THE ASSOCIATION BETWEEN +874 T/A GENETIC VARIATION OF INTERFERON GAMMA GENE AND SERUM INTERFERON GAMMA LEVEL IN PULMONARY TUBERCULOSIS PATIENTS

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Abstract: These days, Mycobacterium tuberculosis (Mtbb) bacillus is estimated to have infected one third of the world's population. World Health Organization (WHO) data shows that in 2011 there were 8.7 million case of tuberculosis (TB) or approximately 125 cases per 100,000 of the world's population. Some studies have been done to examine the role of certain genes toward the susceptibility of TB patients, most of those studies focused on genes that are involved in signal pathway of interferon gamma (IFN-γ). Interferon gamma is one of the cytokine that has a role in pathogenesis of pulmonary tuberculosis. Single nucleotide polymorphism of IFN-γ at +874 T/A position in the first intron is reported to have association with pulmonary TB in different population. The aim of this study is to investigate the genetic variation of IFN-γ in +874 T/A and its relation with IFN-γ plasma level in pulmonary tuberculosis patients. The study’s method is cross sectional. Blood sample was taken from the veins of new AFB (Acid-Fast Bacillus) positive tuberculosis patients. Purification examination of Deoxyribonucleic Acid (DNA) and Polymerase Chain Reaction-Restriciton Fragment Length Polymorphism (PCR-RFLP) for determining the genotype of IFN-γ genes and the level of IFN-γ is measured using Enzyme Linked Analysis Techniques Immunosorbent Assay (ELISA). The data is presented using tables and pictures. We found that there is a significant association between IFN-γ +874T/A genetic variations with the plasma level of IFN-γ. There is a domination of IFN-γ +874T/A heterozygote genotype found in pulmonary TB population with positive +1of AFB and far advanced lesion on chest x-ray.

Keywords: IFN-γ serum level, pulmonary TB, +874 T/A genetic variation of IFN-γ gene.

1. Introduction

Tuberculosis (TB) is an infectious disease that is caused by Mycobacterium tuberculosis (Mtbb) bacillus complex, in human, mostly caused by Mtbb species. This disease often invades lungs (pulmonary TB), but it also can invade other organs (extrapulmonary TB) [1].

Since a few decades ago, pulmonary TB has gained a lot of attention of medical experts. This is the result of the yearly increase prevalence of the disease. Pulmonary tuberculosis patients can be cured if handled from the outset and carefully. In 2012, it is estimated that the global incidence of TB was 8.6 million (ratio 122:100,000 per year) and mortality 1.3 million. From these numbers, there were 50,000 cases of Multidrug Resistance-Tuberculosis (MDR-TB) and 170,000 mortality rate from MDR-TB. Most of the case of global TB happens in South-East Asia and West Pacific totaling 58%, Africa 27% (ratio 255:100,000 per year), India 26%, China 12%, Eastern Mediterranean 8%, Europe 4%, and America 3%. The lowest incidence is mostly found in developed country such as Western Europe, Canada, USA, Japan, Australia, and New Zealand. Whereas in South Africa and Swaziland, incidence ratio of TB is 1000 : 100,000 per year [1]. In 2013, TB incidence in Indonesia was 327.094 with 80.99% being new positive AFB pulmonary TB. The biggest proportion of AFB positive pulmonary TB (21.40%) is found in productive adult age group (25-34 years old) [2].

Pulmonary tuberculosis is caused by positive gram bacillus, Mycobacterium tuberculosis. The Mtbb genome measured 4.4 Mb (mega base) with the most Guanine (G) and Cytosine (C) content. Results from the genes mapping, it is known that there are more than 165 genes and genetic marking that can be divided into 3 groups [3]. The main structure of Mtbb cell wall is mycolic acid, complex waxes, threhalose-dimycosylate which is known as cord factor, and mycobacterial sulpholipids that has a role in virulence. Mycobacterium tuberculosis is transmitted by air particles known as droplet nuclei with 1-5 micron in diameter. Transmission occurs if droplets nuclei that contains Mtbb is inhaled, later it enter the airway passage and finally end at lung alveoli [4], [5], [6], where Mtbb will be phagocytized by phagocytic immune cell (macrophage and dendrite cells) [7], [8]. The inflammation process will activate T helper-1 cell (Th1). One of the cytokine that is produced by Th1 cell is IFN-γ, which has an important role in eliminating Mtbb bacillus. Interferon gamma strengthens the phagocyte
potential from *Mtb*-infected macrophage by stimulating the formation of phagolysosomes. Interleukon gamma also stimulates the formation of free radicals to destroy the bacillus component, DNA, and cell wall [9].

