K ₂ K ₁₀	83.467	85.4679	2.0009	85.4405	0.0274	1.9735	1.369	1.388	1.025	14 menit penyangraian
K ₁ K ₁₄	81.2265	83.2261	1.9996	83.2038	0.0223	1.9773	1.115	1.128	1.125	18 menit penyangraian
K ₂ K ₁₄	86.1323	88.1365	2.0042	88.1182	0.0183	1.9859	0.913	0.921	1.125	18 menit penyangraian
K ₁ K ₁₈	94.0515	96.0528	2.0013	96.029	0.0238	1.9775	1.189	1.204	1.125	18 menit penyangraian
K ₂ K ₁₈	96.6536	98.6539	2.0003	98.6332	0.0207	1.9796	1.035	1.046	1.125	18 menit penyangraian

Keterangan:

K₀K₀ : penyangraian 0 menit

K₁K₁₀ : Penyangraian 10 menit ulangan I

K₂K₁₀ : Penyangraian 10 menit ulangan II

K₁K₁₄ : Penyangraian 14 menit ulangan I

K₂K₁₄ : Penyangraian 14 menit ulangan II

K₁K₁₈ : Penyangraian 18 menit ulangan I

K₂K₁₈ : Penyangraian 18 menit ulangan II

Lampiran 8 : Hasil Konversi sampel biji kakao berdasarkan total padatan terlarut

PERLAKUAN	BERAT CAWAN PETRI KOSONG (W2)	BERAT Cawan petri + sampel awal	berat sampel awal (W))	BERAT Cawan petri + sampel KERING (W1)	W-(W1-W2)	W1-W2	KADAR AIR BERAT BASAH	KADAR AIR BERAT KERING	padatan dalam sampel (100-kadar air basis kering)%	Total padatan dalam sampel (berat sampel x padatan) mg
K0K0	83.5973	85.5982	2.0009	85.483	0.1152	1.8857	5.757	6.109	0.9424	474.9827
K1K10	77.1357	79.137	2.0013	79.11	0.027	1.9743	1.349	1.368	0.9865	502.1330
K2K10	83.467	85.4679	2.0009	85.4405	0.0274	1.9735	1.369	1.388	0.9863	496.1120
K1K14	81.2265	83.2261	1.9996	83.2038	0.0223	1.9773	1.115	1.128	0.9888	494.4239
K2K14	86.1323	88.1365	2.0042	88.1182	0.0183	1.9859	0.913	0.921	0.9909	501.3798
K1K18	94.0515	96.0528	2.0013	96.029	0.0238	1.9775	1.189	1.204	0.9881	501.9587
K2K18	96.6536	98.6539	2.0003	98.6332	0.0207	1.9796	1.035	1.046	0.9897	497.7947

K₀K₀ : penyangraian 0 menit

K₁K₁₀ : Penyangraian 10 menit ulangan I

K₂K₁₀ : Penyangraian 10 menit ulangan II

K₁K₁₄ : Penyangraian 14 menit ulangan I

K₂K₁₄ : Penyangraian 14 menit ulangan II

K₁K₁₈ : Penyangraian 18 menit ulangan I

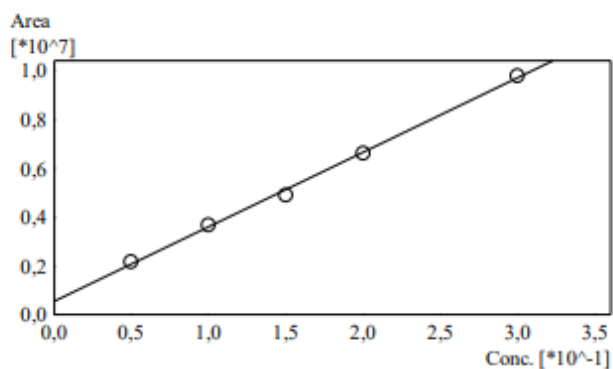
K₂K₁₈ : Penyangraian 18 menit ulangan II

Lampiran 9 : Kurva standar asam amino

Asam Amino asam aspartat

Calibration Curve

ID# : 1 m/z : 134,00
 Name : asam aspartat
 Quantitative Method : External Standard
 Function : $f(x)=3,04814e+007*x+558187$
 Rr1=0,9989208 Rr2=0,9978427
 MeanRF: 3,574048e+007 RF SD: 4,633500e+006 RF %RSD: 12,964293
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	2171193	2171193
2	0,1	3681742	3681742
3	0,15	4907283	4907283
4	0,2	6615933	6615933
5	0,3	9799866	9799866

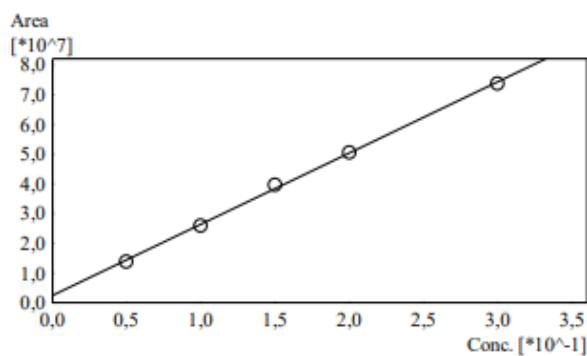
ID#1 Compound Name: asam aspartat

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0.05mM.lcd	4,437	2171193	44834	0.050	0,05
STD 0.1mM.lcd	4,482	3681742	75806	0.100	0,1
STD 0.15mM.lcd	4,459	4907283	103986	0.148	0,15
STD 0.2mM.lcd	4,503	6615933	145313	0.203	0,2
STD 0.3mM.lcd	4,517	9799866	208270	0.303	0,3
A0A0.lcd	4,059	4254054	93657	0,121	--
A1A10.lcd	4,189	5118518	121666	0,150	--
A1A14.lcd	4,252	4065986	78538	0,115	--
A1A18.lcd	4,255	5419136	114746	0,159	--
A2A10.lcd	4,242	4977265	112712	0,145	--
A2A14.lcd	4,255	5434731	111772	0,160	--
A2A18.lcd	4,262	5165395	105616	0,151	--
Average	4,326	5134259	109743	0,150	--
%RSD	3,406	35,730	36,656	40,586	--
Maximum	4,517	9799866	208270	0,303	--
Minimum	4,059	2171193	44834	0,050	--
Standard Deviation	0,147	1834458	40227	0,061	--

