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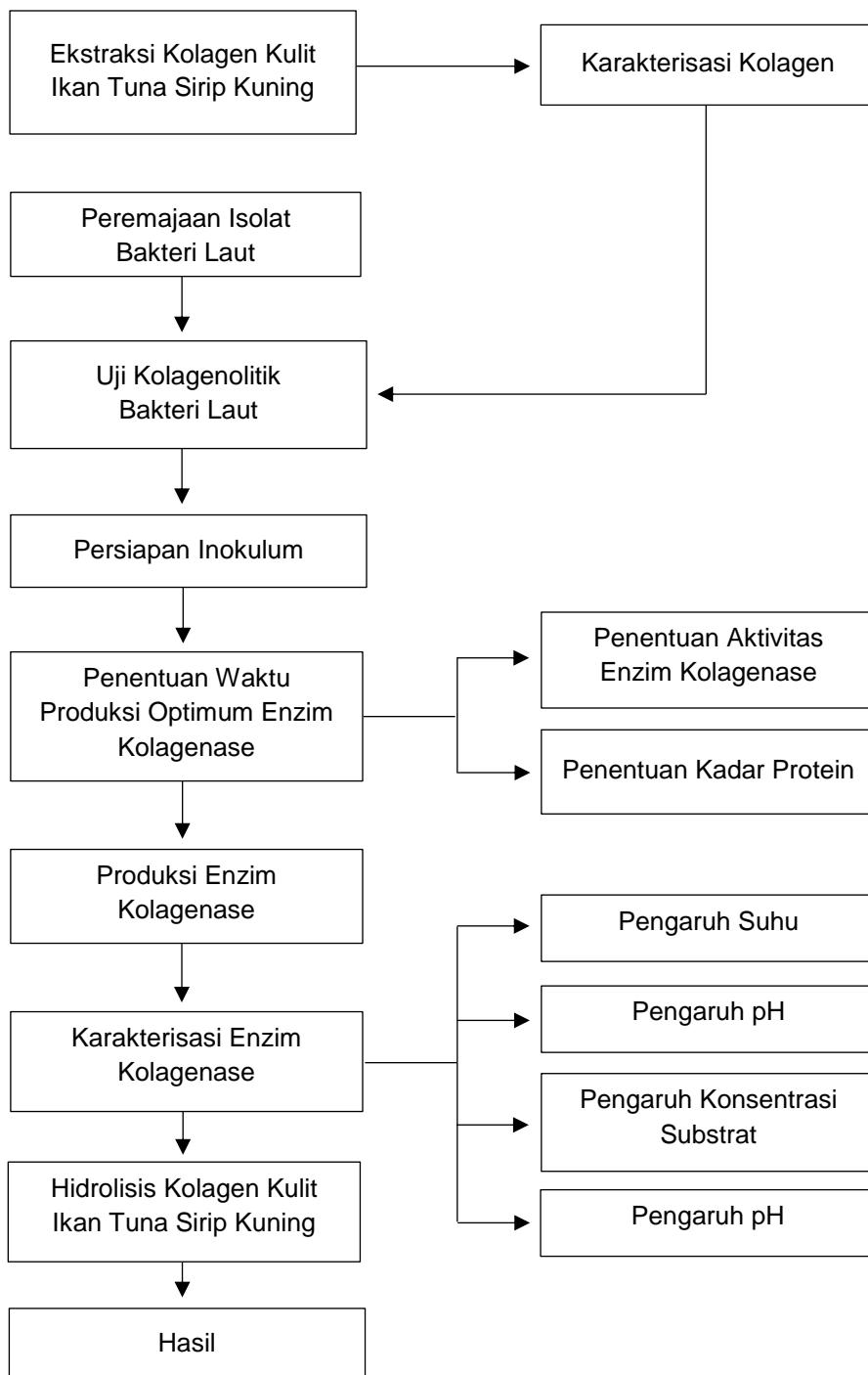
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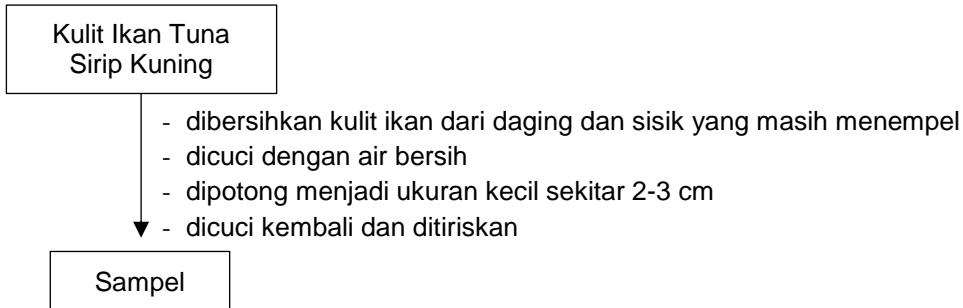
**Lampiran 1. Diagram Alir Penelitian**



