

Daftar Pustaka

1. Rothan HA, Byrareddy SN. The epidemiology and pathogenesis of coronavirus disease (COVID-19) outbreak. *J Autoimmun.* 2020;109(February):18–21.
2. Deng SQ, Peng HJ. Characteristics of and public health responses to the coronavirus disease 2019 outbreak in China. *J Clin Med.* 2020;9(2).
3. Tang Y, Liu J, Zhang D, Xu Z, Ji J, Wen C. Cytokine Storm in COVID-19: The Current Evidence and Treatment Strategies. *Front Immunol.* 2020;11(July):1–13.
4. Dan JM, Mateus J, Kato Y, Hastie KM, Yu ED, Faliti CE, et al. Immunological memory to SARS-CoV-2 assessed for up to 8 months after infection. *Science* (1979). 2021;371(6529):1–22.
5. United Nations. The Impact of COVID-19 on South-East Asia. *Policy Briefs.* 2020;1–29.
6. Ratcliffe R. First coronavirus cases confirmed in Indonesia amid fears nation is ill-prepared for outbreak [Internet]. *The Guardian.* 2020 [cited 2021 Oct 19]. Available from: <https://www.theguardian.com/world/2020/mar/02/first-coronavirus-cases-confirmed-in-indonesia-amid-fears-nation-is-ill-prepared-for-outbreak>
7. Abul K. Abbas. *Basic Immunology: Functions and Disorders of the Immune System*, Sixth Edition. *Journal of Chemical Information and Modeling.* 2020 p. 1689–1699.
8. Breton G, Mendoza P, Hägglöf T, Oliveira TY, Schaefer-Babajew D, Gaebler C, et al. Persistent cellular immunity to SARS-CoV-2 infection. *Journal of Experimental Medicine.* 2021;218(4).
9. Kwok SLL, Cheng SMS, Leung JNS, Leung K, Lee CK, Peiris JSM, et al. Waning antibody levels after COVID-19 vaccination with mRNA Comirnaty and inactivated CoronaVac vaccines in blood donors, Hong Kong, April 2020 to October 2021. *Eurosurveillance. European Centre for Disease Prevention and Control (ECDC);* 2022 Jan 13;27(2).
10. Pollard AJ, Bijker EM. A guide to vaccinology: from basic principles to new developments [Internet]. *Nat Rev Immunol* [Internet]. Available from: <https://doi.org/10.1038/>
11. Ophinni Y, Hasibuan AS, Widhani A, Maria S, Koesnoe S, Yuniastuti E, et al. COVID-19 Vaccines: Current Status and Implication for Use in Indonesia. *Acta Med Indones.* 2020;52(4):388–412.
12. Bradley T, Grundberg E, Selvarangan R, LeMaster C, Fraley E, Banerjee D, et al. Antibody Responses after a Single Dose of SARS-CoV-2 mRNA Vaccine. *New England Journal of Medicine. Massachusetts Medical Society;* 2021 Mar 23;384(20):1959–61.
13. McComb S, Thiriot A, Akache B, Krishnan L, Stark F. Introduction to the Immune System. *Methods in Molecular Biology.* 2019;2024:1–24.
14. Tan W, Lu Y, Zhang J, Wang J, Dan Y, Tan Z, et al. Viral kinetics and antibody responses in patients with COVID-19. *medRxiv.* 2020;
15. Galipeau Y, Greig M, Liu G, Driedger M, Langlois MA. Humoral Responses and Serological Assays in SARS-CoV-2 Infections. *Front Immunol.* 2020;11(December):1–19.
16. Zhang Y, Zeng G, Pan H, Li C, Hu Y, Chu K, et al. Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine in healthy adults aged 18–59 years: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial. *Lancet Infect Dis. Elsevier Ltd;* 2021;21(2):181–92.

17. Lumley SF, O'Donnell D, Stoesser NE, Matthews PC, Howarth A, Hatch SB, et al. Antibodies to SARS-CoV-2 are associated with protection against reinfection. medRxiv. 2020;2020.11.18.20234369.
18. Ge H, Wang X, Yuan X, Xiao G, Wang C, Deng T, et al. The epidemiology and clinical information about COVID-19. *European Journal of Clinical Microbiology and Infectious Diseases*. *European Journal of Clinical Microbiology & Infectious Diseases*; 2020;39(6):1011–9.
19. Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *The Lancet*. 2020;395(10223):507–13.
20. De Wit E, Van Doremalen N, Falzarano D, Munster VJ. SARS and MERS: Recent insights into emerging coronaviruses. *Nat Rev Microbiol*. Nature Publishing Group; 2016;14(8):523–34.
21. WHO. Novel Coronavirus (2019-nCoV) 22 January 2020. *WHO Bulletin*. 2020;(JANUARY):1–7.
22. Sugihantono. PEDOMAN PENCEGAHAN DAN PENGENDALIAN CORONAVIRUS DISEASE (COVID-19). 5th ed. Jakarta: Kementerian Kesehatan RI; 2020 Jul p. 1–214.
23. Organization WH. COVID-19 Weekly Epidemiological Update Edition 44, published 15 June 2021. 2021;(June):1–26.
24. Ministry of Health. COVID-19 Weekly Epidemiological Update. World Health Organization. 2021;(February):1–33.
25. Wu Z, McGoogan JM. Characteristics of and Important Lessons from the Coronavirus Disease 2019 (COVID-19) Outbreak in China: Summary of a Report of 72314 Cases from the Chinese Center for Disease Control and Prevention. *JAMA - Journal of the American Medical Association*. American Medical Association; 2020 p. 1239–42.
26. Burhan E, Sugihantono A, Aryati, Samuedro E, Rinawati W, Sitompul pompini agustina, et al. pedoman pencegahan dan pengendalian COVID-19. 5th ed. kementerian kesehatan RI; 2020 p. 1–214.
27. He W, Yi GY, Zhu Y. Estimation of the basic reproduction number, average incubation time, asymptomatic infection rate, and case fatality rate for COVID-19: Meta-analysis and sensitivity analysis. *J Med Virol*. 2020;92(11):2543–50.
28. Mavrodiev E V, Tursky ML, Mavrodiev NE, Ebach MC, Williams DM. On Classification and Taxonomy of Coronaviruses (Riboviria , Nidovirales , Coronaviridae) with special focus on severe acute respiratory syndrome-related. *bioRxiv preprint*. 2020;2(October):38.
29. Perlman S, Netland J. Coronaviruses post-SARS: Update on replication and pathogenesis. *Nat Rev Microbiol*. 2009;7(6):439–50.
30. R.Fehr A, Perlman S. Coronaviruses: An overview of their replication and pathogenesis. In: Maier helena jane, editor. *Coronaviruses: Methods and Protocols*. 2015 p. 1–282.
31. Maier H, Bickerton E, Britton P. *Coronaviruses: Methods and Protocols*. 2015 Jan 1 p. 1–282.
32. Zhang Q, Xiang R, Huo S, Zhou Y, Jiang S, Wang Q, et al. Molecular mechanism of interaction between SARS-CoV-2 and host cells and interventional therapy [Internet]. Available from: <https://doi.org/10.1038/s41392-021-00653-w>
33. Brochot E, Demey B, Touzé A, Belouzard S, Dubuisson J, Schmit JL, et al. Anti-spike, Anti-nucleocapsid and Neutralizing Antibodies in SARS-CoV-2 Inpatients and Asymptomatic Individuals. *Front Microbiol*. 2020;11(October):1–8.

