DAFTAR PUSTAKA

1. Park J, Son H. Weapon Use in Korean Homicide: Differences Between Homicides Involving Sharp and Blunt Instruments. *J Forensic Sci.* 2018;63(4):1134-1137. http://doi.org/10.1111/1556-4029.13673

2. Vassalini M, Verzeletti A, De Ferrari F. Sharp force injury fatalities: A retrospective study (1982-2012) in Brescia (Italy). *J Forensic Sci.* 2014;59(6):1568-1574. http://doi.org/10.1111/1556-4029.12487

3. Verzeletti A, Bin P, De Ferrari F. Homicide by blunt trauma in Brescia County (Northern Italy) between 1982 and 2012. *Am J Forensic Med Pathol.* 2014;35(1):62-67. http://doi.org/10.1097/PAF.0000000000000080

4. Kemal CJ, Patterson T, Molina DK. Deaths due to sharp force injuries in Bexar County, Texas, with respect to manner of death. *Am J Forensic Med Pathol.* 2013;34(3):253-259. http://doi.org/10.1097/PAF.0b013e31828ced68

5. Mcewen T, Regoeczi W. Forensic Evidence in Homicide Investigations and Prosecutions. *J Forensic Sci.* 2015;60(5):1188-1198. http://doi.org/10.1111/1556-4029.12787

6. Brookman F, Innes M. The problem of success: What is a "good" homicide investigation? *Polic Soc.* 2013;23(3):292-310. http://doi.org/10.1080/10439463.2013.771538

7. Li F, Liu S, Lu X, Ou Y, Yip PSF. Application of the injury scales in homicides. *Forensic Sci Int.* 2018;292:83-89. http://doi.org/10.1016/j.forsciint.2018.09.010

8. Tamsen F, Logan FK, Thiblin I. Homicide Injury Quantification: Correlations and Reliability of Injury Severity Scores Applied to Homicide Victims. *Homicide Stud.* 2015;19(1):88-100. http://doi.org/10.1177/1088767914558142

9. Matzopoulos R, Prinsloo M, Pillay-Van Wyk V, et al. Injury-related mortality in south africa: A retrospective descriptive study of

postmortem investigations. *Bull World Health Organ.* 2015;93(5):303-313. http://doi.org/10.2471/BLT.14.145771

10. Maeda H, Zhu B li, Ishikawa T, Michiue T. Forensic molecular pathology of violent deaths. *Forensic Sci Int.* 2010;203(1-3):83-92. http://doi.org/10.1016/j.forsciint.2010.07.024

11. Li N, Du Q, Bai R, Sun J. Vitality and wound-age estimation in forensic pathology: review and future prospects. *Forensic Sci Res.* 2020;5(1):15-24. http://doi.org/10.1080/20961790.2018.1445441

12. Cecchi R. Estimating wound age: Looking into the future. *Int J Legal Med.* 2010;124(6):523-536. http://doi.org/10.1007/s00414-010-0505-x

13. Khalaf AA, Hassanen EI, Zaki AR, Tohamy AF, Ibrahim MA. Histopathological, immunohistochemical, and molecular studies for determination of wound age and vitality in rats. *Int Wound J.* 2019;16(6):1416-1425. http://doi.org/10.1111/iwj.13206

14. Kasuya A, Tokura Y. Attempts to accelerate wound healing. *J Dermatol Sci.* 2014;76(3):169-172. http://doi.org/10.1016/j.jdermsci.2014.11.001

15. Velnar T, Bailey T, Smrkolj V. The Wound Healing Process : an Overview of the Cellular and Molecular Mechanisms. Published online 2009. http://doi.org/10.1177/147323000903700531

16. Oehmichen M. Vitality and time course of wounds. *Forensic Sci Int.* 2004;144(2-3):221-231. http://doi.org/10.1016/j.forsciint.2004.04.057

17. Kondo T, Ishida Y. Molecular pathology of wound healing. *Forensic Sci Int.* 2010;203(1-3):93-98. http://doi.org/10.1016/j.forsciint.2010.07.004

18. Casse JM, Martrille L, Vignaud JM, Gauchotte G. Skin wounds vitality markers in forensic pathology: An updated review. *Med Sci Law.* 2016;56(2):128-137. http://doi.org/10.1177/0025802415590175

19. Peyron PA, Colomb S, Becas D, et al. Cytokines as new biomarkers of skin wound vitality. *Int J Legal Med.* 2021;135(6):2537-2545. http://doi.org/10.1007/s00414-021-02659-z