Asam Amino Asam Glutamat

Calibration Curve

ID# : 1 m/z : 148,00
 Name : asam glutamat
 Quantitative Method : External Standard
 Function : $f(x)=2.39386e+008*x+2.47526e+006$
 Rr1=0,9993932 Rr2=0,9987867
 MeanRF: 2,602896e+008 RF SD: 1,245939e+007 RF %RSD: 4,786742
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	13931406	13931406
2	0,1	25951981	25951981
3	0,15	39687081	39687081
4	0,2	50603005	50603005
5	0,3	73711402	73711402

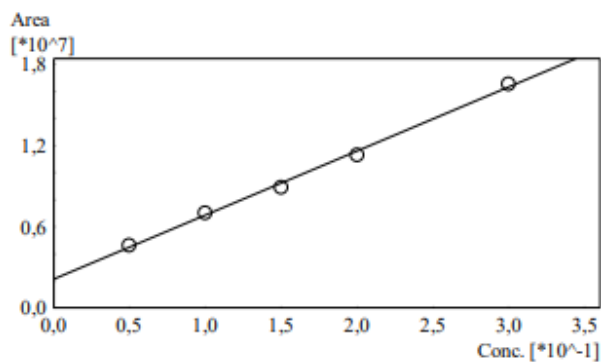
ID#1 Compound Name: asam glutamat

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0.05mM.lcd	4,510	13931406	381576	0,050	0,05
STD 0.1mM.lcd	4,506	25951981	678161	0,100	0,1
STD 0.15mM.lcd	4,507	39687081	1043147	0,151	0,15
STD 0.2mM.lcd	4,530	50603005	1346281	0,198	0,2
STD 0.3mM.lcd	4,525	73711402	1910418	0,298	0,3
A0A0.lcd	3,983	27085659	680750	0,103	--
A1A10.lcd	4,083	39666980	1033673	0,155	--
A1A14.lcd	4,095	25987450	656248	0,098	--
A1A18.lcd	4,107	39228908	1040008	0,154	--
A2A10.lcd	4,112	39950402	1045747	0,157	--
A2A14.lcd	4,121	40113004	1044941	0,157	--
A2A18.lcd	4,128	36998330	947671	0,144	--
Average	4,267	37742967	984052	0,147	
%RSD	5,212	39,441	39,516	41,798	
Maximum	4,530	73711402	1910418	0,298	
Minimum	3,983	13931406	381576	0,050	
Standard Deviation	0,222	14886329	388854	0,061	

Asam Amino Arginin

Calibration Curve

ID# : 1 m/z : 175,00
 Name : arginin
 Quantitative Method : External Standard
 Function : $f(x)=4,73003e+007*x+2,11916e+006$
 Rr1=0,9983726 Rr2=0,9967479
 MeanRF: 6,678926e+007 RF SD: 1,551868e+007 RF %RSD: 23,235293
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	4621444	4621444
2	0,1	7032878	7032878
3	0,15	8919319	8919319
4	0,2	11311043	11311043
5	0,3	16551392	16551392

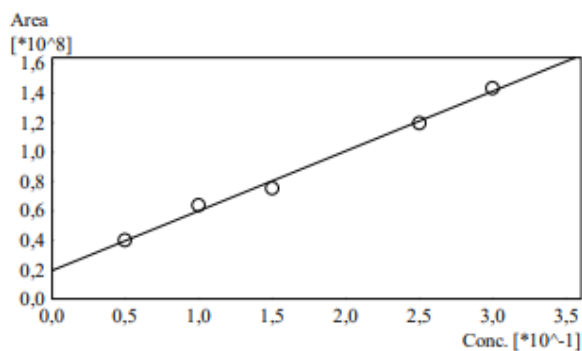
ID#1 Compound Name: arginin

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0.05mM.lcd	3,070	4621444	136074	0,050	0,05
STD 0.1mM.lcd	3,064	7032878	207388	0,100	0,1
STD 0.15mM.lcd	3,064	8919319	266740	0,148	0,15
STD 0.2mM.lcd	3,058	11311043	343022	0,201	0,2
STD 0.3mM.lcd	3,064	16551392	495685	0,305	0,3
A0A0.lcd	3,138	4407860	125972	0,048	--
A1A10.lcd	3,143	5280225	155691	0,067	--
A1A14.lcd	3,129	4197234	117869	0,044	--
A1A18.lcd	3,133	5585534	154244	0,073	--
A2A10.lcd	3,120	5191091	150109	0,065	--
A2A14.lcd	3,128	5416139	151278	0,070	--
A2A18.lcd	3,126	5203288	148611	0,065	--
Average	3,103	6976454	204390	0,103	--
%RSD	1,131	52,523	55,203	76,283	--
Maximum	3,143	16551392	495685	0,305	--
Minimum	3,058	4197234	117869	0,044	--
Standard Deviation	0,035	3664224	112829	0,079	--

Asam Amino Metionin

Calibration Curve

ID# : 1 m/z : 150.00
 Name : metionin
 Quantitative Method : External Standard
 Function : $f(x)=4,06513e+008*x+1,92828e+007$
 Rr1=0,9970133 Rr2=0,9940355
 MeanRF: 5,784580e+008 RF SD: 1,385212e+008 RF %RSD: 23,946629
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	39851434	39851434
2	0,1	63551194	63551194
3	0,15	75439834	75439834
4	0,25	119687648	119687648
5	0,3	143419930	143419930

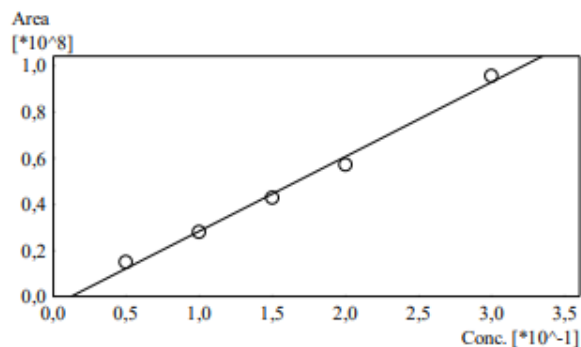
ID#1 Compound Name: metionin

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0,05mM.lcd	4,003	39851434	985994	0,050	0,05
STD 0,1mM.lcd	4,004	63551194	1562420	0,100	0,1
STD 0,15mM.lcd	4,022	75439834	1843420	0,144	0,15
STD 0,2mM.lcd	4,044	119687648	2698157	0,253	0,25
STD 0,3mM.lcd	4,030	143419930	3406434	0,305	0,3
A0A0.lcd	3,612	30199506	764484	0,027	--
A1A10.lcd	3,652	38709427	927192	0,048	--
A1A14.lcd	3,663	26215328	632093	0,017	--
A1A18.lcd	3,662	32987304	782576	0,034	--
A2A10.lcd	3,653	39540236	951900	0,050	--
A2A14.lcd	3,684	39745392	956680	0,050	--
A2A18.lcd	3,691	35660498	852931	0,040	--
Average	3,810	57083978	1363690	0,093	--
%RSD	4,912	66,203	64,022	100,938	--
Maximum	4,044	143419930	3406434	0,305	--
Minimum	3,612	26215328	632093	0,017	--
Standard Deviation	0,187	37791457	873056	0,094	--