The importance of IFN-γ in immune response towards mycobacteria has been shown by the increase of susceptibility, the severity of disease, and poor prognosis in individuals with the defect in IFN-γ, subunit receptor of IFN-γ, and STAT-1, either genetically or acquired, which in all is component of IFN-γ cytokine path [10]. The result of the study also shows that the genetic variation of IFN-γ gene influence the production of IFN-γ by T-cell clusters differentiated-4+ (CD4+) as well as cluster differentiated-8+ (CD8+). Homozygote TT/A and heterozygote T/A each linked with the production of high, low and moderate IFN-γ [9, 10].

Disturbance or decrease in the activity of Th1 cell and its cytokine, IFN-γ, has a meaningful role in influencing the mechanism of body defense mechanism against tuberculosis. Therefore, the information about the role of IFN-γ in body defense mechanism against pulmonary tuberculosis is very important.

2. Method

This study uses a draft research cross sectional. Sample used is the plasma of pulmonary tuberculosis patients in Makassar, South Sulawesi, that are assessed using purification technique Deoxyribonucleic Acid (DNA) and Polymerase Chain Reaction-Restriction Fragment Length Polymorphism (PCR-RFLP) to determine the genotype gene of IFN-γ and Enzyme Linked Immunosorbent Assay (ELISA) examination to measure the plasma level of IFN-γ. Data is presented in tables and figures.

Sample Selection Method

Selection of the study subject uses accidental sampling method. Accidental sampling is the unplanned method of study subject selection, the patients that come for treatment within certain period and meet certain criteria were asked to be the subject. Period and criteria of this study is new case that has been diagnosed with positive AFB pulmonary tuberculosis who come in during September – December 2014 in Makassar.

Examination Procedure of IFN-γ Level Using ELISA Kit

5 ml of the subject’s venous blood was taken and put inside non-EDTA tube. The tube then centrifuged in 5000 rpm for 30 minutes. The blood that has been centrifuged will separate as erythrocyte, buffy coat layer, and blood plasma. Plasma then separated and stored in microtube. Later it was wrapped using paraffin, where it stored in the refrigerator in -80°C until it was used.

Blood plasma and ELISA kit then put in room temperature. Dissolving Lyophilized IFN-γ Standard and Assay Dihuent made standard solution. The standard solution later examined using duplo, while the other well filled with the sample that was added with Assay Dihuent first. Every well then added with Rabbit anti-Human IFN-γ Polyconal Antibody. The plate then closed with sealer (Acetate Plate Sealer) to prevent evaporation and incubated in room temperature for 3 hours.

After incubation, the sealer was opened and the plate then washed using Wash Buffer. Goat anti-Rabbit Conjugated Alkaline Phosphates then added to each well and resealed, later incubated in room temperature for 45 minutes. Sealer then opened and the fluid discarded, after that the plate washed using Wash Buffer. Coloring reagent then added and incubated in room temperature for 6 minutes, afterwards the stop solution is added. The result was read using ELISA reader so that we could find the IFN-γ level.

Data Analysis

The gathered data is presented in tables and figures with description provided.

3. Results

This study used 62 subjects of new positive AFB pulmonary TB patients who has had their +874 T/A IFN-γ gen genetic variation and IFN-γ plasma levels examined when the diagnosis was made. From the 62 subjects, the younges is 17 years old and the oldest is 65 year old, with mean age of 36.5 ±14.5 years old. Almost 75% of pulmonary TB patients are in <50 years old age group.

Most of the pulmonary TB patients in this study come from age group of 16-29 years old. Most of the genetic variation of +874 T/A IFN-γ in our pulmonary TB population heterozygote TA (32 subject or 51.6%) and the lowest is wild type homozygote TT (4 subjects or 6.5%). Distribution of subject’s +874 T/A IFN-γ gen genetic variation based on age group is presented in table 1.

<table>
<thead>
<tr>
<th>Genetic Variation</th>
<th>16-29 n (%)</th>
<th>30-49 n (%)</th>
<th>50-65 n (%)</th>
<th>Total n (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>AA</td>
<td>8 (12.9)</td>
<td>10 (16.1)</td>
<td>3 (4.8)</td>
<td>21 (33.9)</td>
</tr>
<tr>
<td>TA</td>
<td>12 (19.4)</td>
<td>10 (16.1)</td>
<td>10 (16.1)</td>
<td>32 (51.6)</td>
</tr>
<tr>
<td>TT</td>
<td>1 (1.6)</td>
<td>1 (1.6)</td>
<td>2 (3.2)</td>
<td>4 (6.5)</td>
</tr>
<tr>
<td>Total</td>
<td>24 (38.7)</td>
<td>22 (35.5)</td>
<td>16 (25.8)</td>
<td>62 (100.0)</td>
</tr>
</tbody>
</table>

In this study, the most of the male’s genetic variation of +874 T/A IFN-γ found is the TA genotype (32.3%) and the least is TT genotype (3.2%), the same result also found in woman where +874 T/A IFN-γ gen genetic variation found the most is TA genotype (19.4%) and the least is TT genotype (3.2%). The distribution of subject’s +874 T/A IFN-γ gen genetic variation based on gender is presented in table 2.