20. Gauchotte G, Wissler MP, Casse JM, et al. FVIIIra, CD15, and tryptase performance in the diagnosis of skin stab wound vitality in forensic pathology. *Int J Legal Med.* 2013;127(5):957-965. http://doi.org/10.1007/s00414-013-0880-1

21. El-fatah DSA. Cutaneous (tPA) and Skeletal (TnI) mRNA as Markers of Aging in Contused Wound. 2016;(April 2015):1-4. http://doi.org/10.1111/1556-4029.13072

22. Sun J, Wang Y, Zhang L. Time-dependent expression of skeletal muscle troponin I mRNA in the contused skeletal muscle of rats : a possible marker for wound age estimation. Published online 2010:27-33. http://doi.org/10.1007/s00414-009-0323-1

23. Matteis A De, Aquila M, Maiese A, et al. The Troponin-I fast skeletal muscle is reliable marker for the determination of vitality in the suicide hanging. *Forensic Sci Int.* 2019;301:284-288. http://doi.org/10.1016/j.forsciint.2019.05.055

24. Lin L, Hu K. Tissue plasminogen activator and inflammation: from phenotype to signaling mechanisms. *Am J Clin Exp Immunol.* 2014;3(1):30-36.
http://www.ncbi.nlm.nih.gov/pubmed/24660119%0Ahttp://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3960759

25. Zhang Q, Hearn SO, Ph D, Kavalukas SL, Barbul A. Role of High Mobility Group Box 1 (HMGB1) in Wound Healing. *J Surg Res.* 2012;176(1):343-347. http://doi.org/10.1016/j.jss.2011.06.069

26. Biology C, Melgarejo E, Ángel M, Sánchez-jiménez F, Luis J. The International Journal of Biochemistry Monocyte chemoattractant protein-1 : A key mediator in inflammatory processes. 2009;41:998-1001. http://doi.org/10.1016/j.biocel.2008.07.018

27. Bauer M. RNA in forensic science. *Forensic Sci Int Genet.* 2007;1(1):69-74. http://doi.org/10.1016/j.fsigen.2006.11.002

28. Sampaio-Silva F, Magalhães T, Carvalho F, Dinis-Oliveira RJ, Silvestre R. Profiling of RNA Degradation for Estimation of Post Morterm Interval. *PLoS One.* 2013;8(2).

http://doi.org/10.1371/journal.pone.0056507

29. Tozzo P, Scrivano S, Sanavio M, Caenazzo L. The role of DNA degradation in the estimation of post-mortem interval: A systematic review of the current literature. *Int J Mol Sci.* 2020;21(10). http://doi.org/10.3390/ijms21103540

30. Ali MM, Ibrahim SF, Fayed AA. Using Skin Gene Markers for Estimating Early Postmortem Interval at Different Temperatures. *Am J Forensic Med Pathol.* 2017;38(4):323-325. http://doi.org/10.1097/PAF.0000000000000337

31. Tao L, Ma J, Han L, et al. Early postmortem interval estimation based on Cdc25b mRNA in rat cardiac tissue. *Leg Med.* 2018;35(May):18-24. http://doi.org/10.1016/j.legalmed.2018.09.004

32. Wang H, Ma J, Xu H, et al. Early postmortem interval (EPMI) estimation using differentially expressed gene transcripts. *Leg Med.* 2019;38(April):83-91. http://doi.org/10.1016/j.legalmed.2019.04.008

33. Sturmberg JP, Martin CM. Systems and Complexity Thinking in the General Prac- tice Literature: An Integrative, Historical Narrative Review. Published online 2014:66-74. http://doi.org/10.1370/afm.1593.INTRODUCTION

34. Xue H. Applications of systems modelling in obesity research. Published online 2018:1-16. http://doi.org/10.1111/obr.12695

35. Rikken QGH, Chadid A, Peters J, Geeraedts LMG, Giannakopoulos GF, Tan ECTH. Epidemiology of penetrating injury in an urban versus rural level 1 trauma center in the Netherlands. Published online 2020. http://doi.org/10.1177/1024907920904190

36. Byun CS, Park IH, Oh JH, Bae KS, Lee KH, Lee E. Epidemiology of Trauma Patients and Analysis of 268 Mortality Cases : Trends of a Single Center in Korea. 2015;56(1):220-226.