Asam Amino

Calibration Curve

ID# : 1 m/z : 147,00
 Name : lisin
 Quantitative Method : External Standard
 Function : $f(x)=3,22858e+008*x-4,04745e+006$
 Rr1=0,9960089 Rr2=0,9920337
 MeanRF: 2,927347e+008 RF SD: 1,604637e+007 RF %RSD: 5,481541
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	14835756	14835756
2	0,1	27849444	27849444
3	0,15	42657228	42657228
4	0,2	57036870	57036870
5	0,3	95669482	95669482

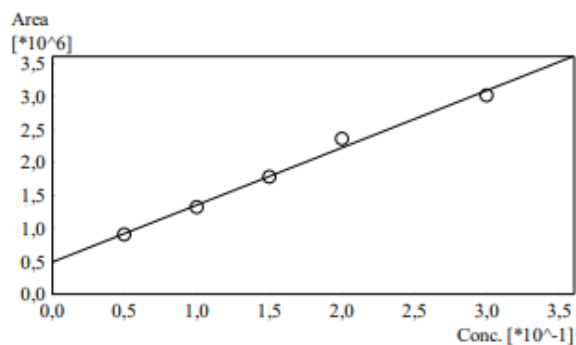
ID#1 Compound Name: lisin

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0,05mM.lcd	3,117	14835756	421982	0,050	0,05
STD 0,1mM.lcd	3,114	27849444	768746	0,100	0,1
STD 0,15mM.lcd	3,114	42657228	1240356	0,151	0,15
STD 0,2mM.lcd	3,108	57036870	1649740	0,201	0,2
STD 0,3mM.lcd	3,125	95669482	2618296	0,309	0,3
A0A0.lcd	3,155	4156350	119389	0,025	--
A1A10.lcd	3,170	4340616	123915	0,026	--
A1A14.lcd	3,161	2786184	82649	0,021	--
A1A18.lcd	3,169	3369127	97588	0,023	--
A2A10.lcd	3,152	4671391	131679	0,027	--
A2A14.lcd	3,161	3821860	109787	0,024	--
A2A18.lcd	3,162	3623017	104713	0,024	--
Average	3,142	22068110	622403	0,082	--
%RSD	0,778	133,042	131,086	113,608	--
Maximum	3,170	95669482	2618296	0,309	--
Minimum	3,108	2786184	82649	0,021	--
Standard Deviation	0,024	29359761	815884	0,093	--

Asam Amino Alanin

Calibration Curve

ID# : 1 m/z : 90,00
 Name : alanin
 Quantitative Method : External Standard
 Function : $f(x)=8,66012e+006*x+487993$
 Rr1=0,9955172 Rr2=0,9910545
 MeanRF: 1,297463e+007 RF SD: 3,023337e+006 RF %RSD: 23,301909
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	900308	900308
2	0,1	1316953	1316953
3	0,15	1781055	1781055
4	0,2	2354778	2354778
5	0,3	3014964	3014964

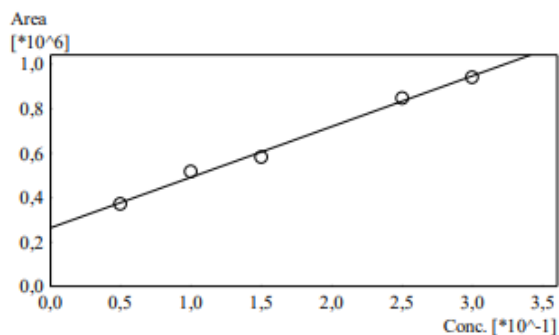
ID#1 Compound Name: alanin

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0,05mM.lcd	3,998	900308	20617	0,050	0,05
STD 0,1mM.lcd	4,021	1316953	29354	0,100	0,1
STD 0,15mM.lcd	4,030	1781055	38598	0,151	0,15
STD 0,2mM.lcd	4,054	2354778	48768	0,204	0,2
STD 0,3mM.lcd	4,047	3014964	55809	0,292	0,3
A0A0.lcd	3,648	1524867	35873	0,120	--
A1A10.lcd	3,699	1837588	46331	0,156	--
A1A14.lcd	3,701	1492202	33424	0,116	--
A1A18.lcd	3,708	1927837	47551	0,166	--
A2A10.lcd	3,702	2204680	49111	0,198	--
A2A14.lcd	3,725	1876553	46372	0,160	--
A2A18.lcd	3,733	1920190	42784	0,165	--
Average	3,839	1845998	41216	0,157	
%RSD	4,439	29,099	24,139	38,558	
Maximum	4,054	3014964	55809	0,292	
Minimum	3,648	900308	20617	0,050	
Standard Deviation	0,170	537158	9949	0,060	

Asam Amino Glisin

Calibration Curve

ID# : 1 m/z : 76,00
 Name : glisin
 Quantitative Method : External Standard
 Function : $f(x)=2,26997e+006*x+264948$
 Rr1=0,9966509 Rr2=0,9933129
 MeanRF: 4,592117e+006 RF SD: 1,752588e+006 RF %RSD: 38,165147
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	369650	369650
2	0,1	517397	517397
3	0,15	581811	581811
4	0,25	845511	845511
5	0,3	939846	939846

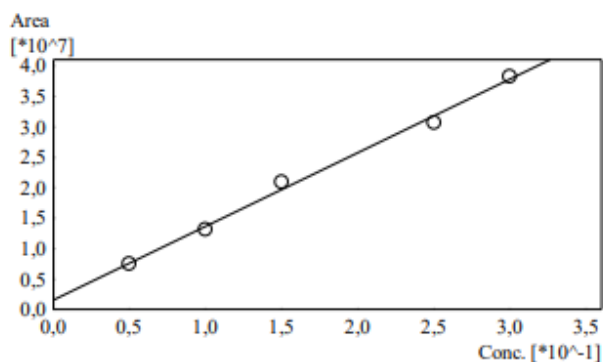
ID#1 Compound Name: glisin

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0,05mM.lcd	4,092	369650	7302	0,050	0,05
STD 0,1mM.lcd	4,128	517397	9706	0,100	0,1
STD 0,15mM.lcd	4,134	581811	10663	0,143	0,15
STD 0,25mM.lcd	4,181	845511	15090	0,253	0,25
STD 0,3mM.lcd	4,156	939846	15599	0,297	0,3
A0A0.lcd	3,707	553813	11919	0,127	--
A1A10.lcd	3,763	633338	14818	0,162	--
A1A14.lcd	3,773	497527	10715	0,102	--
A1A18.lcd	3,774	759051	16186	0,218	--
A2A10.lcd	3,765	729574	15377	0,205	--
A2A14.lcd	3,785	719235	15621	0,200	--
A2A18.lcd	3,792	671818	13920	0,179	--
Average	3,921	651548	13076	0,170	--
%RSD	4,952	24,457	22,242	41,264	--
Maximum	4,181	939846	16186	0,297	--
Minimum	3,707	369650	7302	0,050	--
Standard Deviation	0,194	159347	2908	0,070	--