34. Meyer B, Drosten C, Müller MA. Serological assays for emerging coronaviruses: Challenges and pitfalls. *Virus Res.* 2014;194:175–83.
35. Wang L, Xiang Y. Spike glycoprotein-mediated entry of sars coronaviruses. *Viruses.* 2020;12(11).
36. Wang L, Xiang Y. Spike glycoprotein-mediated entry of sars coronaviruses. *Viruses.* 2020;12(11).
37. Wrapp D, Wang N, Corbett KS, Goldsmith JA, Hsieh CL, Abiona O, et al. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science (1979).* 2020;367(6483):1260–3.
38. Bergmann CC, Silverman RH. COVID-19: Coronavirus replication, pathogenesis, and therapeutic strategies. *Cleve Clin J Med.* 2020;87(5):321–7.
39. Baker D, Amor S, Kang AS, Schmierer K, Giovannoni G. The underpinning biology relating to multiple sclerosis disease modifying treatments during the COVID-19 pandemic. *Mult Scler Relat Disord.* 2020;43(April).
40. Zhou G, Zhao Q. Perspectives on therapeutic neutralizing antibodies against the novel coronavirus sars-cov-2. *Int J Biol Sci.* 2020;16(10):1718–23.
41. Chowdhury MA, Hossain N, Kashem MA, Shahid MA, Alam A. Immune response in COVID-19: A review. *J Infect Public Health.* 2020 Nov;13(11):1619–29.
42. Ghaffari A, Meurant R, Ardakani A. COVID-19 serological tests: how well do they actually perform? *Diagnostics.* 2020;10(7):1–14.
43. Aprillya SR. Penyintas Covid Artinya Apa? Ini Penjelasannya [Internet]. *detiknews.* 2021 [cited 2021 Oct 19]. Available from: <https://news.detik.com/berita/d-5762316/penyintas-covid-artinya-apa-ini-penjelasannya>
44. World Health Organization. Criteria for releasing COVID-19 patients from isolation. *Scientific brief.* 2020;(17 June):1–5.
45. Kemenkes RI. Peraturan Menteri Kesehatan PMK Nomor 230 Tahun 2020 tentang Ruang Rawat Gabung. 2020;2019:1–46.
46. Amenta EM, Spallone A, Rodriguez-Barradas MC, El Sahly HM, Atmar RL, Kulkarni PA. Postacute COVID-19: An Overview and Approach to Classification. *Open Forum Infect Dis.* 2020 Dec 1;7(12):ofaa509.
47. Greenhalgh T, Knight M, A'Court C, Buxton M, Husain L. Management of post-acute covid-19 in primary care. *The BMJ.* 2020;370.
48. Hosseini A, Hashemi V, Shomali N, Asghari F, Gharibi T, Akbari M, et al. Innate and adaptive immune responses against coronavirus. *Biomedicine and Pharmacotherapy.* Elsevier Masson SAS; 2020;132:110859.
49. Tahaghoghi-Hajghorbani S, Zafari P, Masoumi E, Rajabinejad M, Jafari-Shakib R, Hasani B, et al. The role of dysregulated immune responses in COVID-19 pathogenesis. *Virus Res. Elsevier B.V.;* 2020;290(October):198197.
50. Chaari A, Bendriss G, Zakaria D, McVeigh C. Importance of Dietary Changes During the Coronavirus Pandemic: How to Upgrade Your Immune Response. *Front Public Health.* 2020;8(August):4–12.
51. Saad N, Moussa S. Immune response to COVID-19 infection: a double-edged sword. *Immunol Med.* Taylor & Francis; 2021;44(3):187–96.
52. Toor SM, Saleh R, Sasidharan Nair V, Taha RZ, Elkord E. T-cell responses and therapies against SARS-CoV-2 infection. *Immunology.* 2021;162(1):30–43.