37. Thomsen AH, Hougen HP, Sc DM, Villesen P, Ph D, Brink O. PAPER PATHOLOGY / BIOLOGY. Published online 2019:1-7. http://doi.org/10.1111/1556-4029.14244

38. Bianchi ME. DAMPs from cell death to new life. 2015;(September).

http://doi.org/10.3389/fimmu.2015.00422

39. Roh JS, Sohn DH. Damage-Associated Molecular Patterns in Inflammatory Diseases. 2018;18(4):1-14.

40. Rodrigues M, Kosaric N, Bonham CA, Gurtner GC. Wound healing: A cellular perspective. *Physiol Rev.* 2019;99(1):665-706. http://doi.org/10.1152/physrev.00067.2017

41. Straino S, Carlo A Di, Mangoni A, et al. High-Mobility Group Box 1 Protein in Human and Murine Skin : Involvement in Wound Healing. Published online 2008:1545-1553. http://doi.org/10.1038/sj.jid.5701212

42. Gomes A V, Potter JD, Szczesna-cordary D. The Role of Troponins in Muscle Contraction. Published online 2002:323-333. http://doi.org/10.1080/15216540290114676

43. Harper D, Young A, Mcnaught C. The physiology of wound healing. *Surgery.* 2014;32(9):445-450. http://doi.org/10.1016/j.mpsur.2014.06.010

44. Qing C. SC. *Chinese J Traumatol.* Published online 2017. http://doi.org/10.1016/j.cjtee.2017.06.001

45. Qu D, Tan X, Zhang K, Wang Q, Wang H. ATF3 mRNA , but not BTG2 , as a possible marker for vital reaction of skin contusion. *Forensic Sci Int.* 2019;303:109937. http://doi.org/10.1016/j.forsciint.2019.109937

46. Balandiz H, Pehlivan S, Çiçek AF, Tu H. Evaluation of Vitality in the Experimental Hanging Model of Rats by Using Immunohistochemical IL-1 β Antibody Staining. 2015;36(4):317-322. http://doi.org/10.1097/PAF.0000000000000199

47. Bai R, Wan L, Shi M. The time-dependent expressions of IL-1β, COX-2, MCP-1 mRNA in skin wounds of rabbits. *Forensic Sci Int.* 2008;175(2-3):193-197. http://doi.org/10.1016/j.forsciint.2007.07.006

48. He J, Huang H, Qu D, et al. CXCL1 and CXCR2 as potential markers for vital reactions in skin contusions. 2018;(1023).

49. Wang Q. IL-6 and IL-20 as potential markers for vitality of skin contusion. Published online 2018.

http://doi.org/10.1016/j.jflm.2018.07.010

50. Weis S, Llenos IC, Dulay JR, Elashoff M, Mart F, Miller CL. Quality control for microarray analysis of human brain samples : The impact of postmortem factors , RNA characteristics , and histopathology. 2007;165:198-209. http://doi.org/10.1016/j.jneumeth.2007.06.001

51. Pozhitkov AE, Noble PA. Insights & Perspectives Gene expression in the twilight of death. 2017;1700066:1-7. http://doi.org/10.1002/bies.201700066

52. Wen-can L, Kai-jun M, Ye-hui L, et al. Science and Justice Postmortem interval determination using 18S-rRNA and microRNA ☆. *Sci Justice.* Published online 2014:1-4. http://doi.org/10.1016/j.scijus.2014.03.001

53. Du Q xiang, Sun J hong, Zhang L yu, et al. Time-dependent expression of SNAT2 mRNA in the contused skeletal muscle of rats: A possible marker for wound age estimation. *Forensic Sci Med Pathol.* 2013;9(4):528-533. http://doi.org/10.1007/s12024-013-9482-y

54. Johnson G, Nolan T, Bustin SA. Chapter 1 Real-Time Quantitative PCR , Pathogen Detection and MIQE. 943:1-16. http://doi.org/10.1007/978-1-60327-353-4

55. Article M. System Dynamics Modeling in Health and Medicine : A Systematic Literature Review. 2020;(June 2019):1-45. http://doi.org/10.1002/sdr.1646

56. Malczynski LA. Best Practices for System Dynamics Model Design and Construction with Powersim Studio. 2011;(June).

57. Syawal P, Budu B, Hatta M, Massi MN, Ichsan AM, Minhajat R. Comparison between the triamcinolone and bevacizumab subconjunctivals and changes in Interleukin-1 mRNA expression in pterygium. *J Taibah Univ Med Sci.* 2022;17(1):67-71. http://doi.org/10.1016/j.jtumed.2021.07.009

58. Fachri M, Hatta M, Massi MN, et al. The strong correlation between ADAM33 expression and airway inflammation in chronic obstructive

pulmonary disease and candidate for biomarker and treatment of COPD. *Sci Rep.* 2021;11(1):1-13. http://doi.org/10.1038/s41598-021-02615-2