Asam Amino Prolin

Calibration Curve

ID# : 1 m/z : 116,00
 Name : proline
 Quantitative Method : External Standard
 Function : $f(x)=1,20579e+008*x+1,62622e+006$
 Rr1=0,9973338 Rr2=0,9946747
 MeanRF: 1,344322e+008 RF SD: 1,083478e+007 RF %RSD: 8,059657
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	7506666	7506666
2	0,1	13178706	13178706
3	0,15	20999537	20999537
4	0,25	30675326	30675326
5	0,3	38262757	38262757

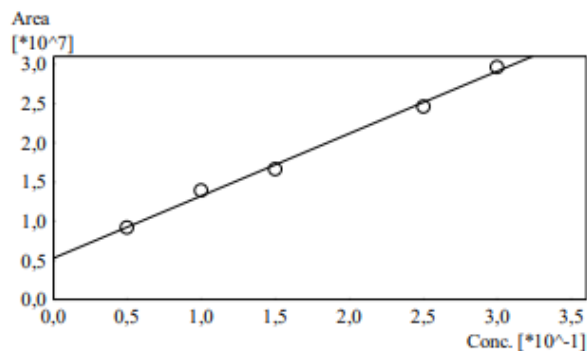
ID#1 Compound Name: proline

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0,05mM.lcd	4,493	7506666	182863	0,050	0,05
STD 0,1mM.lcd	4,505	13178706	335259	0,100	0,1
STD 0,15mM.lcd	4,506	20999537	522610	0,153	0,15
STD 0,25mM.lcd	4,503	30675326	786235	0,245	0,25
STD 0,3mM.lcd	4,523	38262757	952814	0,304	0,3
A0A0.lcd	4,040	10958952	275249	0,077	--
A1A10.lcd	4,128	15840790	402955	0,118	--
A1A14.lcd	4,153	10273636	252581	0,072	--
A1A18.lcd	4,163	15905750	400883	0,118	--
A2A10.lcd	4,160	16262526	405255	0,121	--
A2A14.lcd	4,166	15722020	389100	0,117	--
A2A18.lcd	4,178	14786584	358610	0,109	--
Average	4,293	17531104	438701	0,132	--
%RSD	4,455	50,078	50,614	55,152	--
Maximum	4,523	38262757	952814	0,304	--
Minimum	4,040	7506666	182863	0,050	--
Standard Deviation	0,191	8779171	222045	0,073	--

Asam Amino Valin

Calibration Curve

ID# : 1 m/z : 118,00
 Name : valine
 Quantitative Method : External Standard
 Function : $f(x)=7,94221e+007*x+5,29649e+006$
 Rr1=0,9973316 Rr2=0,9946703
 MeanRF: 1,261416e+008 RF SD: 3,593016e+007 RF %RSD: 28,483980
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	9152372	9152372
2	0,1	13949437	13949437
3	0,15	16643856	16643856
4	0,25	24583068	24583068
5	0,3	29662515	29662515

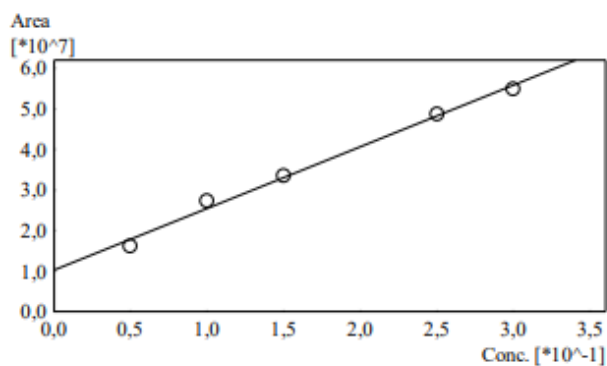
ID#1 Compound Name: valine

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0.05mM.lcd	3,932	9152372	217962	0,050	0,05
STD 0.1mM.lcd	3,938	13949437	330949	0,100	0,1
STD 0.15mM.lcd	3,950	16643856	390213	0,145	0,15
STD 0.25mM.lcd	3,957	24583068	571891	0,250	0,25
STD 0.3mM.lcd	3,941	29662515	730546	0,307	0,3
A0A0.lcd	3,565	8903550	197709	0,045	--
A1A10.lcd	3,623	9461053	222778	0,052	--
A1A14.lcd	3,630	6779055	147457	0,019	--
A1A18.lcd	3,632	9335245	192133	0,051	--
A2A10.lcd	3,626	9774015	220417	0,056	--
A2A14.lcd	3,648	9600940	217208	0,054	--
A2A18.lcd	3,658	9170619	192423	0,049	--
Average	3,758	13084644	302641	0,098	
%RSD	4,393	54,470	59,035	92,440	
Maximum	3,957	29662515	730546	0,307	
Minimum	3,565	6779055	147457	0,019	
Standard Deviation	0,165	7127186	178664	0,091	

Asam Amino Leusin-Isoleusin

Calibration Curve

ID# : 1 m/z : 132,00
 Name : leusin-isoleusin
 Quantitative Method : External Standard
 Function : $f(x)=1,51563e+008*x+1,03735e+007$
 Rr1=0,9960071 Rr2=0,9920302
 MeanRF: 2,395674e+008 RF SD: 5,804916e+007 RF %RSD: 24,230828
 FitType : Linear
 ZeroThrough : Not Through
 Weighted Regression : None



#	Conc.(Ratio)	MeanArea	Area
1	0,05	16128193	16128193
2	0,1	27356012	27356012
3	0,15	33554271	33554271
4	0,25	48738183	48738183
5	0,3	54919538	54919538