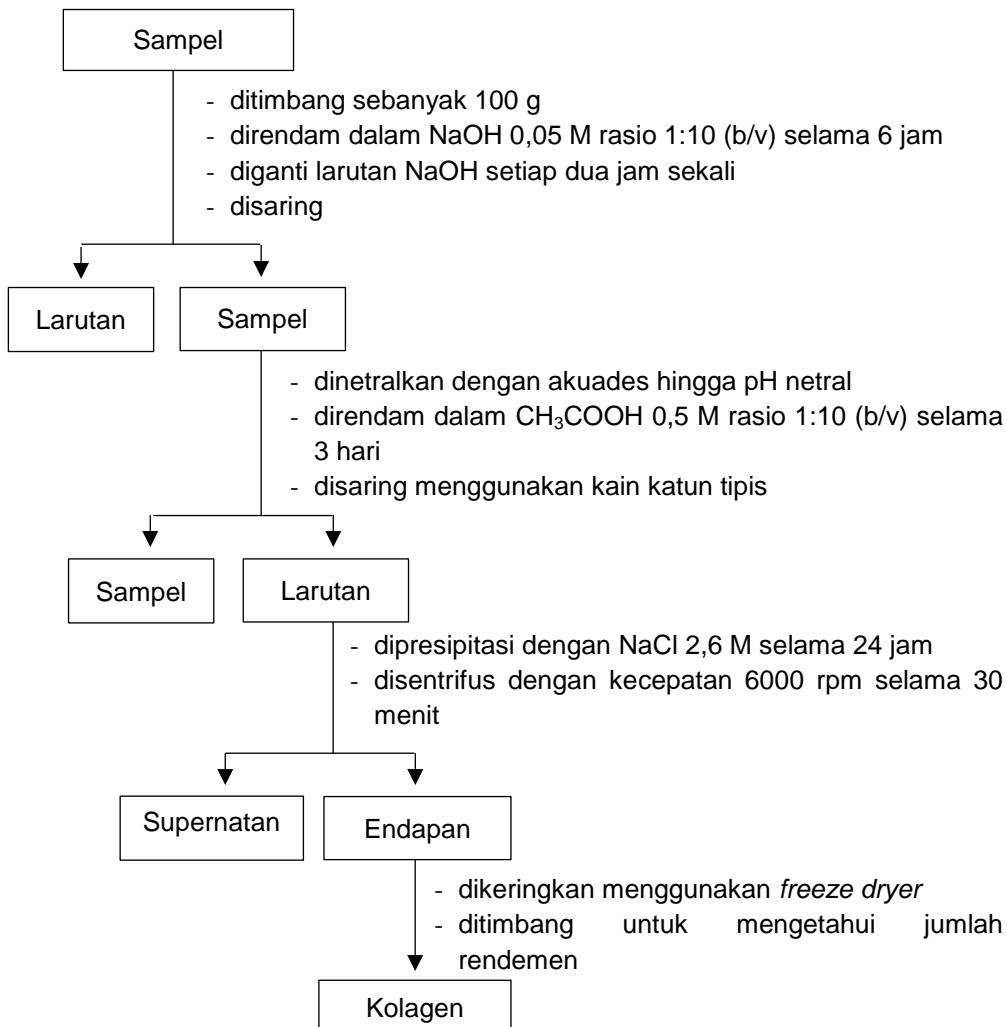
## Lampiran 2. Prosedur Penelitian

### 1. Ekstrasi Kolagen Kulit Ikan Tuna Sirip Kuning

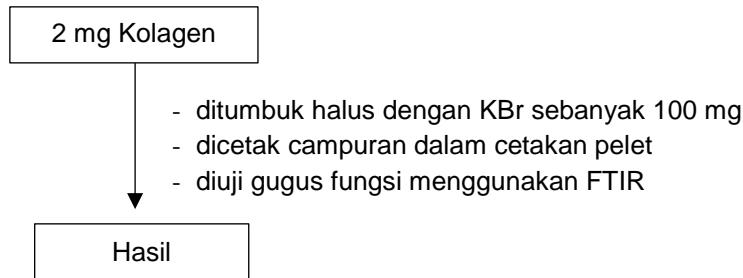
#### a. Preparasi Sampel



#### b. Ekstraksi Kolagen

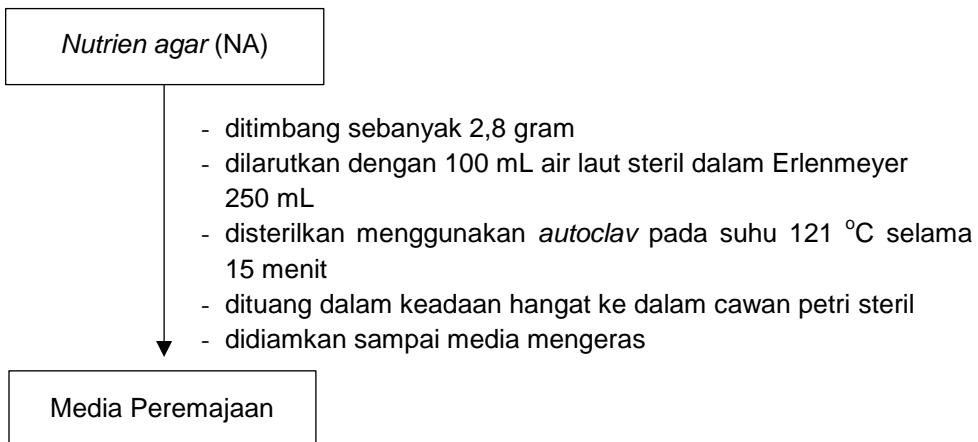


### c. Karakterisasi Kolagen



## 2. Pembuatan Media

### a. Media Peremajaan



### b. Media Substrat

2 g agar base, 0,5 g yeast,  
0,5 g pepton, 0,5 g NaCl,  
substrat 1%

- ditimbang masing-masing dan dimasukkan ke dalam Erlenmeyer 250 mL
- dilarutkan dengan air laut steril hingga volume 100 mL
- disterilkan menggunakan *autoclav* pada suhu 121 °C selama 15 menit
- dituang dalam keadaan hangat ke dalam cawan petri steril
- didiamkan hingga mengeras