59. Farsida, Hatta M, Patellongi I, et al. The correlation of Foxp3 + gene and regulatory T cells with scar BCG formation among children with Tuberculosis. *J Clin Tuberc Other Mycobact Dis.* 2020;21:100202. http://doi.org/10.1016/j.jctube.2020.100202

60. Hatta M, Surachmanto EE, Islam AA, Wahid S. Expression of mRNA IL - 17F and sIL - 17F in atopic asthma patients. *BMC Res Notes.* Published online 2017:1-5. http://doi.org/10.1186/s13104-017-2517-9

61. Lu Q, Pan B, Bai H, Zhao W. ScienceDirect Intranuclear cardiac troponin I plays a functional role in regulating Atp2a2 expression in cardiomyocytes. 2021;(xxxx). http://doi.org/10.1016/j.gendis.2021.04.007

62. Kumar S, Shah S, Tang HM, Smith M, Borrás T, Danias J. Tissue plasminogen activator in trabecular meshwork attenuates steroid induced outflow resistance in mice. *PLoS One.* 2013;8(8):1-11. http://doi.org/10.1371/journal.pone.0072447

63. Schiller KR, Zillhardt MR, Alley J, Borjesson DL, Beitz AJ, Mauro LJ. tumor-bone coculture model. 2009;17:1-17. http://doi.org/10.1186/1471-2407-9-45

64. Article O. Systemic lidocaine inhibits high - mobility group box 1 messenger ribonucleic acid expression and protein in BALB / c mice after closed fracture musculoskeletal injury. Published online 2018:395-398. http://doi.org/10.4103/sja.SJA

65. Made L, Rusyati M, Kedokteran F, et al. Higher Treg FoxP3 and TGF-β mRNA Expression in Type 2 Reaction ENL (Erythema Nodosum Leprosum) Patients in Mycobacterium leprae Infection. 2020;(December). http://doi.org/10.2174/1874434602014010304

66. Hatta M, Massi MN, Natzir R, Ichsan AM, Sujuti H. The Effectiveness of Triamcinolone Injection on Risk of Postoperative Operations with the Conjunctiva Autograft Technique and Its Association with Change

of VEGF mRNA Expression. 2020;13(June):543-549.

67. Madea B, Grellner W, Kondo T. Vital Reactions and Wound Age Estimation. *Handb Forensic Med*. Published online 2014:237-252. http://doi.org/10.1002/9781118570654.ch18

68. Alshehhi S, Haddrill PR. Estimating time since deposition using quantification of RNA degradation in body fluid-specific markers. *Forensic Sci Int*. 2019;298:58-63. http://doi.org/10.1016/j.forsciint.2019.02.046

69. Kubo H, Hayashi T, Ago K, Ago M, Kanekura T. Temporal expression of wound healing-related genes in skin burn injury. *Leg Med*. 2014;16(1):8-13. http://doi.org/10.1016/j.legalmed.2013.10.002

70. Ye MY, Xu D, Liu JC, et al. IL-6 and IL-20 as potential markers for vitality of skin contusion. *J Forensic Leg Med*. 2018;59:8-12. http://doi.org/10.1016/j.jflm.2018.07.010

71. Ibrahim SF, Issak M, Bayoumy AA, Abd El-Fatah DS. Cutaneous (tPA) and Skeletal (TnI) mRNA as Markers of Aging in Contused Wound. *J Forensic Sci*. 2016;61(4):1007-1010. http://doi.org/10.1111/1556-4029.13072

72. Takamiya M, Saigusa K, Kumagai R, Nakayashiki N, Aoki Y. Studies on mRNA expression of tissue-type plasminogen activator in bruises for wound age estimation. *Int J Legal Med*. 2005;119(1):16-21. http://doi.org/10.1007/s00414-004-0453-4

73. Sun JH, Wang YY, Zhang L, Gao CR, Zhang LZ, Guo Z. Time-dependent expression of skeletal muscle troponin i mRNA in the contused skeletal muscle of rats: A possible marker for wound age estimation. *Int J Legal Med*. 2010;124(1):27-33. http://doi.org/10.1007/s00414-009-0323-1

74. Abd-Elhakim YM, Omran BHF, Ezzeldein SA, Ahmed AI, El-Sharkawy NI, Mohamed AA-R. Time-dependent expression of high-mobility group box-1 and toll-like receptors proteins as potential determinants of skin wound age in rats: Forensic implication. *Int J Legal Med*. 2022;136(6):1781-1789. http://doi.org/10.1007/s00414-022-02788-z