ID#1 Compound Name: leusin-isoleusin

Title	Ret. Time	Area	Height	Conc.	Std. Conc.
STD 0,05mM.lcd	3,898	16128193	409703	0,050	0,05
STD 0,1mM.lcd	3,894	27356012	641399	0,100	0,1
STD 0,15mM.lcd	3,900	33554271	793618	0,145	0,15
STD 0,25mM.lcd	3,922	48738183	1199268	0,247	0,25
STD 0,3mM.lcd	3,928	54919538	1371592	0,294	0,3
A0A0.lcd	3,574	16357438	397007	0,039	--
A1A10.lcd	3,611	21154535	479598	0,071	--
A1A14.lcd	3,612	14113846	313639	0,025	--
A1A18.lcd	3,619	18218900	400170	0,052	--
A2A10.lcd	3,611	20149146	468864	0,064	--
A2A14.lcd	3,636	19341706	447634	0,059	--
A2A18.lcd	3,637	17641968	401916	0,048	--
Average	3,737	25639478	610367	0,100	--
%RSD	4,082	52,259	56,016	86,627	--
Maximum	3,928	54919538	1371592	0,294	--
Minimum	3,574	14113846	313639	0,025	--
Standard Deviation	0,153	13399013	341905	0,086	--

Lampiran : Hasil analisa asam amino pada sampel biji kakao berdasarkan total padatan

Asam Amino Asam Aspartat

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Aspartic acid (mM)	Aspartic acid (ppm)	Aspartic acid (%)	Kadar air berat basah	Massa Aspartic acid (mg)	Aspartic acid (%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.1210	16.106	0.799	5.757	4.02658	0.8477	0.8477
2	A1A10	509	25	0.1500	19.967	0.981	1.349	4.99163	0.9941	0.9833
3	A2A10	503	25	0.1450	19.301	0.959	1.369	4.82524	0.9726	
4	A1A14	500	25	0.1150	15.308	0.765	1.115	3.82691	0.7740	0.9180
5	A2A14	506	25	0.1600	21.298	1.052	0.913	5.3244	1.0619	
6	A1A18	508	25	0.1590	21.164	1.042	1.189	5.29112	1.0541	1.0318
7	A2A18	503	25	0.1510	20.100	0.999	1.035	5.0249	1.0094	

Asam Amino Asam Glutamat

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Glutamic acid (mM)	Glutamic acid (ppm)	Glutamic acid (%)	Kadar air berat basah	Massa Glutamic acid (mg)	Glutamic acid (%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.1030	15.154	0.752	5.757	3.7886	0.7976	0.7976
2	A1A10	509	25	0.1550	22.805	1.120	1.349	5.70129	1.1354	
3	A2A10	503	25	0.1570	23.099	1.148	1.369	5.77485	1.1640	1.1497
4	A1A14	500	25	0.0980	14.419	0.721	1.115	3.60469	0.7291	
5	A2A14	506	25	0.1570	23.099	1.141	0.913	5.77485	1.1518	0.9404
6	A1A18	508	25	0.1540	22.658	1.115	1.189	5.66451	1.1285	
7	A2A18	503	25	0.1440	21.187	1.053	1.035	5.29668	1.0640	1.0963

Asam Amino Arginin

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Arginine (mM)	Arginine (ppm)	Arginine (%)	Kadar air berat basah	Massa Arginine (mg)	Arginine (%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.0480	8.362	0.415	5.757	2.0904	0.4401	0.4401
2	A1A10	509	25	0.0670	11.671	0.573	1.349	2.91785	0.5811	0.5758
3	A2A10	503	25	0.0650	11.323	0.563	1.369	2.83075	0.5706	
4	A1A14	500	25	0.0440	7.665	0.383	1.115	1.9162	0.3876	0.4978
5	A2A14	506	25	0.0700	12.194	0.602	0.913	3.0485	0.6080	
6	A1A18	508	25	0.0730	12.717	0.626	1.189	3.17915	0.6333	0.6010
7	A2A18	503	25	0.0650	11.323	0.563	1.035	2.83075	0.5687	

Asam Amino Metionin

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Methionine (mM)	Methionine (ppm)	Methionine (%)	Kadar air berat basah	Massa Methionine (mg)	Methionine (%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.0270	4.029	0.200	5.757	1.00717	0.2120	0.2120
2	A1A10	509	25	0.0480	7.162	0.352	1.349	1.79052	0.3566	0.3663
3	A2A10	503	25	0.0500	7.461	0.371	1.369	1.86513	0.3759	
4	A1A14	500	25	0.0170	2.537	0.127	1.115	0.63414	0.1283	0.2501
5	A2A14	506	25	0.0500	7.461	0.369	0.913	1.86513	0.3720	
6	A1A18	508	25	0.0340	5.073	0.250	1.189	1.26829	0.2527	0.2762
7	A2A18	503	25	0.0400	5.968	0.297	1.035	1.4921	0.2997	

Asam Amino Lisin

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Lysine (mM)	Lysine (ppm)	Lysine (%)	Kadar air berat basah	Massa Lysine (mg)	Lysine (%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.0250	3.655	0.181	5.757	0.91369	0.1924	0.1924
2	A1A10	509	25	0.0260	3.801	0.187	1.349	0.95024	0.1892	0.1941
3	A2A10	503	25	0.0270	3.947	0.196	1.369	0.98678	0.1989	
4	A1A14	500	25	0.0210	3.070	0.153	1.115	0.7675	0.1552	0.1651
5	A2A14	506	25	0.0240	3.509	0.173	0.913	0.87714	0.1749	
6	A1A18	508	25	0.0230	3.362	0.165	1.189	0.84059	0.1675	0.1718
7	A2A18	503	25	0.0240	3.509	0.174	1.035	0.87714	0.1762	

Asam Amino Alanin

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Alanine (mM)	Alanine (ppm)	Alanine (%)	Kadar air berat basah	Massa Alanin (mg)	Alanine (%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.1200	10.691	0.530	5.757	2.6727	0.5627	0.5627
2	A1A10	509	25	0.1560	13.898	0.683	1.349	3.4745	0.6920	
3	A2A10	503	25	0.1980	17.640	0.877	1.369	4.4100	0.8889	0.7904
4	A1A14	500	25	0.1160	10.334	0.517	1.115	2.5836	0.5225	0.6167
5	A2A14	506	25	0.1600	14.254	0.704	0.913	3.5636	0.7108	
6	A1A18	508	25	0.1660	14.789	0.728	1.189	3.6972	0.7366	0.7374
7	A2A18	503	25	0.1650	14.700	0.731	1.035	3.6750	0.7382	