Media Substrat

**Catatan:** prosedur yang sama dilakukan untuk pembuatan media substrat 2% dan 3%

### c. Media Inokulum

0,1 g *Bacto* pepton,  
0,5 g yeast extract,  
dan 0,1 g NaCl

- ditimbang masing-masing di masukkan ke dalam Erlenmeyer 250 mL
- dilarutkan dengan 100 mL air laut steril
- disterilkan menggunakan *autoclav* pada suhu 121 °C selama 15 menit
- didiamkan hingga dingin

Media Inokulum

#### d. Media Produksi

0,5 g yeast extract, 0,1 bacto pepton, 0,1 g NaCl, 0,7 g  $(\text{NH}_4)_2\text{SO}_4$ , 0,1 g  $\text{KH}_2\text{PO}_4$ , 0,01 g  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , dan 0,05 g  $\text{CaCl}_2$ , dan 1 mL Substrat

- dimasukkan ke dalam Erlenmeyer 250 mL
- dilarutkan dengan 100 mL air laut steril
- disterilkan menggunakan *autoclav* pada suhu 121 °C selama 15 menit
- ↓ - didiamkan hingga dingin

Media Produksi

#### 3. Peremajaan Isolat Bakteri Laut

Isolat Bakteri Laut

- diambil satu ose
- digoreskan pada media peremajaan
- ↓ - diinkubasi pada suhu 37 °C selama 24 jam

Hasil

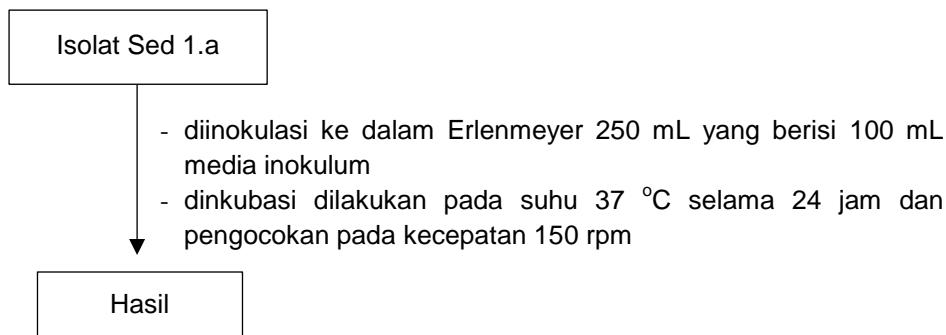
#### 4. Uji Kolagenolitik Bakteri Laut

Isolat Bakteri Laut

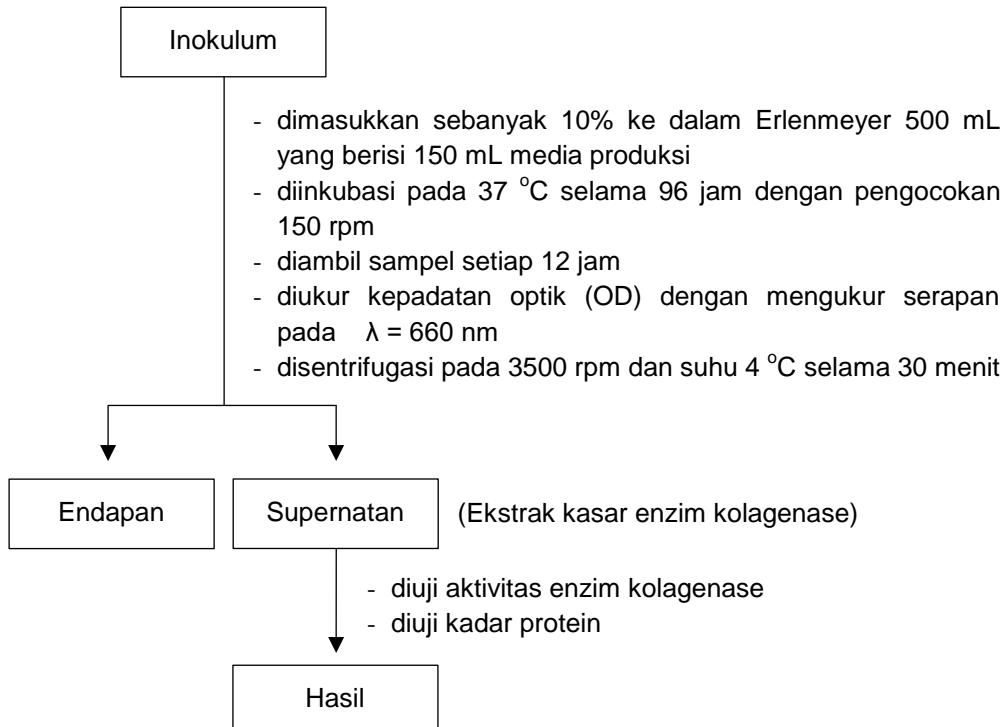
- diambil satu ose
- digoreskan pada media substrat dengan 3 konsentrasi berbeda
- ↓ - dinkubasi pada suhu 37 °C selama 3x24 jam
- diamati setiap 1x24 jam zona bening yang terbentuk