<table>
<thead>
<tr>
<th>Genetic Variation</th>
<th>Male</th>
<th>Female</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>AA</td>
<td>16 (19.4)</td>
<td>9 (14.5)</td>
<td>21 (33.9)</td>
</tr>
<tr>
<td>TA</td>
<td>20 (23.3)</td>
<td>12 (19.4)</td>
<td>32 (51.6)</td>
</tr>
<tr>
<td>TT</td>
<td>2 (2.4)</td>
<td>2 (3.2)</td>
<td>4 (6.5)</td>
</tr>
<tr>
<td>Total</td>
<td>37 (59.7)</td>
<td>25 (40.3)</td>
<td>62 (100)</td>
</tr>
</tbody>
</table>

Table 2. Distribution of +874 T/A IFN-γ gen genetic variation based on gender
Toward all 62 subjects after the diagnosis of pulmonary TB was enforced, the IFN-γ level examination was executed. Obtained the average of 22.13±22.33 pg/mL, with the lowest level in 6.25 pg/mL and the highest 112.97 pg/mL.

To understand the relation between +874 T/A IFN-γ gen genetic variation and IFN-γ plasma level, we use the Spearman nonparametric correlation test considering the abnormal distribution of IFN-γ plasma level. The result of Spearman correlation test shows that there is a significant correlation between +874 T/A IFN-γ gen genetic variation and IFN-γ plasma level with p=0.001 (p<0.01). (figure 1)

Figure 1. IFN-γ plasma level (phase 0) based on +874 T/A IFN-γ gen genetic variation. Red line shows the median of IFN-γ level in each genotype.

The AFB level in sputum test and the lesion area in chest x-ray can determine the severity of pulmonary TB. In this study based on the lesion area, the most common to find is far advanced lesion (48.4%). The most genetic variation of +874 T/A IFN-γ gen found in pulmonary TB patients with broad lesion is TA genotype (29.0%) and the least is TT genotype (0.0%). The distribution of +874 T/A IFN-γ gen genetic variation based on lesion area in chest x-ray is presented in table 3.

Table 3. Distribution of +874 T/A IFN-γ gen genetic variation based on lesion area in chest x-ray

<table>
<thead>
<tr>
<th>Genetic Variation</th>
<th>Degree of Lesion in Chest X-ray</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Minimal</td>
<td>Moderate</td>
</tr>
<tr>
<td>AA</td>
<td>3 (4.8)</td>
<td>8 (12.9)</td>
</tr>
<tr>
<td>TA</td>
<td>5 (8.1)</td>
<td>9 (14.5)</td>
</tr>
<tr>
<td>TT</td>
<td>2 (3.2)</td>
<td>2 (3.2)</td>
</tr>
<tr>
<td>TA/TT</td>
<td>0 (0.0)</td>
<td>3 (4.8)</td>
</tr>
<tr>
<td>Total</td>
<td>10 (16.1)</td>
<td>22 (35.5)</td>
</tr>
</tbody>
</table>

Based on the positivity of AFB sputum test, in this study, the most common to find is AFB positive one (59.7%). Genetic variation of +874 T/A IFN-γ gen found the most in the subject with AFB positive one is TA genotype (33.9%), and the least is TA/TT genotype (1.6%). Distribution of +874 T/A IFN-γ gen genetic variation based on positivity of AFB sputum test is presented in table 4.

Table 4. Distribution of +874 T/A IFN-γ gen genetic variation based on positivity of AFB sputum test

<table>
<thead>
<tr>
<th>Genetic Variation</th>
<th>AFB Sputum Test</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>+1</td>
<td>+2</td>
</tr>
<tr>
<td>AA</td>
<td>13 (21.0)</td>
<td>5 (8.1)</td>
</tr>
<tr>
<td>TA</td>
<td>21 (33.9)</td>
<td>5 (8.1)</td>
</tr>
<tr>
<td>TT</td>
<td>2 (3.2)</td>
<td>1 (1.6)</td>
</tr>
<tr>
<td>TA/TT</td>
<td>1 (1.6)</td>
<td>0 (0.0)</td>
</tr>
<tr>
<td>Total</td>
<td>37 (59.7)</td>
<td>11 (17.1)</td>
</tr>
</tbody>
</table>

4. Discussion

This study aims to investigate the role of IFN-γ +874 T/A gen polymorphism and IFN-γ plasma level with the severity of pulmonary TB patients based on the positivity of AFB sputum test and lesion area in chest x-ray.