Asam Amino Glisin

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Glycine (mM)	Glycine (ppm)	Glycine (%)	Kadar air berat basah	Massa Glycine (mg)	Glycine (%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.1270	9.534	0.473	5.757	2.38347	0.5018	0.5018
2	A1A10	509	25	0.1620	12.161	0.597	1.349	3.04034	0.6055	0.6905
3	A2A10	503	25	0.2050	15.389	0.765	1.369	3.84734	0.7755	
4	A1A14	500	25	0.1020	7.657	0.383	1.115	1.91429	0.3872	0.5679
5	A2A14	506	25	0.2000	15.014	0.742	0.913	3.7535	0.7486	
6	A1A18	508	25	0.2180	16.365	0.805	1.189	4.09132	0.8151	0.7450
7	A2A18	503	25	0.1790	13.438	0.668	1.035	3.35938	0.6749	

Asam Amino Prolin

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Proline (mM)	Proline (ppm)	Proline (%)	Kadar air berat basah	Massa Proline (mg)	Proline(%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.0770	8.865	0.440	5.757	2.21625	0.4666	0.4666
2	A1A10	509	25	0.1180	13.585	0.667	1.349	3.39634	0.6764	0.6892
3	A2A10	503	25	0.1210	13.931	0.692	1.369	3.48268	0.7020	
4	A1A14	500	25	0.0720	8.289	0.414	1.115	2.07234	0.4191	0.5454
5	A2A14	506	25	0.1170	13.470	0.666	0.913	3.36755	0.6717	
6	A1A18	508	25	0.1180	13.585	0.669	1.189	3.39634	0.6766	0.6534
7	A2A18	503	25	0.1090	12.549	0.624	1.035	3.13729	0.6302	

Asam Amino Valin

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Valine (mM)	Valine (ppm)	Valine (%)	Kadar air berat basah	Massa Valine (mg)	Valine(%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.0450	5.272	0.261	5.757	1.31795	0.2775	0.2775
2	A1A10	509	25	0.0520	6.092	0.299	1.349	1.52296	0.3033	0.3169
3	A2A10	503	25	0.0560	6.560	0.326	1.369	1.64011	0.3306	
4	A1A14	500	25	0.0190	2.226	0.111	1.115	0.55647	0.1125	0.2140
5	A2A14	506	25	0.0540	6.326	0.313	0.913	1.58154	0.3154	
6	A1A18	508	25	0.0510	5.975	0.294	1.189	1.49368	0.2976	0.2929
7	A2A18	503	25	0.0490	5.740	0.285	1.035	1.4351	0.2883	

Asam Amino Leusin-isoleusin

No	Sampel	Berat Sampel (mg)	Volume Ekstrak (ml)	Leucine-Isoleucine (mM)	Leucine-Isoleucine (ppm)	Leucine-Isoleucine (%)	Kadar air berat basah	Massa Leucine-Isoleucine (mg)	Leucine-Isoleucine(%) berdasarkan total padatan	Rerata
1	A0A0	504	25	0.0390	5.116	0.254	5.757	1.27891	0.2693	0.2693
2	A1A10	509	25	0.0710	9.313	0.457	1.349	2.32827	0.4637	0.4434
3	A2A10	503	25	0.0640	8.395	0.417	1.369	2.09872	0.4230	0.2758
4	A1A14	500	25	0.0250	3.279	0.164	1.115	0.81981	0.1658	0.2758
5	A2A14	506	25	0.0590	7.739	0.382	0.913	1.93476	0.3859	0.3280
6	A1A18	508	25	0.0520	6.821	0.336	1.189	1.70521	0.3397	0.3280
7	A2A18	503	25	0.0480	6.296	0.313	1.035	1.57404	0.3162	0.3280

Keterangan:

A₀A₀ : penyangraian 0 menit

A₁A₁₀ : Penyangraian 10 menit ulangan I

A₂A₁₀ : Penyangraian 10 menit ulangan II

A₁A₁₄ : Penyangraian 14 menit ulangan I

A₂A₁₄ : Penyangraian 14 menit ulangan II

A₁A₁₈ : Penyangraian 18 menit ulangan I

A₂A₁₈ : Penyangraian 18 menit ulangan II

Lampiran 11 : Data hasil perhitungan asam amino hidrofobik, asam amino hidrofilik dan total asam amino

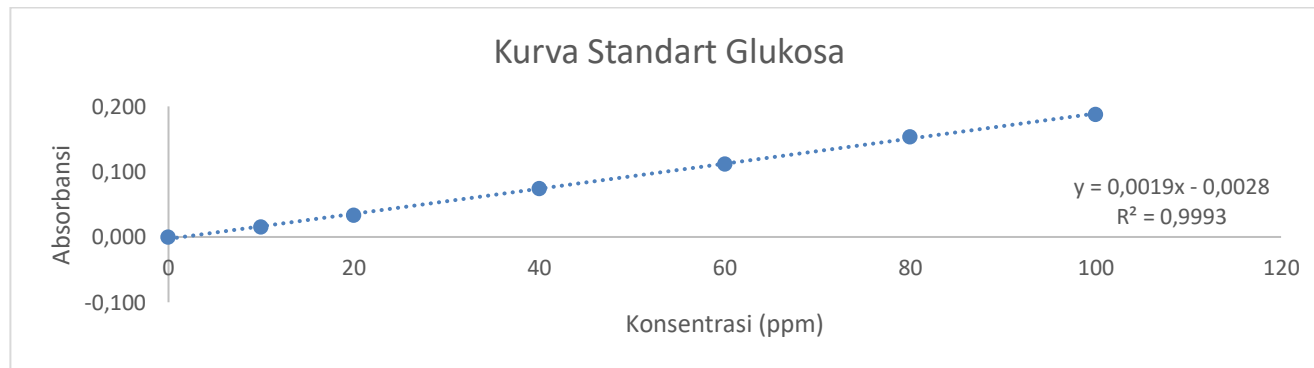
Asam Amino Hidrofilik	Kandungan asam amino berdasarkan total padatan (%)			
	0 menit	10 menit	14 menit	18 mneit
Asam aspartat	0.85	0.98	0.92	1.01
Asam glutamat	0.80	1.15	0.94	1.10
Arginin	0.44	0.58	0.50	0.60
Metionin	0.21	0.37	0.25	0.28
Lisin	0.19	0.19	0.17	0.17
total hidrofilik	2.49	3.27	2.77	3.16

Asam Amino Hidrofobik	Kandungan asam amino berdasarkan total padatan (%)			
	0 menit	10 menit	14 menit	18 mneit
Alanin	0.56	0.79	0.62	0.74
Glisin	0.50	0.69	0.57	0.75
Prolin	0.47	0.69	0.55	0.65
Valin	0.28	0.32	0.21	0.29
Leusin-Isoleusin	0.27	0.44	0.28	0.33
Totak Hidrofobik	2.08	2.93	2.22	2.76