Hasil

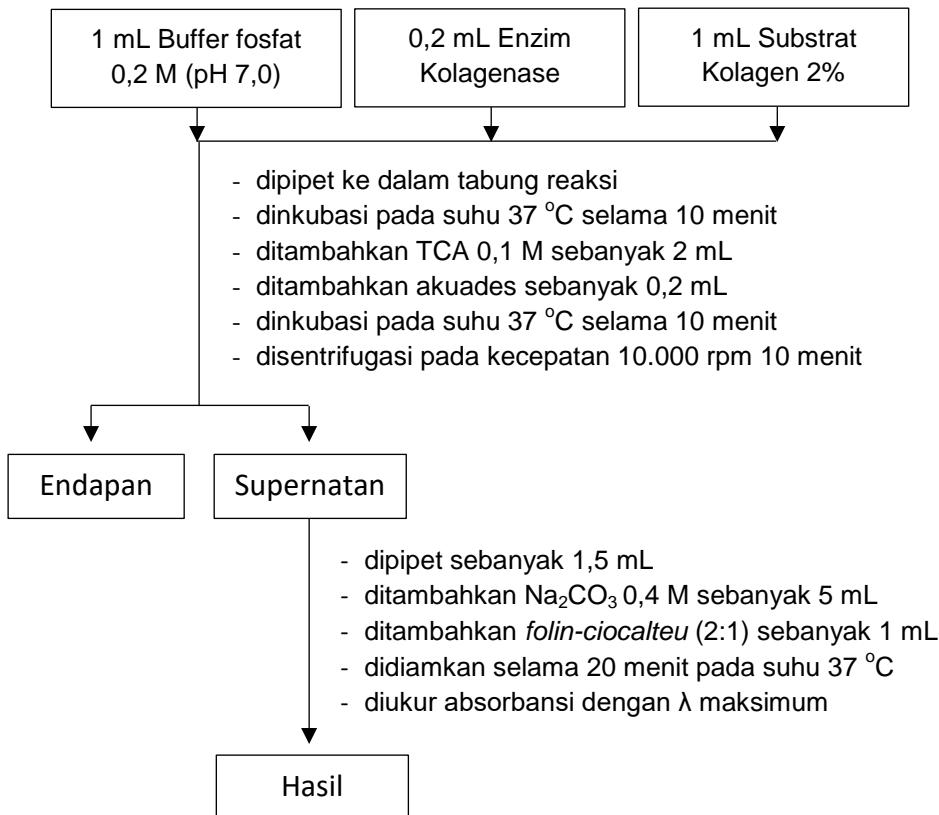
## 5. Persiapan Inokulum



## 6. Penentuan Waktu Produksi Optimum Enzim Kolagenase

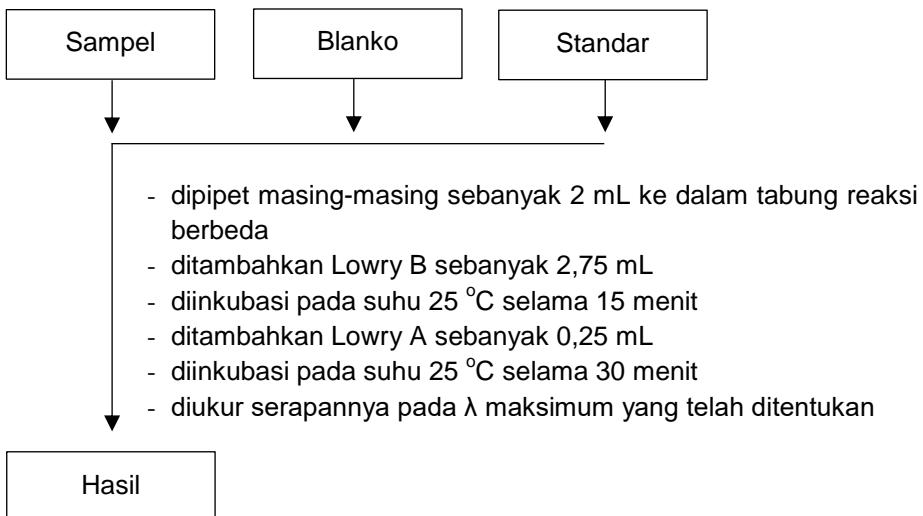


## 7. Penentuan Aktivitas Kolagenase

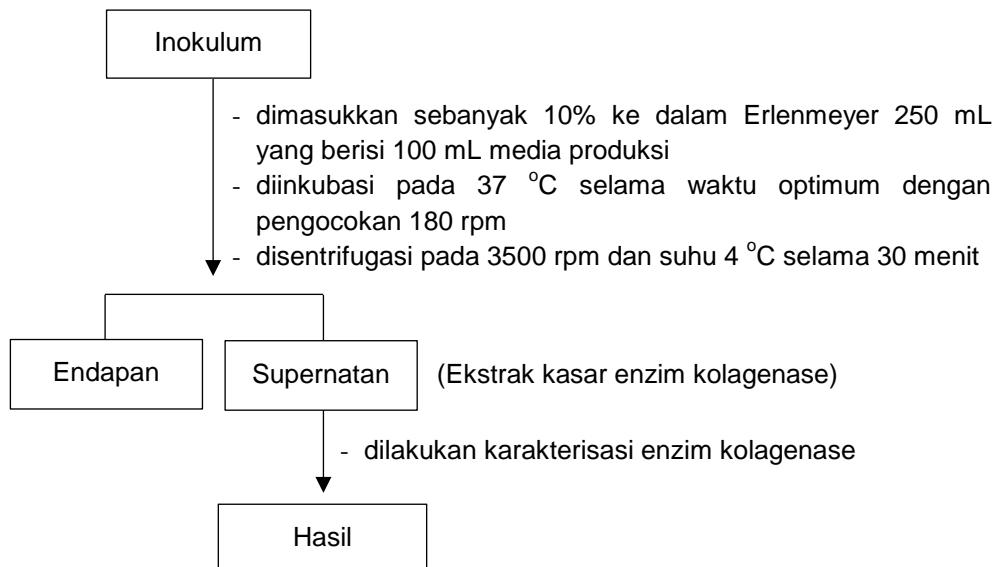


**Catatan:** Larutan tirosin digunakan sebagai larutan standar enzim