53. Boechat JL, Chora I, Morais A, Delgado L. The immune response to SARS-CoV-2 and COVID-19 immunopathology – Current perspectives. *Pulmonology*. Sociedade Portuguesa de Pneumologia; 2021;27(5):423–37.
54. Brüssow H. Immunology of COVID-19. *Environ Microbiol*. 2020;22(12):4895–908.
55. Zhang G, Nie S, Zhang Z, Zhang Z. Longitudinal Change of Severe Acute Respiratory Syndrome Coronavirus 2 Antibodies in Patients with Coronavirus Disease 2019. *J Infect Dis*. Oxford University Press; 2020 Jun 29;222(2):183–8.
56. Carsetti R, Zaffina S, Piano Mortari E, Terreri S, Corrente F, Capponi C, et al. Different Innate and Adaptive Immune Responses to SARS-CoV-2 Infection of Asymptomatic, Mild, and Severe Cases. *Front Immunol*. 2020;11(December):1–16.
57. Chen Y, Zuiani A, Fischinger S, Mullur J, Atyeo C, Travers M, et al. Quick COVID-19 Healers Sustain Anti-SARS-CoV-2 Antibody Production. *Cell*. Elsevier Inc.; 2020;183(6):1496-1507.e16.
58. McAndrews KM, Dowlathshahi DP, Dai J, Becker LM, Hensel J, Snowden LM, et al. Heterogeneous antibodies against SARS-CoV-2 spike receptor binding domain and nucleocapsid with implications for COVID-19 immunity. *JCI Insight*. 2020;5(18):1–14.
59. Deming D, Sheahan T, Heise M, Yount B, Davis N, Sims A, et al. Vaccine efficacy in senescent mice challenged with recombinant SARS-CoV bearing epidemic and zoonotic spike variants. *PLoS Med*. 2006;3(12):2359–75.
60. Fialkowski A, Gernez Y, Arya P, Weinacht KG, Kinane TB, Yonker LM. Insight into the pediatric and adult dichotomy of COVID-19: Age-related differences in the immune response to SARS-CoV-2 infection. *Pediatr Pulmonol*. 2020;55(10):2556–64.
61. Bao Y, Ling Y, Chen Y-Y, Tian D, Zhao G-P, Zhang X-H, et al. Dynamic anti-spike protein antibody profiles in COVID-19 patients. *Int J Infect Dis*. 2021 Feb;103:540–8.
62. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, et al. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. *Clinical Infectious Diseases*. 2020;71(15):762–8.
63. Goodnow CC. COVID-19, varying genetic resistance to viral disease and immune tolerance checkpoints. *Immunol Cell Biol*. 2021;99(2):177–91.
64. Katona P, Katona-Apte J. The interaction between nutrition and infection. *Clinical Infectious Diseases*. 2008;46(10):1582–8.
65. Briguglio M, Pregliasco FE, Lombardi G, Perazzo P, Banfi G. The Malnutritional Status of the Host as a Virulence Factor for New Coronavirus SARS-CoV-2. *Front Med (Lausanne)*. 2020;7(April):1–5.
66. Calder PC, Carr AC, Gombart AF, Eggersdorfer M. Reply to “comment on: Optimal nutritional status for a well-functioning immune system is an important factor to protect against viral infections. *nutrients* 2020, 12, 1181.” *Nutrients*. 2020;12(8):1–3.
67. Soresina A, Moratto D, Chiarini M, Paolillo C, Baresi G, Focà E, et al. Two X-linked agammaglobulinemia patients develop pneumonia as COVID-19 manifestation but recover. *Pediatric Allergy and Immunology*. 2020;31(5):565–9.
68. Cao Y, Su B, Guo X, Sun W, Deng Y, Bao L, et al. Potent Neutralizing Antibodies against SARS-CoV-2 Identified by High-Throughput Single-Cell Sequencing of Convalescent Patients’ B Cells. *Cell*. 2020;182(1):73-84.e16.
69. Su S, Wong G, Shi W, Liu J, Lai ACK, Zhou J, et al. Epidemiology, Genetic Recombination, and Pathogenesis of Coronaviruses. *Trends Microbiol*. 2016;24(6):490–502.
70. Jia M, Wang X, Gong W, Zhong J, Leng Z, Ren L, et al. Humoral responses after inactivated COVID-19 vaccination in individuals with and without prior SARS-CoV-2

