

BAB VIII

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LAMPIRAN 1

1. Prosedur pewarnaan jaringan biologis penderita kanker payudara

<p>1. Potong blok paraffin dengan mikrotom pada ketebalan 3-4μ</p> 	<p>2. Celupkan kedalam Waterbath</p> 
<p>3. Ambil potongan jaringan dengan slide lalu tiriskan</p> 	<p>4. Tulis pada slide kode sesua blok paraffin dengan pensil</p> 
<p>5. Panaskan slide diatas Hot Plate selama 1 jam</p> 	<p>6. Dinginkan slide lalu masukkan kedalam keranjang slide</p> <p>7. Deparafinasi (Xylol I, Xylol II, Xylol III) masing-masing 5 menit</p> 



<p>8. Rehidrasi (Alkohol 96%, Alkohol 80%, Alkohol 70%), masing-masing selama 5 menit</p> 	<p>9. Cuci air mengalir selama 5 menit</p> 
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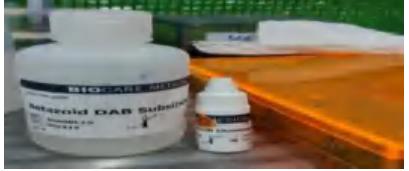
<p>10. Angkat dari air lalu masukkan slide kedalam larutan Blocking Endogen Peroksida, rendam selama 15 menit</p> 	<p>11. Cuci air mengalir selama 5 menit</p> 
<p>12. Masukkan keranjang berisi slide kedalam decloaking yang berisi larutan Antigen Retrieval Decloaking Chamber, lalu letakkan slide pada rack holder</p> 	<p>13. Masukkan rack holder kedalam decloaking, lalu tutup</p> 
<p>14. Atur waktu yaitu selama 40 menit pada suhu 95 derajat.</p> 	<p>15. Dinginkan, dengan mengeluarkan slide dari decloaking dan disimpan pada suhu ruangan</p> <p>16. Setelah dingin, cuci dalam larutan PBS 2x masing-masing selama 5 menit</p> <p>17. Tandai slide dengan memberi lingkaran sekitar jaringan</p>



	
18. Atur slide pada baki slide 	19. Ambil satu per satu lalu tetesi Background Sniper lalu inkubasi selama 30 menit 

20. Buang larutan background sniper dengan cara ditiriskan pada tisu 21. Tetesi Antibody Primer lalu inkubasi selama 1 jam pada suhu ruang 22. Cuci PBS 2x selama 5 menit 	23. Tetesi Trekkie Universal lalu diamkan selama 30 menit 
24. Tiriskan pada tissue lalu tetesi Trekkavidin-HRP lalu diamkan selama 30 menit 	25. Cuci PBS 2x dengan cara merendam slide selama 5 menit 
mobil menunggu pencucian, at larutan DAB dengan cara mpurkan cromogen DAB 1	27. Amati jaringan jika sudah menunjukkan warna coklat, langsung direndam ke dalam air selama 5 menit



<p>tetes + Substrat buffer 1 ml (dicampur dalam tabung bersih) lalu tetesi ke jaringan.</p> 	
<p>28. Rendam dengan hematoxylin Meyer 5 menit</p> 	<p>29. Cuci air mengalir selama 5 menit</p> 
<p>30. Dehidrasi (Alkohol 70%, Alkohol 80%, Alkohol 96%) masing-masing 5 menit</p> 	<p>31. Clearing (Xylol I, Xylol II, Xylol III)</p> 
<p>32. Keringkan slide lalu tetesi dengan entelan lalu tutup dengan deck glass.</p>	<p>33. Amati di mikroskop</p>