kolagenase

## 8. Penentuan Kadar Protein

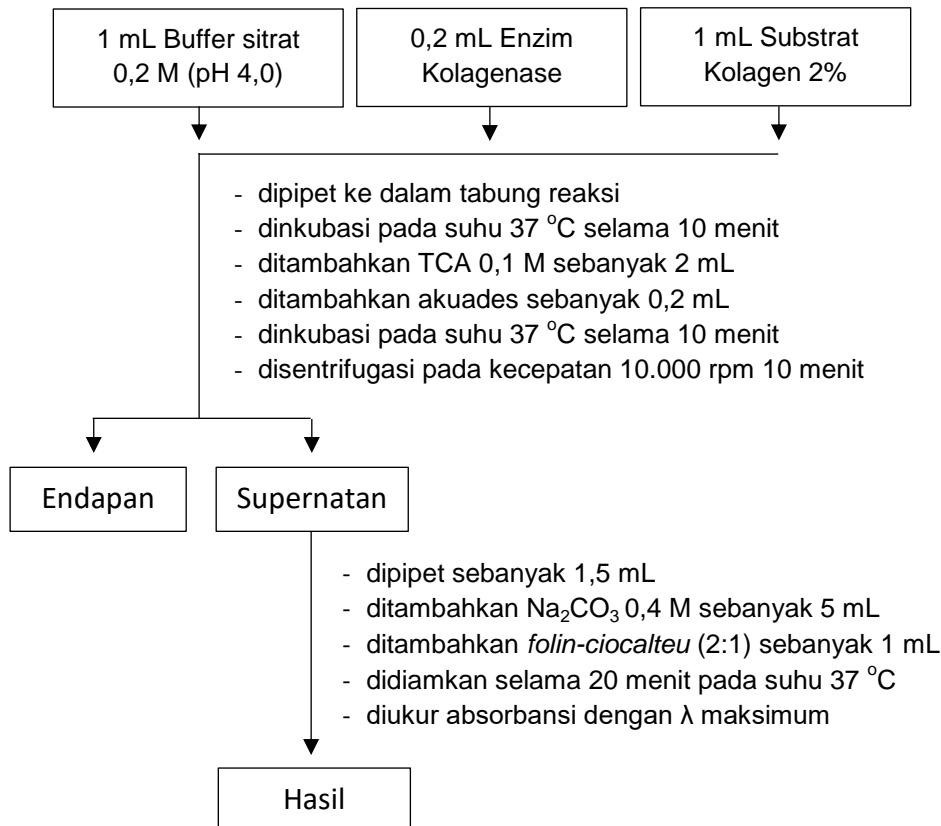


## 9. Produksi Enzim Kolagenase



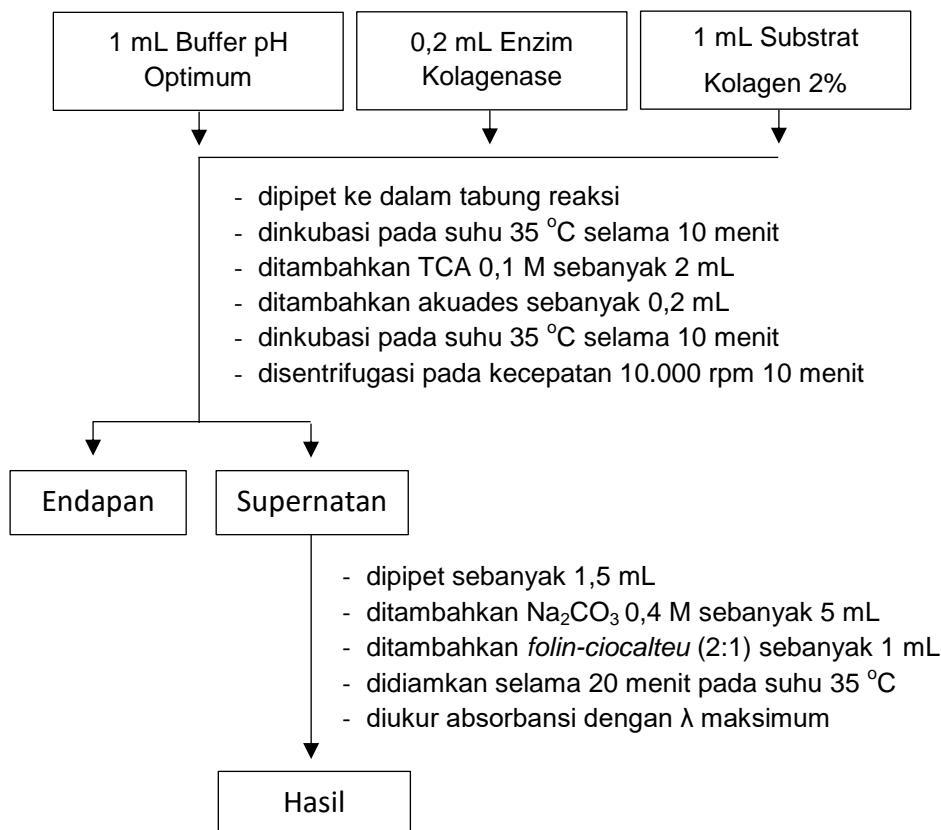
## 10. Karakterisasi Enzim Kolagenase

### a. Pengaruh pH Terhadap Aktivitas Enzim Kolagenase



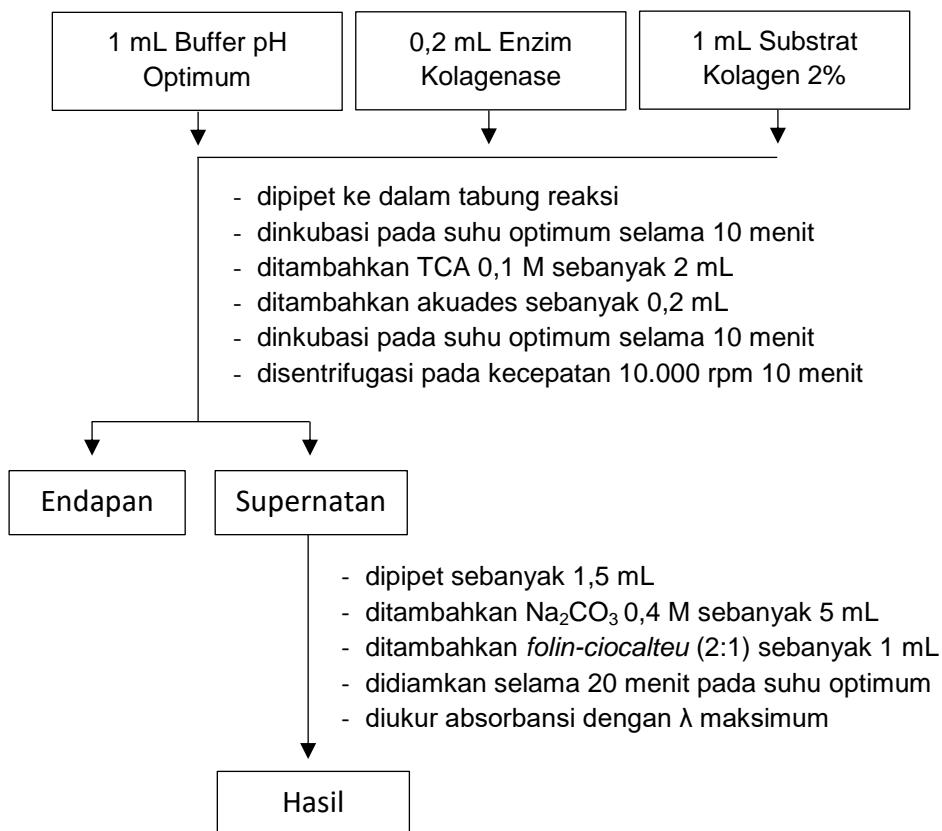