- infection: A prospective cohort study. *J Med Virol*. John Wiley and Sons Inc; 2022 Dec 1;94(12):5746–57.
71. Kaneko N, Kuo HH, Boucau J, Farmer JR, Allard-Chamard H, Mahajan VS, et al. Loss of Bcl-6-Expressing T Follicular Helper Cells and Germinal Centers in COVID-19. *Cell*. 2020;183(1):143-157.e13.
 72. Long QX, Liu BZ, Deng HJ, Wu GC, Deng K, Chen YK, et al. Antibody responses to SARS-CoV-2 in patients with COVID-19. *Nat Med*. 2020;26(6):845–8.
 73. Zhao J, Yuan Q, Wang H, Liu W, Liao X, Su Y, et al. Antibody Responses to SARS-CoV-2 in Patients with Novel Coronavirus Disease 2019. *Clinical Infectious Diseases*. 2020;71(16):2027–34.
 74. Goto A, Go H, Miyakawa K, Yamaoka Y, Ohtake N, Kubo S, et al. Sustained Neutralizing Antibodies 6 Months Following Infection in 376 Japanese COVID-19 Survivors. *Front Microbiol*. 2021;12(May):1–9.
 75. Peng P, Hu J, Deng H jun, Liu B zhong, Fang L, Wang K, et al. Changes in the humoral immunity response in SARS-CoV-2 convalescent patients over 8 months. *Cell Mol Immunol*. Springer US; 2021;18(2):490–1.
 76. Rao S, Lau A, So H-C. Exploring Diseases/Traits and Blood Proteins Causally Related to Expression of ACE2, the Putative Receptor of SARS-CoV-2: A Mendelian Randomization Analysis Highlights Tentative Relevance of Diabetes-Related Traits [Internet]. *Diabetes Care* [Internet]. 2020 May 19;43(7):1416–26. Available from: <https://doi.org/10.2337/dc20-0643>
 77. Fernandez C, Rysä J, Almgren P, Nilsson J, Engström G, Orho-Melander M, et al. Plasma levels of the proprotein convertase furin and incidence of diabetes and mortality [Internet]. *J Intern Med* [Internet]. John Wiley & Sons, Ltd; 2018 Oct 1;284(4):377–87. Available from: <https://doi.org/10.1111/joim.12783>
 78. Shang J, Wan Y, Luo C, Ye G, Geng Q, Auerbach A, et al. Cell entry mechanisms of SARS-CoV-2 [Internet]. *Proceedings of the National Academy of Sciences* [Internet]. *Proceedings of the National Academy of Sciences*; 2020 May 26;117(21):11727–34. Available from: <https://doi.org/10.1073/pnas.2003138117>
 79. Ejaz H, Alsrhani A, Zafar A, Javed H, Junaid K, Abdalla AE, et al. COVID-19 and comorbidities: Deleterious impact on infected patients [Internet]. *J Infect Public Health* [Internet]. 2020;13(12):1833–9. Available from: <https://www.sciencedirect.com/science/article/pii/S1876034120305943>
 80. Yang J, Zheng Y, Gou X, Pu K, Chen Z, Guo Q, et al. Prevalence of comorbidities and its effects in coronavirus disease 2019 patients: A systematic review and meta-analysis. *International Journal of Infectious Diseases*. Elsevier B.V.; 2020 May 1;94:91–5.
 81. Bhatraju PK, Ghassemieh BJ, Nichols M, Kim R, Jerome KR, Nalla AK, et al. Covid-19 in Critically Ill Patients in the Seattle Region — Case Series. *New England Journal of Medicine*. Massachusetts Medical Society; 2020 May 21;382(21):2012–22.
 82. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical Characteristics of 138 Hospitalized Patients with 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA - Journal of the American Medical Association*. American Medical Association; 2020 Mar 17;323(11):1061–9.
 83. Qiu H, Tong Z, Ma P, Hu M, Peng Z, Wu W, et al. Intensive care during the coronavirus epidemic. *Intensive Care Medicine*. Springer; 2020 p. 576–8.
 84. Yin Y, Wunderink RG. MERS, SARS and other coronaviruses as causes of pneumonia. *Respirology*. Blackwell Publishing; 2018 p. 130–7.

85. Wan Y, Shang J, Graham R, Baric RS, Li F. Receptor Recognition by the Novel Coronavirus from Wuhan: an Analysis Based on Decade-Long Structural Studies of SARS Coronavirus. *J Virol. American Society for Microbiology*; 2020 Mar 17;94(7).
86. Liu W, Tao ZW, Wang L, Yuan ML, Liu K, Zhou L, et al. Analysis of factors associated with disease outcomes in hospitalized patients with 2019 novel coronavirus disease. *Chin Med J (Engl). NLM (Medline)*; 2020 May 5;133(9):1032–8.
87. Zhao Q, Meng M, Kumar R, Wu Y, Huang J, Lian N, et al. The impact of COPD and smoking history on the severity of COVID-19: A systemic review and meta-analysis. *J Med Virol. John Wiley and Sons Inc.*; 2020 Oct 1;92(10):1915–21.
88. Chen WW, GLZWW et al. Chinacardiovascular diseases report 2018: an updated summary. *J Geriatr Cardiol.* 2020;17(1):1–8.
89. Fang L, KGRM. Are patients with hypertension and diabetes mellitus at increased risk for COVID-19 infection? *Lancet Respir Med.* 2020 Mar 11;8:e21.
90. Zheng YY, Ma YT, Zhang JY, Xie X. COVID-19 and the cardiovascular system. *Nature Reviews Cardiology. Nature Research*; 2020 p. 259–60.
91. Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *The Lancet. Lancet Publishing Group*; 2020 Mar 28;395(10229):1054–62.
92. Bonow RO, Fonarow GC, O’Gara PT, Yancy CW. Association of Coronavirus Disease 2019 (COVID-19) with Myocardial Injury and Mortality. *JAMA Cardiology. American Medical Association*; 2020 p. 751–3.
93. Uhlén M, Fagerberg L, Hallström BM, Lindskog C, Oksvold P, Mardinoglu A, et al. Tissue-based map of the human proteome. *Science (1979). American Association for the Advancement of Science*; 2015 Jan 23;347(6220).
94. Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *The Lancet. Lancet Publishing Group*; 2020 Feb 15;395(10223):507–13.
95. Jin X, Lian JS, Hu JH, Gao J, Zheng L, Zhang YM, et al. Epidemiological, clinical and virological characteristics of 74 cases of coronavirus-infected disease 2019 (COVID-19) with gastrointestinal symptoms. *Gut. BMJ Publishing Group*; 2020 Jun 1;69(6):1002–9.
96. Liu Y, Sun W, Li J, Chen L, Zhang L, Yu L. Title: Clinical features and progression of acute respiratory distress syndrome in Word count: 2,681 (including Research in context) 2 0 [Internet]. Available from: <https://doi.org/10.1101/2020.02.17.20024166>
97. Sun J, Zhu A, Li H, Zheng K, Zhuang Z, Chen Z, et al. Isolation of infectious SARS-CoV-2 from urine of a COVID-19 patient. *Emerging Microbes and Infections. Taylor and Francis Ltd.*; 2020 p. 991–3.
98. Chen YT, Shao SC, Lai ECC, Hung MJ, Chen YC. Mortality rate of acute kidney injury in SARS, MERS, and COVID-19 infection: A systematic review and meta-analysis. *Crit Care. BioMed Central*; 2020 Jul 16;24(1).
99. Zhu F, Cao Y, Xu S, Zhou M. Reply to Comments on ‘Co-infection of SARS-CoV-2 and HIV in a patient in Wuhan city, China.’ *Journal of Medical Virology. John Wiley and Sons Inc.*; 2020 p. 1417–8.
100. Martinez MA. Compounds with Therapeutic Potential against Novel Respiratory 2019 Coronavirus [Internet]. 2020 . Available from: <https://doi.org/10>

