

DAFTAR PUSTAKA

- Abdullah, M., Maryam, S., 2020. Uji Aktivitas Antioksidan Isolat Fungi Endofit Daun Galing-galing (*Cayratia trifolia* L.) dengan Metode 1,1-Diphenyl-2-Picrylhydrazil (DPPH), Jurnal Farmasi Desember.
- Aini, K., Elfita, Widjajanti, H., Setiawan, A., Kurniawati, A.R., 2022. Antibacterial activity of endophytic fungi isolated from the stem bark of jambu mawar (*Syzygium jambos*). Biodiversitas 23, 521–532. <https://doi.org/10.13057/biodiv/d230156>
- Asomadu, R.O., Ezeorba, T.P.C., Ezike, T.C., Uzoechina, J.O., 2024. Exploring the antioxidant potential of endophytic fungi: a review on methods for extraction and quantification of total antioxidant capacity (TAC). 3 Biotech 14, 127. <https://doi.org/10.1007/s13205-024-03970-3>
- Azim, M., Novasari, N., Hariadi, P., Febriani, Y., Yuliana, T.P., 2022. Endophyte Extract From Nettle (*Urtica dioica* L.) Against *Staphylococcus aureus* and *Escherichia coli*. ALKIMIA : Jurnal Ilmu Kimia dan Terapan 6, 196–203. <https://doi.org/10.19109/alkimia.v6i1.11277>
- Bahadori Ganjabadi, P., Farzaneh, M., Mirjalili, M.H., 2023. Development and Optimization of Culture Medium for the Production of Glabridin by *Aspergillus eucalypticola*: An Endophytic Fungus Isolated from *Glycyrrhiza glabra* L. (Fabaceae). Mycobiology 51, 230–238. <https://doi.org/10.1080/12298093.2023.2225253>
- Bains, A., Chawla, P., 2020. In vitro bioactivity, antimicrobial and anti-inflammatory efficacy of modified solvent evaporation assisted *Trametes versicolor* extract. 3 Biotech 10. <https://doi.org/10.1007/s13205-020-02397-w>
- Baron, N.C., Rigobelo, E.C., 2022. Endophytic fungi: a tool for plant growth promotion and sustainable agriculture. Mycology. <https://doi.org/10.1080/21501203.2021.1945699>
- Breijyeh, Z., Jubeh, B., Karaman, R., 2020. Resistance of gram-negative bacteria to current antibacterial agents and approaches to resolve it. Molecules. <https://doi.org/10.3390/molecules25061340>
- Chaudhary, A., Kumar, P., Jaswal, V.S., Thakur, A., 2019. Analytical Techniques for the Identification and Quantification of Flavonoids, in: Current Aspects of Flavonoids: Their Role in Cancer Treatment. Springer Singapore, Singapore, pp. 9–22. https://doi.org/10.1007/978-981-13-5874-6_2
- Coskun, O., 2016. Separation techniques: Chromatography. North Clin Istanb 3, 156–160. <https://doi.org/10.14744/nci.2016.32757>
- De Gregorio, M.A., Zhang, L., Mahomoodally, M.F., Zengin, G., Jugreet, S., Yildiztugay, E., Fiorini, A., Lucini, L., 2024. Metabolomic Profiles and Biopharmaceutical Properties of *Petrosimonia brachiata* and *P. nigdeensis* from Turkey. Plants 13, 2073. <https://doi.org/10.3390/plants13152073>
- Destryana, R., Estiasih, T., Sukardi, Pranowo, D., 2024. Zingiberaceae Rhizome Essential Oil: A Review of Chemical Composition, Biological Activity, and Application in Food Industry. IOP Conf Ser Earth Environ Sci 1299, 012010. <https://doi.org/10.1088/1755-1315/1299/1/012010>
- Dolok, T.N.S., Nursaadah, E., Primairyani, A., 2023. Keanekaragaman Hayati Tumbuhan an Pemanfaatannya. BIOEDUSAINS:Jurnal Pendidikan Biologi 7453. <https://doi.org/10.31539/bioedusains.v6i2.7547>
- i, A.S., do Vale, H.M.M., 2022a. Methods used for the study of review on methodologies and challenges, and associated tips. s://doi.org/10.1007/s00203-022-03283-0