**Catatan:** Prosedur yang sama dilakukan untuk pH berbeda menggunakan larutan buffer sitrat 0,2 M (pH 4,0; 5,0; dan 6,0) dan buffer fosfat 0,2 M (pH 7,0; dan 8,0)

### b. Pengaruh Suhu Terhadap Aktivitas Enzim Kolagenase



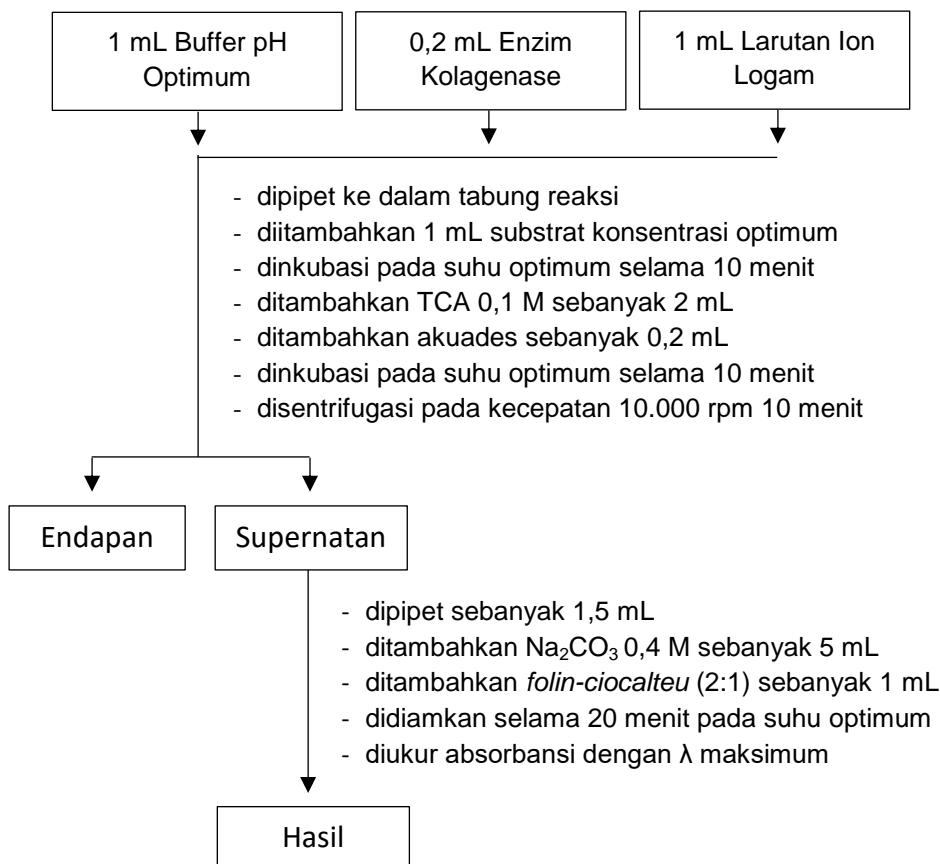
**Catatan:** Prosedur yang sama dilakukan pada variasi suhu (40, 45, 50, dan 55 °C)