101. Day PM, Kines RC, Thompson CD, Jagu S, Roden RB, Lowy DR, et al. In vivo mechanisms of vaccine-induced protection against HPV infection. *Cell Host Microbe*. 2010 Sep 16;8(3):260–70.
102. Burton DR, Topol EJ. Toward superhuman SARS-CoV-2 immunity? *Nature Medicine*. Nature Research; 2021 p. 5–6.
103. Sanders B, Koldijk M, Schuitemaker H. Inactivated viral vaccines. In *Vaccine Analysis: Strategies, Principles, and Control*. Springer Berlin Heidelberg; 2015 Jan 1 p. 45–80.
104. Ura T, Okuda K, Shimada M. Developments in viral vector-based vaccines. *Vaccines*. MDPI AG; 2014 p. 624–41.
105. Iwasaki A, Yang Y. The potential danger of suboptimal antibody responses in COVID-19. *Nature Reviews Immunology*. Nature Research; 2020 p. 339–41.
106. Kutzler MA, Weiner DB. DNA vaccines: Ready for prime time? *Nature Reviews Genetics*. 2008 p. 776–88.
107. Pardi N, Hogan MJ, Porter FW, Weissman D. mRNA vaccines—a new era in vaccinology. *Nature Reviews Drug Discovery*. Nature Publishing Group; 2018 p. 261–79.
108. Liu G, Carter B, Gifford DK. Predicted Cellular Immunity Population Coverage Gaps for SARS-CoV-2 Subunit Vaccines and Their Augmentation by Compact Peptide Sets. *Cell Syst*. Cell Press; 2021 Jan 20;12(1):102-107.e4.
109. Amanat F, Stadlbauer D, Strohmeier S, O Nguyen TH, Chromikova V, McMahon M, et al. A serological assay to detect SARS-CoV-2 seroconversion in humans [Internet]. *Nat Med* [Internet]. Available from: <https://doi.org/10.1038/s41591-020-0913-5>
110. Chi X, Yan R, Zhang J, Zhang G, Zhang Y, Hao M, et al. A neutralizing human antibody binds to the N-terminal domain of the Spike protein of SARS-CoV-2.
111. Setiadi W, Rozi IE, Safari D, Ode W, Daningrat Id D, Id EJ, et al. Prevalence and epidemiological characteristics of COVID-19 after one year of pandemic in Jakarta and neighbouring areas, Indonesia: A single center study [Internet]. 2022; Available from: <https://doi.org/10.1371/journal.pone.0268241.g001>
112. Danielsen AC, Lee KM, Boulicault M, Rushovich T, Gompers A, Tarrant A, et al. Sex disparities in COVID-19 outcomes in the United States: Quantifying and contextualizing variation. *Soc Sci Med*. Elsevier Ltd; 2022 Feb 1;294.
113. Song H, Seddighzadeh B, Cooperberg MR, Huang FW. Expression of ACE2, the SARS-CoV-2 receptor, and TMPRSS2 in prostate epithelial cells [Internet]. Available from: <https://doi.org/10.1101/2020.04.24.056259>
114. Luo B, Rossato M, Vona R, Carlos Ramírez-Soto M, Peruana U, Heredia C, et al. Sex-differences in COVID-19 diagnosis, risk factors and disease comorbidities: A large US-based cohort study [Internet]. Available from: <https://covid19researchdatabase.org>
115. Takahashi T, Ellingson MK, Wong P, Israelow B, Lucas C, Klein J, et al. Sex differences in immune responses that underlie COVID-19 disease outcomes Overview of the study design [Internet]. *Nature* [Internet]. 2020;588:315. Available from: <https://doi.org/10.1038/s41586-020-2700-3>
116. Arnold CG, Libby A, Vest A, Hopkinson A, Monte AA. Immune mechanisms associated with sex-based differences in severe COVID-19 clinical outcomes. *Biology of Sex Differences*. BioMed Central Ltd; 2022.
117. Liu Y, Gayle AA, Wilder-Smith A, Rocklöv J. The reproductive number of COVID-19 is higher compared to SARS coronavirus. *J Travel Med*. Oxford University Press; 2020 Mar 1;27(2).

118. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, et al. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell*. Cell Press; 2020 Apr 16;181(2):271-280.e8.
119. Duque GA, Descoteaux A. Macrophage cytokines: Involvement in immunity and infectious diseases. *Frontiers in Immunology*. Frontiers Media S.A.; 2014.
120. Fialkowski A, Gernez Y, Arya P, Weinacht KG, Kinane TB, Yonker LM. Insight into the pediatric and adult dichotomy of COVID-19: Age-related differences in the immune response to SARS-CoV-2 infection. *Pediatric Pulmonology*. John Wiley and Sons Inc; 2020 p. 2556–64.
121. Selva KJ, van de Sandt CE, Lemke MM, Lee CY, Chua BY, Nguyen TH, et al. Distinct systems serology features in children, elderly and COVID patients 1 2 [Internet]. *Naphak 6 Modhiran* [Internet]. 11:17. Available from: <https://doi.org/10.1101/2020.05.11.20098459>
122. Dhar Chowdhury S, Oommen AM, Chowdhury D. Epidemiology of COVID-19 [Internet]. Available from: <https://doi.org/>
123. Zhou F, Yu T, Du R, Fan Guohui', Liu Ying. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *the lancet*. 2020 Mar 28;395:1054–62.
124. Iyer AS, Jones FK, Nodoushani A, Kelly M, Becker M, Slater D, et al. Persistence and decay of human antibody responses to the receptor binding domain of SARS-CoV-2 spike protein in COVID-19 patients. *Sci Immunol*. American Association for the Advancement of Science; 2020 Oct 16;5(52).
125. L'Huillier AG, Meyer B, Andrey DO, Arm-Vernez I, Baggio S, Didierlaurent A, et al. Antibody persistence in the first 6 months following SARS-CoV-2 infection among hospital workers: a prospective longitudinal study. *Clinical Microbiology and Infection*. Elsevier B.V.; 2021 May 1;27(5):784.e1-784.e8.
126. Ko JH, Müller MA, Seok H, Park GE, Lee JY, Cho SY, et al. Serologic responses of 42 MERS-coronavirus-infected patients according to the disease severity. *Diagn Microbiol Infect Dis*. Elsevier Inc.; 2017 Oct 1;89(2):106–11.
127. Costa Clemens SA, Weckx L, Clemens R, Almeida Mendes AV, Ramos Souza A, Silveira MBV, et al. Heterologous versus homologous COVID-19 booster vaccination in previous recipients of two doses of CoronaVac COVID-19 vaccine in Brazil (RHH-001): a phase 4, non-inferiority, single blind, randomised study. *The Lancet*. Elsevier B.V.; 2022 Feb 5;399(10324):521–9.
128. Zeng G, Wu Q, Pan H, Li M, Yang J, Wang L, et al. Immunogenicity and safety of a third dose of CoronaVac, and immune persistence of a two-dose schedule, in healthy adults: interim results from two single-centre, double-blind, randomised, placebo-controlled phase 2 clinical trials. *Lancet Infect Dis*. Elsevier Ltd; 2022 Apr 1;22(4):483–95.
129. Gudbjartsson DF, Norddahl GL, Melsted P, Gunnarsdottir K, Holm H, Eythorsson E, et al. Humoral Immune Response to SARS-CoV-2 in Iceland. *New England Journal of Medicine*. Massachusetts Medical Society; 2020 Oct 29;383(18):1724–34.
130. Rode OĐ, Bodulić K, Zember S, Balent NC, da Novokmet A, Čulo M, et al. Decline of Anti-SARS-CoV-2 IgG Antibody Levels 6 Months after Complete BNT162b2 Vaccination in Healthcare Workers to Levels Observed Following the First Vaccine Dose. *Vaccines* (Basel). MDPI; 2022 Feb 1;10(2).
131. Ma ML, Shi DW, Li Y, Hong W, Lai DY, Xue JB, et al. Systematic profiling of SARS-CoV-2-specific IgG responses elicited by an inactivated virus vaccine identifies peptides and

- proteins for predicting vaccination efficacy. *Cell Discov.* Springer Nature; 2021 Dec 1;7(1).
132. Zhang J, Xing S, Liang D, Hu W, Ke C, He J, et al. Differential Antibody Response to Inactivated COVID-19 Vaccines in Healthy Subjects. *Front Cell Infect Microbiol.* Frontiers Media S.A.; 2021 Dec 16;11.
 133. Badano MN, Sabbione F, Keitelman I, Pereson M, Aloisi N, Colado A, et al. Humoral response to the BBIBP-CorV vaccine over time in healthcare workers with or without exposure to SARS-CoV-2. *Mol Immunol.* Elsevier Ltd; 2022 Mar 1;143:94–9.
 134. Zhang Y, Zeng G, Pan H, Li C, Hu Y, Chu K, et al. Safety, tolerability, and immunogenicity of an inactivated SARS-CoV-2 vaccine in healthy adults aged 18–59 years: a randomised, double-blind, placebo-controlled, phase 1/2 clinical trial. *Lancet Infect Dis.* Lancet Publishing Group; 2021 Feb 1;21(2):181–92.
 135. Ebinger JE, Fert-Bober J, Printsev I, Wu M, Sun N, Prostko JC, et al. Antibody responses to the BNT162b2 mRNA vaccine in individuals previously infected with SARS-CoV-2. *Nat Med.* Nature Research; 2021 Jun 1;27(6):981–4.
 136. Jiang S, Hillyer C, Du L. Neutralizing Antibodies against SARS-CoV-2 and Other Human Coronaviruses [Internet]. Available from: <https://biorender.com/>
 137. Tan CW, Chia WN, Qin X, Liu P, Chen MIC, Tiu C, et al. A SARS-CoV-2 surrogate virus neutralization test based on antibody-mediated blockage of ACE2–spike protein–protein interaction. *Nat Biotechnol.* Nature Research; 2020 Sep 1;38(9):1073–8.
 138. Wang H, Zhang Y, Huang B, Deng W, Quan Y, Wang W, et al. Development of an Inactivated Vaccine Candidate, BBIBP-CorV, with Potent Protection against SARS-CoV-2. *Cell.* Cell Press; 2020 Aug 6;182(3):713–721.e9.
 139. Chandrashekar A, Liu J, Martinot AJ, McMahan K, Mercado NB, Peter L, et al. SARS-CoV-2 infection protects against rechallenge in rhesus macaques [Internet]. Available from: <https://www.science.org>
 140. Nicola C. Poor correlation between antibody titers and neutralizing activity in sera from SARS-CoV-2 infected subjects [Internet]. Available from: <https://doi.org/10.1101/2020.07.10.20150375>
 141. Grzelak L, Temmam S, Planchais C, Demeret C, Tondeur L, Huon C, et al. Mireille Nowakowski 15 , Marija Backovic 16 [Internet]. Sandrine Pellerin-Fernandes. 2020 . Available from: <https://www.science.org>
 142. Chi X, Yan R, Zhang J, Zhang G, Zhang Y, Hao M, et al. A neutralizing human antibody binds to the N-terminal domain of the Spike protein of SARS-CoV-2.
 143. Favresse J, Gillot C, Di Chiaro L, Eucher C, Elsen M, Van Eeckhoudt S, et al. Neutralizing Antibodies in COVID-19 Patients and Vaccine Recipients after Two Doses of BNT162b2 [Internet]. 2021; Available from: <https://doi.org/10.3390/v13071364>
 144. Galipeau Y, Greig M, Liu G, Driedger M, Langlois MA. Humoral Responses and Serological Assays in SARS-CoV-2 Infections. *Front Immunol.* 2020;11(December):1–19.