- dos Reis, J.B.A., Lorenzi, A.S., do Vale, H.M.M., 2022b. Methods used for the study of endophytic fungi: a review on methodologies and challenges, and associated tips. *Arch Microbiol.* <https://doi.org/10.1007/s00203-022-03283-0>
- Fadiji, A.E., Babalola, O.O., 2020. Exploring the potentialities of beneficial endophytes for improved plant growth. *Saudi J Biol Sci.* <https://doi.org/10.1016/j.sjbs.2020.08.002>
- Kancherla, N., Dhakshinamoothi, A., Chitra, K., Komaram, R.B., 2019. Preliminary Analysis of Phytoconstituents and Evaluation of Anthelmintic Property of *Cayratia auriculata* (In Vitro). *Maedica (Bucur)* 14, 350–356. <https://doi.org/10.26574/maedica.2019.14.4.350>
- KAPA Biosystems, 2013. KAPA HiFi HotStart ReadyMix PCR Kit (No. KR0370), v5.13. Boston, Massachusetts.
- Khairuddin, K., Taebe, B., Kombong, O.I., 2021. Measurement of Specific and Non-Specific Parameters of Sampare Leaves Ethanol Extract (*Glochidion* sp var. Biak) as a Traditional Antimalarial Agent. *Journal of Pharmaceutical and Medicinal Sciences* 6.
- Kowalska, T., Sajewicz, M., 2022. Thin-Layer Chromatography (TLC) in the Screening of Botanicals-Its Versatile Potential and Selected Applications. *Molecules* 27. <https://doi.org/10.3390/molecules27196607>
- LibreTexts Chemistry, 2024. General Lab Techniques, 1st ed.
- Linh, N.N., Hop, N.Q., Nhungh, P.T.T., Dao, P.T.B., Manh, V.Q., Pham, T.V., Son, N.T., 2024. *Glochidion* Species: A Review on Phytochemistry and Pharmacology. *Nat Prod Commun.* <https://doi.org/10.1177/1934578X241276962>
- Magalhães, L.M., Almeida, M.I.G.S., Barreiros, L., Reis, S., Segundo, M.A., 2012. Automatic Aluminum Chloride Method for Routine Estimation of Total Flavonoids in Red Wines and Teas. *Food Anal Methods* 5, 530–539. <https://doi.org/10.1007/s12161-011-9278-1>
- Mamangkey, J., Mendes, L.W., Harahap, A., Briggs, D., Kayacilar, C., 2022. Endophytic Bacteria and Fungi from Indonesian Medicinal Plants with Antibacterial, Pathogenic Antifungal and Extracellular Enzymes Activities: A Review. *International Journal of Science* 3, 245–255.
- Martin, K.J., Rygiewicz, P.T., 2005. Fungal-specific PCR primers developed for analysis of the ITS region of environmental DNA extracts. *BMC Microbiol* 5. <https://doi.org/10.1186/1471-2180-5-28>
- McKinnon, A.C., 2016. Plant Tissue Preparation for the Detection of an Endophytic Fungus In *Planta*, in: Glare Travis R. and Moran-Diez, M.E. (Ed.), *Microbial-Based Biopesticides: Methods and Protocols*. Springer New York, New York, NY, pp. 167–173. https://doi.org/10.1007/978-1-4939-6367-6_13
- Mishra, V.K., Passari, A.K., Leo, V.V., Singh, B.P., 2017. Molecular Diversity and Detection of Endophytic Fungi Based on Their Antimicrobial Biosynthetic Genes, in: Singh, B.P., Gupta, V.K. (Eds.), *Molecular Markers in Mycology: Diagnostics and Marker Developments*. Springer International Publishing, Cham, pp. 1–35. https://doi.org/10.1007/978-3-319-34106-4_1
- Murdiah, S., 2017. Endophytic Fungi of Various Medicinal Plants Collected From Evergreen Forest Baluran National Park and Its Potential as Laboratory Manual for *Jurnal Pendidikan Biologi Indonesia* 3, 64–71.
- B., Bawadikji, A.A., Lim, J.W., Tong, W.Y., Leong, C.R., Khaw, 22. Recent Developments in Metabolomics Studies of Endophytic Fungi. <https://doi.org/10.3390/jof8010028>
- P., Segun, P.A., 2019. Medicinal mushrooms with methicillin *occus aureus* (MRSA) inhibitory activity. *Journal of Pharmacy & I.* <https://doi.org/10.4314/jpb.v16i1.5>