Variabel X/Y	Kandungan asam amino berdasarkan total padatan (%)			
	0 menit	10 menit	14 menit	18 mneit
Total Asam Amino	4.57	6.2	4.99	5.92
Total Hidrofilik	2.49	3.27	2.77	3.16
Total Hidrofobik	2.08	2.93	2.22	2.76

Lampiran 12: Hasil pengujian gula reduksi menggunakan Spektrofotometer UV-Vis

Konsentrasi ppm	Abs
0	0.000
10	0.015
20	0.033
40	0.074
60	0.112
80	0.153
100	0.188



Sampel	Kode	Berat (mg)	ml Ekstrak	FP	Absorbansi	ppm Kurva	% Karbohidrat	Kadar air berat basah	padatan dalam sampel (100-kadar air basis kering)%	Total padatan dalam sampel (berat sampel x padatan) mg	massa gula reduksi (mg)	% Karbohidrat (total padatan)	Rata-Rata nilai % Karbohidrat (total padatan)	
G ₀ G ₀	A	1	2004.0	250	1	0.141	75.684	0.944	5.7574	0.9424	1888.6217	18.921053	1.0018	0.9757
			2004.0	250	1	0.140	75.158	0.938	5.7574	0.9424	1888.6217	18.789474	0.9949	
		2	2004.0	250	1	0.135	72.526	0.905	5.7574	0.9424	1888.6217	18.131579	0.9600	
			2004.0	250	1	0.133	71.474	0.892	5.7574	0.9424	1888.6217	17.868421	0.9461	
G ₁ G ₁₀	B	1	2002.0	250	1	0.097	52.526	0.656	1.3491	0.9865	1974.9910	13.131579	0.6649	0.6985
			2002.0	250	1	0.099	53.579	0.669	1.3491	0.9865	1974.9910	13.394737	0.6782	
		2	2011.0	250	1	0.103	55.684	0.692	1.3491	0.9865	1983.8696	13.921053	0.7017	
			2011.0	250	1	0.103	55.684	0.692	1.3491	0.9865	1983.8696	13.921053	0.7017	
G ₂ G ₁₀	E	1	2007.0	250	1	0.101	54.632	0.681	1.3694	0.9863	1979.5161	13.657895	0.6900	0.6985
			2007.0	250	1	0.100	54.105	0.674	1.3694	0.9863	1979.5161	13.526316	0.6833	
		2	2005.0	250	1	0.108	58.316	0.727	1.3694	0.9863	1977.5435	14.578947	0.7372	
			2005.0	250	1	0.107	57.789	0.721	1.3694	0.9863	1977.5435	14.447368	0.7306	
G ₁ G ₁₄	C	1	2021.0	250	1	0.144	77.263	0.956	1.1152	0.9888	1998.4618	19.315789	0.9665	0.5917
			2021.0	250	1	0.141	75.684	0.936	1.1152	0.9888	1998.4618	18.921053	0.9468	
		2	2003.0	250	1	0.139	74.632	0.931	1.1152	0.9888	1980.6625	18.657895	0.9420	
			2003.0	250	1	0.140	75.158	0.938	1.1152	0.9888	1980.6625	18.789474	0.9486	
G ₂ G ₁₄	F	1	2007.0	250	1	0.030	17.263	0.215	0.9131	0.9909	1988.6741	4.3157895	0.2170	0.5917
			2007.0	250	1	0.030	17.263	0.215	0.9131	0.9909	1988.6741	4.3157895	0.2170	
		2	2027.0	250	1	0.035	19.895	0.245	0.9131	0.9909	2008.4915	4.9736842	0.2476	

			2027.0	250	1	0.035	19.895	0.245	0.9131	0.9909	2008.4915	4.9736842	0.2476	
G ₁ G ₁₈	D	1	2010.0	250	1	0.045	25.158	0.313	1.1892	0.9881	1986.0971	6.2894737	0.3167	0.2739
			2010.0	250	1	0.045	25.158	0.313	1.1892	0.9881	1986.0971	6.2894737	0.3167	
		2	2047.0	250	1	0.042	23.579	0.288	1.1892	0.9881	2022.6571	5.8947368	0.2914	
			2047.0	250	1	0.044	24.632	0.301	1.1892	0.9881	2022.6571	6.1578947	0.3044	
G ₂ G ₁₈	G	1	2003.0	250	1	0.035	19.895	0.248	1.0348	0.9897	1982.2730	4.9736842	0.2509	
			2003.0	250	1	0.036	20.421	0.255	1.0348	0.9897	1982.2730	5.1052632	0.2575	
		2	2069.0	250	1	0.032	18.316	0.221	1.0348	0.9897	2047.5900	4.5789474	0.2236	
			2069.0	250	1	0.033	18.842	0.228	1.0348	0.9897	2047.5900	4.7105263	0.2301	

Keterangan :

G₀G₀ : penyangraian 0 menit

G₁G₁₀ : Penyangraian 10 menit ulangan I

G₂G₁₀ : Penyangraian 10 menit ulangan II

G₁G₁₄ : Penyangraian 14 menit ulangan I

G₂G₁₄ : Penyangraian 14 menit ulangan II

G₁G₁₈ : Penyangraian 18 menit ulangan I

G₂G₁₈ : Penyangraian 18 menit ulangan II

Lampiran 13 : Hasil Uji Anova pada sampel asam amino dan gula reduksi

ANOVA

		Sum of Squares	df	Mean Square	F	Sig.
Alanine	Between Groups	.066	3	.022	2.387	.210
	Within Groups	.037	4	.009		
	Total	.104	7			
as_aspartat	Between Groups	.038	3	.013	1.200	.417
	Within Groups	.043	4	.011		
	Total	.081	7			
Glisin	Between Groups	.074	3	.025	1.105	.445
	Within Groups	.090	4	.022		
	Total	.164	7			
as_glutamat	Between Groups	.152	3	.051	2.211	.229
	Within Groups	.092	4	.023		
	Total	.244	7			
leusin_isoleusin	Between Groups	.039	3	.013	2.050	.250
	Within Groups	.025	4	.006		
	Total	.064	7			
Lisin	Between Groups	.001	3	.000	6.102	.057
	Within Groups	.000	4	.000		
	Total	.002	7			