DAFTAR LAMPIRAN

Lampiran 1. Elisa

Alat dan Bahan:

- 96-well microplate (Corning, #3590)
- Bicarbonate buffer (1.59 g of Na₂CO₃ + 2.93 g of NaHCO₃ per 1 l)
- PBS (-)
- PBS-T (PBS, 0.05% Tween 20)
- 1% BSA in PBS
- Cit/phos buffer (0.1 M citrate, 0.2 M disodium phosphate, pH 4.4-4.6)
- SARS-CoV-2 recombinant S1 (Sino Biological, #40591-V08H)
- SARS-CoV-2 recombinant RBD (GenScript, #Z03483)
- Heat-inactivated serum samples (56°C, 30 min)
- Representative serum samples as inter-day reliability control
- Anti-human IgG HRP conjugate (Bio-Rad, #172-1050)
- ABTS (Merck, #A1888)
- H₂O₂

Prosedur:

(Day 0)

1. Coat the microplate with 0.2 µg/50 µl/well of recombinant protein in bicarbonate buffer.
2. Incubate overnight at 4°C.

(Day 1)

3. Aspirate the bicarbonate buffer in the microplate.
4. Add 100 µl/well of 1% BSA in PBS and incubate for 1 h at room temperature (25°C).
5. Dilute serum samples with 1% BSA in PBS (1:100 for S1 and RBD). (These dilution factors roughly correspond to the EC₅₀ values for the RayBiotech patient samples)
6. Aspirate the blocking buffer in the microplate.
7. Add 50 µl/well of the diluted samples (duplicates) and incubate for 1 hour at room temperature.
8. Wash the microplate three times with PBS-T and once with PBS.
9. Add 50 µl/well of the diluted anti-human IgG HRP conjugates (1:2000 in 1% BSA/PBS) and incubate for 1 h at room temperature.
10. Prepare the substrate as follows:
 - ABTS: 4 mg
 - cit/phos buffer: 10 ml
 - H₂O₂: 6 µl (add just before use)
11. Wash the microplate three times with PBS-T and once with PBS.
12. Add 100 µl/well of the substrate and incubate for 30 min (precisely, if possible) at room temperature.
13. Measure the absorbance at 414 nm on a microplate reader.

Lampiran 2. Neutralizing Antibody Baculovirus Immunized Sera (Mice)

1. Add DMEM-CM (96 well)

Control 2020C 2003							DMEM	Serum		7-10 times pipetting
	#1	#2	#5	#6	#9	#10				
A							260µl	10,8µl	90µl	
B							180µl		90µ	
C									90µl	
D									90µl	
E									90µl	

2. Add 10,8µl serum in "A"
3. serial dilution (90µl ea)
4. prepare white plate:
A1 B1 C1: 100µl DMEM
A2 B2 C2: 50µl DMEM
5. SAG-LUC VSV
200ml/ 5ml DMEM in 15ml falcon tube
6. 50µl/ well VSV (w/o A1 B1 C1)
7. add diluted serum: 50µl/ well
8. 37C 1 hour
9. 293 T/ h ACE2 + hTMPRSS2
Cell number 200 x2/4= 10⁶/well
Requirement: 2x10⁵/ml
Dilution: 1:5 2 2mol cell + 9ml B
10. 100µl cells/well
11. incubated 37C over night
12. lysis reagent 400µl in 1,7ml DW
13. discard medium --> A/C
14. 20µl/ well lysis reagent
15. luceferase assay by Glomax

Neutralizing activity of the serum was examined, using a VSV-based pseudovirus, as previously describe. Briefly, the pseudovirus was engineered to express the Wuhan-Hu-1 SARS-CoV-2 Spike protein on the viral surface, in which the luciferase gene was incorporated in the viral genome. The serum was diluted with the medium, and the virus was added by triplicate. Final dilution rate of the serum was 1:100. The mixture of the virus and the serum was incubated with human embryonic kidney (HEK) 293T cells that expressed human ACE2 and human TMPRSS2. The cells were examined by Luciferase Assay System (Promega, Madison, WI, USA) after 24 hours incubation.

Lampiran 3. Form Anamnesis dan Data Pasien

FORM ANAMNESIS DAN DATA PASIEN

ID Pasien :

NO	Pertanyaan	Catatan
1	Nama	
2	Umur	
3	Jenis Kelamin	
4	Alamat	
5	Pekerjaan	
6	Suku	
7	Agama	
8	Tanggal masuk RS/Klinik	
9	Tanggal wawancara	
10	Keluhan Utama	
11	Riwayat Penyakit: <ul style="list-style-type: none"> - Kapan terdiagnosis? - Dimana? - Riwayat kontak? - Demam? Berapa lama? Frekuensi? Waktu tertentu? Tipe demam (continus, intermitten) - Banyak berkeringat? - Sakit kepala? - Nyeri otot? Di daerah mana? - Gusi bengkak? - Batuk (kering/berdahak)? - Sesak napas? - Nyeri menelan - Nyeri di bagian dada - Penurunan penciuman (bau) - Penurunan pengecap - Mual/muntah - Diare - Keluhan lain : ... - Riwayat rawat inap 	
12	Riwayat penyakit sebelumnya? <ul style="list-style-type: none"> - Riwayat saluran napas - Riwayat alergi (atopi) - Komorbid lain DM/hipertensi - Riwayat malaria 	
13	Riwayat keluarga <ul style="list-style-type: none"> - Apakah ada anggota keluarga yang juga positif COVID 19? - Kapan terakhir kontak dengan penderita sebelum sakit? 	
14	Riwayat Pengobatan <ul style="list-style-type: none"> - Obat apa saja yang telah dikonsumsi? 	