### c. Pengaruh Konsentrasi Substrat Terhadap Aktivitas Enzim Kolagenase



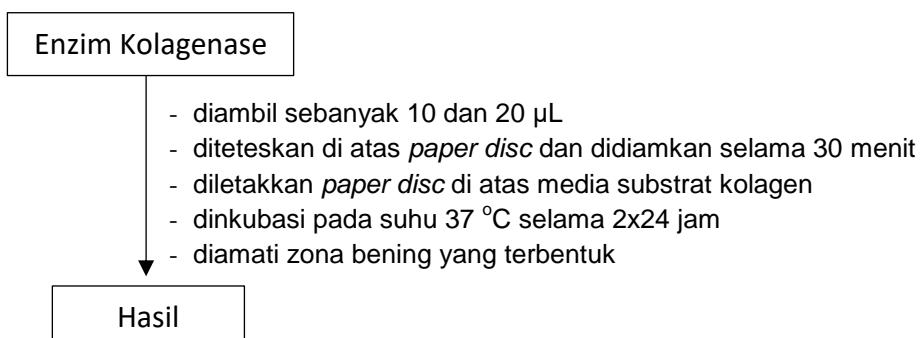
**Catatan:** Prosedur yang sama dilakukan pada variasi konsentrasi substrat 1%, 1,5%, 2,5% dan 3%.

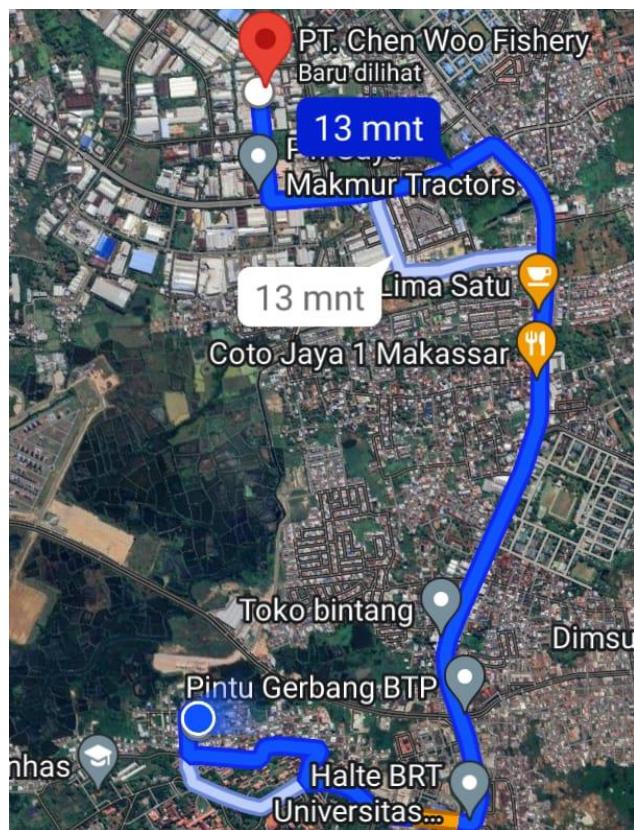
#### d. Pengaruh Ion Logam Terhadap Aktivitas Enzim Kolagenase



**Catatan:** Prosedur yang sama dilakukan pada variasi ion logam ( $K^+$ ,  $Na^+$ ,  $Ca^{2+}$ ,  $Mg^{2+}$ , dan  $Hg^{2+}$ ) dengan konsentrasi 5 dan 10 mM

#### 11. Hidrolisis Kolagen Kulit Ikan Tuna Sirip Kuning



**Lampiran 3.** Peta Lokasi Pengambilan Sampel

**Lampiran 4. Perhitungan Pembuatan Larutan**

**1. Pembuatan NaOH 0,05 M sebanyak 1000 mL**

$$\begin{aligned} g &= V \times M \times Mr \\ &= 1 \text{ L} \times 0,05 \text{ M} \times 40 \text{ g/mol} \\ &= 2 \text{ g} \end{aligned}$$

**2. Pembuatan CH<sub>3</sub>COOH 0,5 M sebanyak 1000 mL**

$$\begin{aligned} M &= \frac{\% \times bJ \times 1000}{Mr} \\ &= \frac{\% \times 1,05 \text{ g/cm}^3 \times 1000}{60 \text{ g/mol}} \\ &= 17,5 \text{ M} \end{aligned}$$

$$V_1 \times M_1 = V_2 \times M_2$$

$$V_1 \times 17,5 \text{ M} = 1000 \text{ mL} \times 0,5 \text{ M}$$

$$\begin{aligned} V_1 &= \frac{1000 \text{ mL} \times 0,5 \text{ M}}{17,5 \text{ M}} \\ V_1 &= 28,57 \text{ mL} \end{aligned}$$

**3. Pembuatan NaCl 2,6 M sebanyak 850 mL**

$$\begin{aligned} g &= V \times M \times Mr \\ &= 0,8 \text{ L} \times 2,6 \text{ M} \times 58,5 \text{ g/mol} \\ &= 121,68 \text{ g} \end{aligned}$$

**4. Pembuatan TCA 0,1 M sebanyak 200 mL**

$$\begin{aligned} g &= V \times M \times Mr \\ &= 0,2 \text{ L} \times 0,1 \text{ M} \times 163,4 \text{ g/mol} \\ &= 3,268 \text{ g} \end{aligned}$$