Metionin	Between Groups	.026	3	.009	1.111	.443
	Within Groups	.031	4	.008		
	Total	.057	7			
Prolin	Between Groups	.062	3	.021	2.487	.200
	Within Groups	.033	4	.008		
	Total	.095	7			
Valin	Between Groups	.012	3	.004	.738	.582
	Within Groups	.021	4	.005		
	Total	.033	7			
Ariginin	Between Groups	.033	3	.011	1.640	.315
	Within Groups	.026	4	.007		
	Total	.059	7			
gula_reduksi	Between Groups	1.488	3	.496	11.327	.000
	Within Groups	1.051	24	.044		
	Total	2.539	27			

Lampiran 14: Uji lanjut Tukey Test untuk Gula reduksi

gula_reduksiTukey HSD^{a,b}

Perlakuan	N	Subset for alpha = 0.05		
		1	2	3
penyangraian 18 menit	8	.273913		
penyangraian 14 menit	8	.591637	.591637	
penyangraian 10 menit	8		.698450	.698450
penyangraian 0 menit	4			.975700
Sig.		.055	.798	.110

Means for groups in homogeneous subsets are displayed.

a. Uses Harmonic Mean Sample Size = 6.400.

b. The group sizes are unequal. The harmonic mean of the group sizes is used. Type I error levels are not guaranteed.

Lampiran 15: Dokumentasi-dokumentasi selama penelitian

1. Persiapan sampel biji kakao



2. Penyangraian biji Kakao



3. Biji Kakao yang telah disangrai dengan beberapa perlakuan waktu penyangraian



4. Pemisahan Kulit dengan Nib biji Kakao sekaligus penimbangan untuk keperluan analisa sampel



Lampiran 16 : Dokumentasi Pengujian Senyawa Volatil Dengan Alat GCMS-QP 2010 Plus Shimadzu

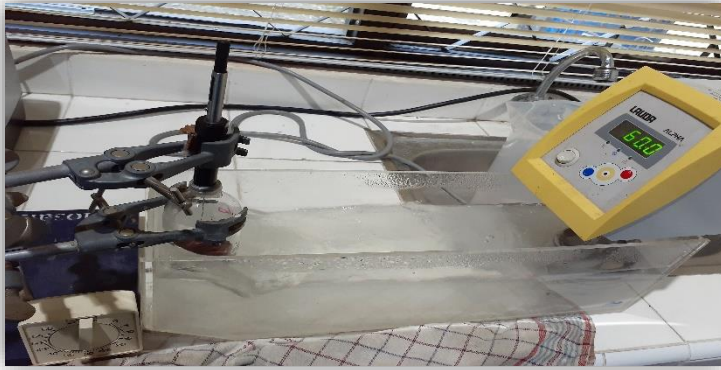
1. Proses Grinding/penghalusan sampel



2. Setelah dihaluskan sampel dimasukkan dalam tabung vial sebanyak masing-masing 5 gram



3. Proses inkubasi sampel yang sudah dipasangkan SPME dalam waterbath dengan suhu 60 °C selama 30 menit



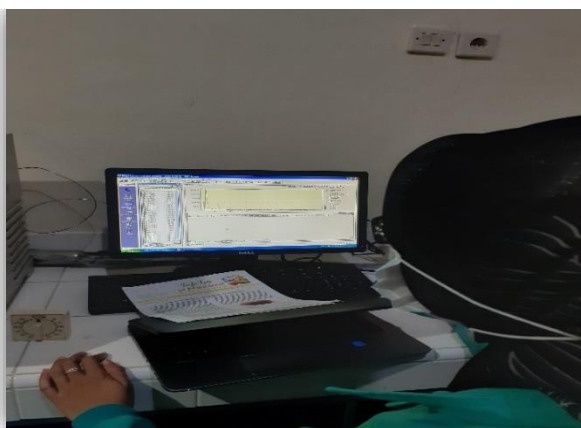
4. Alat SPME yang digunakan



5. Proses injeksi SPME ke dalam injector port GCMS



6. Pengolahan data senyawa volatil



Lampiran 17 : Dokumentasi Pengujian Asam Amino dengan Alat UFLC-LCMS 2020, Shimadzu

1. Biji kakao dihaluskan lalu ditimbang kemudian dimasukkan ke dalam Erlenmeyer lalu dimasukkan ke dalam penangas air



2. Sampel disaring menggunakan kertas saring, filtrat yang sudah terpisah diambil menggunakan jarum suntik dan disaring kembali



3. Filter kemudian diencerkan lalu dimasukkan ke dalam tabung vial untuk dianalisa menggunakan alat kromatografi *Liquid chromatography-mass spectrometry* (LC-MS)



4. Alat kromatografi *Liquid chromatography–mass spectrometry* (LC–MS) UFLC/LCMC 2020 Shimadzu



5. Pengolahan data senyawa Asam Amino



Lampiran 18 : Analisa Gula pereduksi

1. Biji kakao digerus hingga halus, lalu di timbang dan dimasukkan ke dalam labu takar, setelah itu ditambahkan aquades lalu dikocok sedikit. Kemudian ditambahkan dengan larutan Pb acetat



Sampel diberikan $(\text{NH}_4)_2\text{HPO}_4$ 10% sambil dilihat apakah terjadi endapan. Setelah itu ditambahkan air hingga tanda tera, kemudian labu ukur digoyang sebanyak 12 kali lalu sampel disaring untuk mendapatkan filtratnya



3. Larutan standar dan larutan sampel masing-masing diambil sebanyak 1 ml

Lampiran 19 : Profil Fermentasi biji kakao yang dilaksanakan oleh petani di kecamatan palolo

1. Biji kakao fermentasinya di ambil di daerah sentra produksi biji kakao Desa Sukaria, kec. Palolo, kab, Sigi Sulawesi Tengah.
2. Biji kakaonya dari Tanaman kakao tipe Kakao Lindak, jenis forestero, dari Klon Sulawesi -1 dan Sulawesi 2, jenis Hibrida.
3. Fermentasi dilakukan dalam kotak kayu dengan kapasitas 20 kg untuk setiap kotaknya.
4. Pengadukan biji kakao fermentasi dilakukan setiap 12 jam.
5. Fermentasi dilakukan selama 5-6 Malam dengan rincian sebagai berikut:
 - Lama waktu fermentasi di kotak atas selama 12 jam
 - Pengadukan dilakukan 1 kali setiap 12 jam
 - Setelah 24 jam biji kakao dipindah ke kotak bawah.
 - Biji kakao di kotak bawah selama 3 hari
 - Pengadukan dilakukan 1 kali 12 jam setiap hari setelah biji kakao dipindah ke kotak
6. Kadar air biji kakao fermentasi setelah di keringkan berkisar 6-8 %