- Oktalia, G., Chrystomo, L.Y., Karim, A.K., 2017. Uji Aktivitas Sitotoksik dan Analisis Fitokimia Ekstrak Etanol Daun Sampare (*Glochidion* sp.). Jurnal Biologi Papua 9, 49–54.

Ouchari, L., Boukeskasse, A., Bouizgarne, B., Ouhdouch, Y., 2019. Antimicrobial potential of actinomycetes isolated from the unexplored hot Merzouga desert and their taxonomic diversity. Biol Open 8. <https://doi.org/10.1242/bio.035410>

Pekal, A., Pyrzynska, K., 2014. Evaluation of Aluminium Complexation Reaction for Flavonoid Content Assay. Food Anal Methods 7, 1776–1782. <https://doi.org/10.1007/s12161-014-9814-x>

Prabu, S.L., Suriya Prakash, T.N.K., Thirumurugan, R., 2015. Cleaning Validation and Its Regulatory Aspects in the Pharmaceutical Industry, in: Developments in Surface Contamination and Cleaning. Elsevier, pp. 129–186. <https://doi.org/10.1016/B978-0-323-31303-2.00005-4>

Pyka, A., 2014. Detection progress of selected drugs in TLC. Biomed Res Int 2014, 732078. <https://doi.org/10.1155/2014/732078>

Qin, Z., Liu, H.-M., Ma, Y.-X., Wang, X.-D., 2021. Developments in extraction, purification, and structural elucidation of proanthocyanidins (2000–2019). pp. 347–391. <https://doi.org/10.1016/B978-0-12-819485-0.00008-6>

Rianto, A., Isrul, M., Anggarini, S., Saleh, A., 2018. Isolasi Dan Identifikasi Fungi Endofit Daun Jambu Mete (*Anacardium occidentale* L.) Sebagai Antibakteri Terhadap *Salmonella typhimurium*. Jurnal Mandala Pharmacon Indonesia 4.

Rohde, M., 2024. Chapter 3 - Bacterial ultrastructure, in: Tang, Y.-W., Hindiyeh, M.Y., Liu, D., Sails, A., Spearman, P., Zhang, J.-R. (Eds.), Molecular Medical Microbiology (Third Edition). Academic Press, pp. 23–43. <https://doi.org/https://doi.org/10.1016/B978-0-12-818619-0.00163-5>

Sahu, P.K., Tilgam, J., Mishra, S., Hamid, S., Gupta, A., Jayalakshmi, K., Verma, S.K., Kharwar, R.N., 2022a. Surface sterilization for isolation of endophytes: Ensuring what (not) to grow. J Basic Microbiol 62, 647–668. <https://doi.org/10.1002/JOBM.202100462>

Sahu, P.K., Tilgam, J., Mishra, S., Hamid, S., Gupta, A., K., J., Verma, S.K., Kharwar, R.N., 2022b. Surface sterilization for isolation of endophytes: Ensuring what (not) to grow. J Basic Microbiol 62, 647–668. <https://doi.org/10.1002/jobm.202100462>