**5. Pembuatan Na<sub>2</sub>CO<sub>3</sub> 0,4 M sebanyak 500 mL**

$$\begin{aligned} g &= V \times M \times Mr \\ &= 0,5 \text{ L} \times 0,4 \text{ M} \times 106 \text{ g/mol} \\ &= 21,2 \text{ g} \end{aligned}$$

**6. Pembuatan Lowry A dan Lowry B**

a) **Lowry A**

5 mL folin ciocalteu : 5 mL akuades (1:1)

**b) Lowry B**

- **Pembuatan NaOH 0,1 N sebanyak 100 mL**

$$\begin{aligned} g &= V \times N \times Be \\ &= 0,1 \text{ L} \times 0,1 \text{ N} \times 40 \\ &= 0,4 \text{ g} \end{aligned}$$

- **Pembuatan Na<sub>2</sub>CO<sub>3</sub> 2% sebanyak 100 mL**

$$\begin{aligned} \% \text{ b/v} &= \frac{\text{g zat terlarut}}{\text{V larutan}} \times 100\% \\ 2\% &= \frac{x}{100 \text{ mL}} \times 100\% \end{aligned}$$

$$x = 2 \text{ g}$$

- **Pembuatan 50 mL Na-K-Tatrat 1%**

$$\begin{aligned} \% \text{ b/v} &= \frac{\text{g zat terlarut}}{\text{V larutan}} \times 100\% \\ 1\% &= \frac{x}{50 \text{ mL}} \times 100\% \end{aligned}$$

$$x = 0,5 \text{ g}$$

**7. Pembuatan Deret Standar**

**a) Konsentrasi 0,01 mg/mL**

$$\begin{aligned} V_1 \times M_1 &= V_2 \times M_2 \\ V_1 \times 0,1 \text{ mg/mL} &= 2 \text{ mL} \times 0,01 \text{ mg/mL} \\ V_1 &= \frac{2 \text{ mL} \times 0,01 \text{ mg/mL}}{0,1 \text{ mg/mL}} \\ V_1 &= 0,2 \text{ mL} \end{aligned}$$

Volume akuades = 2 mL - 0,2 mL = 1,8 mL

**b) Konsentrasi 0,02 mg/mL**

$$\begin{aligned} V_1 \times M_1 &= V_2 \times M_2 \\ V_1 \times 1 \text{ mg/mL} &= 2 \text{ mL} \times 0,02 \text{ mg/mL} \\ V_1 &= \frac{2 \text{ mL} \times 0,02 \text{ mg/mL}}{1 \text{ mg/mL}} \\ V_1 &= 0,04 \text{ mL} \end{aligned}$$

Volume akuades = 2 mL - 0,04 mL = 1,96 mL

**c) Konsentrasi 0,04 mg/mL**

$$\begin{aligned} V_1 \times M_1 &= V_2 \times M_2 \\ V_1 \times 1 \text{ mg/mL} &= 2 \text{ mL} \times 0,04 \text{ mg/mL} \\ V_1 &= \frac{2 \text{ mL} \times 0,04 \text{ mg/mL}}{1 \text{ mg/mL}} \end{aligned}$$

$$\begin{aligned} V_1 &= 0,08 \text{ mL} \\ \text{Volume akuades} &= 2 \text{ mL} - 0,08 \text{ mL} = 1,92 \text{ mL} \end{aligned}$$

**d) Konsentrasi 0,08 mg/mL**

$$\begin{aligned} V_1 \times M_1 &= V_2 \times M_2 \\ V_1 \times 1 \text{ mg/mL} &= 2 \text{ mL} \times 0,08 \text{ mg/mL} \\ V_1 &= \frac{2 \text{ mL} \times 0,08 \text{ mg/mL}}{1 \text{ mg/mL}} \\ V_1 &= 0,16 \text{ mL} \\ \text{Volume akuades} &= 2 \text{ mL} - 0,16 \text{ mL} = 1,84 \text{ mL} \end{aligned}$$

**e) Konsentrasi 0,16 mg/mL**

$$\begin{aligned} V_1 \times M_1 &= V_2 \times M_2 \\ V_1 \times 1 \text{ mg/mL} &= 2 \text{ mL} \times 0,16 \text{ mg/mL} \\ V_1 &= \frac{2 \text{ mL} \times 0,16 \text{ mg/mL}}{1 \text{ mg/mL}} \\ V_1 &= 0,32 \text{ mL} \\ \text{Volume akuades} &= 2 \text{ mL} - 0,32 \text{ mL} = 1,68 \text{ mL} \end{aligned}$$

**Lampiran 5. Dokumentasi Penelitian**

Pengambilan sampel



Preparasi sampel



Perendaman dengan NaOH

Perendaman dengan  $\text{CH}_3\text{COOH}$ 

Pembuatan media



Peremajaan bakteri laut



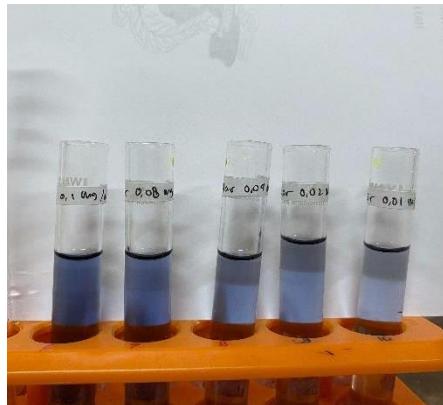
Hasil uji kolagenolitik substrat kolagen ikan tuna sirip kuning 1%, 2%, dan 3%



Produksi enzim di shaker waterbath



Ekstrak kasar kolagenase



Penentuan kadar protein



Karakterisasi ekstrak kasar enzim