In this study, the most genetic variation of IFN-γ +874 T/A gen that was found is TA genotypes, namely 51.6% and 32.3%, are male. Etokebe [12] study also found that the group with TA genotype is the group with the highest frequency of genetic namely 50.4%, different result found in Vallinoto study [11] that has AA genotype group namely 56.6%, as well as Hwang [15] who also found similar major genotype frequency that is AA group namely 82.5%. The relation between the susceptibility of TB and IFN-γ polymorphisms has been observed inside different population [10], [12].

This study finds that there is a significant correlation between IFN-γ +874 T/A gene genetic variation with IFN-γ plasma level (p=0.001). Pravica, [16] shows that genetic variation of single nucleotide (T→A) in +874 position at first intron from IFN-γ gen influence the production of IFN-γ, possibly caused by the +874 position is the are that responsible for NF-κB factor transcription. The correlation between genetic variation of IFN-γ +874 T/A and ex-vivo production of IFN-γ by peripheral mononuclear cells in Lopez [10] study indicates that the linkage with functional variants in other locus in that gene regulation area. The specific bond of NF-κB to the DNA order that contains +874 alleles causes functional consequence towards IFN-γ gene’s transcription and its production [17], [9]. Study conducted by Vallinoto [11] shows that tuberculosis patients with +874 A/A genotype show profound low IFN-γ plasma level compared to patients with +874 T/A and +874 T/T genotype, which can be strong evidence that the polymorphisms decreases the production of IFN-γ and cellular immunity activation that can increase the possibility of infection.

Genetic variation based on lesion area found the most in this study is far advanced lesion with TA genotype as the group with highest genetic frequency namely 29.0%, different result found in a study by Lopez [10] where the most patients are patients with moderate lesion and AA genotype.

Analysis of genetic variation of IFN-γ +874 T/A towards subjects in this study shows that from the four genotype types, homozygous +874 T/A constituting the most genotype (51.6%). Polymorphisms of IFN-γ +874 has shown to have meaningful correlation with tuberculosis. Studies by Lopez [10] and Pravica [16] found that frequency of allele A is high in pulmonary TB patients and control with TST (+) and TST (-) compared with miliary tuberculosis patients and pleural TB, whereas allele T that connects with the IFN-γ production.
that is high in-vitro, increases in patients with miliary and
pleural TB. Vallinoto [11] shows that there is a correlation
between allele +874 A and active TB, at the same times
shows a high frequency of allele +874 T in control group.
These results show the protective role of allele +874 T, on
the other hand, allele +874 A is the predisposition factor of
Mtb infection. The result of these studies strengthen the
correlation between genotype +874 AA with the susceptibility
towards bacteria, whereas genotype +874 T/T may have
connected with the protection against Mtb infection, or even
the partial protection if allele T shows in heterozygote.

The expression and production of IFN-γ is regulated
normally, study by Lopez [10] shows that this polimorphisms influence the production of IFN-γ inducted by PPD in TB patients, healthy contact, or normal subject. The lowest production of IFN-γ found in AA genotype both before and after therapy. Production of IFN-γ with genotype AA is twice as low compared with the other two genotype (TA and TT) and still low even after 6 month post treatment even compared to control. Settlement of low IFN-γ production by peripheral mononuclear cell more than 6 months show that there is a genetic defect in the production of IFN-γ in the patients with homozygote allele A that become the basis of the increase of reactivation ratio from one latent tuberculosis focus. Statistic analysis also shows that the presence of AA genotype and lymphocyte count is the independent predictor toward the production of IFN-γ.

In our study, the most commonly genetic variation found in patients with AFB sputum test result of positive 3 is genotype TA. This is in accordance with the study of Etokobe [12], in tuberculosis patients with positive AFB sputum test result the most commonly found genotype group is TA, but it is said to not have any influence toward the severity of the disease.

References

Author Profile

Erwin Arief received the M.D. and specialist in Internal Medicine from Medical Faculty Hasanuddin University, Makassar, South Sulawesi in 1998 and 2009 respectively. In 2008, he received Pulmonology Specialist from Medical Faculty Airlangga University. He now work as a staff of Department of Pulmonology and Respiratory Medicine, Medical Faculty of Hasanuddin University, Makassar, South Sulawesi, Indonesia.

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