Schoch, C.L., Seifert, K.A., Huhndorf, S., Robert, V., Spouge, J.L., Levesque, C.A., Chen, W., Bolchacova, E., Voigt, K., Crous, P.W., Miller, A.N., Wingfield, M.J., Aime, M.C., An, K.D., Bai, F.Y., Barreto, R.W., Begerow, D., Bergeron, M.J., Blackwell, M., Boekhout, T., Bogale, M., Boonyuen, N., Burgaz, A.R., Buyck, B., Cai, L., Cai, Q., Cardinali, G., Chaverri, P., Coppins, B.J., Crespo, A., Cubas, P., Cummings, C., Damm, U., de Beer, Z.W., de Hoog, G.S., Del-Prado, R., Dentinger, B., Diéguez-Uribondo, J., Divakar, P.K., Douglas, B., Dueñas, M., Duong, T.A., Eberhardt, U., Edwards, J.E., Elshahed, M.S., Fliegerova, K., Furtado, M., García, M.A., Ge, Z.W., Griffith, G.W., Griffiths, K., Groenewald, J.Z., Groenewald, M., Grube, M., Gryzenhout, M., Guo, L.D., Hagen, F., Hambleton, S., Hamelin, R.C., Hansen, K., Harrold, P., Heller, G., Herrera, C., Hirayama, K., Hirooka, Y., Ho, H.M., Hoffmann, K., Hofstetter, Högnabba, F., Hollingsworth, P.M., Hong, S.B., Hosaka, K., ghes, K., Huhtinen, S., Hyde, K.D., James, T., Johnson, E.M., 1ston, P.R., Jones, E.B.G., Kelly, L.J., Kirk, P.M., Knapp, D.G., G.M., Kurtzman, C.P., Landvik, S., Leavitt, S.D., Liggenstoffer, C., Lombard, L., Luangsa-ard, J.J., Lumbsch, H.T., Maganti, H., Ira, S.S.N., Martin, M.P., May, T.W., McTaggart, A.R., Methven, Moncalvo, J.M., Mongkolsamrit, S., Nagy, L.G., Nilsson, R.H., si, I., Okada, G., Okane, I., Olariaga, I., Otte, J., Papp, T., Park,



- D., Petkovits, T., Pino-Bodas, R., Quaedvlieg, W., Raja, H.A., Redecker, D., Rintoul, T.L., Ruibal, C., Sarmiento-Ramírez, J.M., Schmitt, I., Schüßler, A., Shearer, C., Sotome, K., Stefani, F.O.P., Stenroos, S., Stielow, B., Stockinger, H., Suetrong, S., Suh, S.O., Sung, G.H., Suzuki, M., Tanaka, K., Tedersoo, L., Telleria, M.T., Tretter, E., Untereiner, W.A., Urbina, H., Vágvölgyi, C., Vialle, A., Vu, T.D., Walther, G., Wang, Q.M., Wang, Y., Weir, B.S., Weiß, M., White, M.M., Xu, J., Yahr, R., Yang, Z.L., Yurkov, A., Zamora, J.C., Zhang, N., Zhuang, W.Y., Schindel, D., 2012. Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. Proc Natl Acad Sci U S A 109, 6241–6246. <https://doi.org/10.1073/pnas.1117018109>
- Singh, P., Sharma, A., Bordoloi, M., Nandi, S.P., 2020. Molecular identification of endophytic fungi isolated from medicinal plant. Biointerface Res Appl Chem 10, 6436–6443. <https://doi.org/10.33263/BRIAC105.64366443>
- Singh, V.K., Kumar, A., 2023. Secondary metabolites from endophytic fungi: Production, methods of analysis, and diverse pharmaceutical potential. Symbiosis 1–15. <https://doi.org/10.1007/s13199-023-00925-9>
- Soković, M., Glamočlija, J., Ćirić, A., Petrović, J., Stojković, D., 2018. Mushrooms as Sources of Therapeutic Foods, in: Therapeutic Foods. Elsevier, pp. 141–178. <https://doi.org/10.1016/b978-0-12-811517-6.00005-2>
- Vallavan, V., Krishnasamy, G., Zin, N.M., Latif, M.A., Kebangsaan Malaysia, U., Raja, J., Aziz, M.A., 2020. A Review on Antistaphylococcal Secondary Metabolites from Basidiomycetes. Molecules 2020, Vol. 25, Page 5848 25, 5848. <https://doi.org/10.3390/MOLECULES25245848>



Optimized using
trial version
www.balesio.com