- Penggunaan vitamin D
- Riwayat berjemur selama sakit? Frekuensi?
Durasi?

Apakah dirawat di ICU? Berapa lama?

15 Riwayat Vaksinasi

- COVID 19 kapan?
- Smallpox (cacar) ya/tidak?

16 Keluhan setelah sembuh:

- Apakah ada keluhan yang masih dirasakan?
- Atau ada keluhan lain yang kira-kira berkaitan?

Lampiran 4. Case Report Form

CASE REPORT FORM

ID Pasien :

No	Pemeriksaan	Tanggal	Hasil
1	Anamnesis: <ul style="list-style-type: none"> - Keluhan utama - Riwayat penyakit sekarang - Riwayat penyakit dahulu - Riwayat Keluarga 		
2	Pemeriksaan fisik: <ul style="list-style-type: none"> - Keadaan umum - TB/BB - Pemeriksaan kepala-kaki 		
3	Pemeriksaan sampel darah <ul style="list-style-type: none"> - Reaktivitas terhadap protein spike dan nukleokapsid SARS-CoV-2 - Kadar antibody dalam serum 		
4	Pemeriksaan penunjang lainnya Radiologi: GGO? Pneumonia? Lab Darah: Trombositopeni? NLR?		

Lampiran 5. Informed Consent

1. PERSETUJUAN SESUDAH PENJELASAN (PSP) untuk orang dewasa (>16 tahun)**Bagian I. Lembar Informasi / Penjelasan kepada Volunteer**

Nama saya, bekerja di

Kami sedang melakukan riset mengenai penyakit infeksi yang disebabkan oleh virus korona. Penyakit ini biasanya ditandai dengan gejala demam, batuk, sesak napas, hilangnya rasa pengecap dan penciuman, dan berbagai gejala lainnya.

Kami akan memberikan informasi kepada Bapak/Ibu dan mengundang Bapak/Ibu jika ingin berpartisipasi pada riset ini. Sebelum memutuskan untuk berpartisipasi, Bapak/Ibu dapat berdiskusi dengan seseorang yang Bapak/Ibu percaya. Apabila terdapat kata-kata yang tidak dipahami, silakan bertanya ke saya, dan saya akan menyediakan waktu untuk menjelaskannya. Selain itu jika nanti Bapak/Ibu memiliki pertanyaan, dapat ditanyakan hal tersebut kepada saya, dokter atau petugas yang ditunjuk.

Keikutsertaan Bapak/Ibu dalam riset ini sepenuhnya sukarela. Kalau Bapak/Ibu memilih untuk tidak ikut, Bapak/Ibu tetap mendapatkan layanan kesehatan seperti biasanya. Bahkan, meskipun Bapak/Ibu setuju saat ini, tetapi kemudian berubah pikiran dan menarik persetujuan keikutsertaan Bapak/Ibu, maka layanan yang anda terima dari rumah sakit (RS) di daerah ini tetap dilanjutkan.

Selama masa riset, darah akan diambil 1 kali, sebanyak +/- 3 ml menggunakan jarum suntik kecil. Hal ini mungkin akan menimbulkan rasa sakit, namun akan kami minimalisir dengan teknik yang sebaik-baiknya. Dalam pengambilan darah ini, kami akan melakukan prosedur bebas-hama untuk menghindarkan Bapak/Ibu dari efek samping infeksi akibat pengambilan darah tersebut dengan menguapkan kapas alcohol sebelum dan setelah memasukkan jarum suntik. Darah Bapak/Ibu akan kami cek apakah mengandung antibody terhadap virus korona. Hal ini penting diketahui karena jika kadar antibody dalam darah Bapak/Ibu tinggi, maka akan memberikan perlindungan terhadap infeksi virus korona di kemudian hari. Namun, jika kadarnya terus menurun, maka data ini akan sangat bermanfaat untuk pembuatan dan pengembangan vaksin. Dengan adanya vaksin, dapat memicu terbentuknya antibody dalam jangka waktu tertentu untuk menghasilkan mekanisme perlindungan dan pertahanan tubuh yang baik terhadap virus korona.

Penerjemah: Saya telah menanyakan Bapak/Ibu bahwa keikutsertaan pada penelitian ini sepenuhnya sukarela:(inisial).

Jika Bapak/Ibu memutuskan untuk ikut dalam riset ini, setiap kesakitan yang berkaitan dengan penelitian ini, maka Bapak/Ibu dapat menghubungi dr. Sitti Nurisyah, Sp.P di nomor telepon: 081230308383.

Kami tidak akan memberitahukan identitas Bapak/Ibu di riset ini pada orang lain. Informasi yang kami kumpulkan pada riset ini akan dijaga kerahasiannya. Setiap informasi yang diperoleh tentang Bapak/Ibu akan diberi kode tertentu, tanpa disertai nama Bapak/Ibu. Hanya anggota peneliti yang mengetahui kode Bapak/Ibu dan kami akan menyimpan informasi tersebut dalam laci yang terkunci. Kami akan membagi pengetahuan yang kami peroleh dari riset ini kepada Bapak/Ibu sebelum diumumkan ke public. Informasi rahasia tidak akan dipaparkan. Kami akan menyelenggarakan pertemuan kecil dengan partisipan dan hal ini akan diumumkan. Setelah itu, kami akan mempublikasikan hasil ini dan membuat sedemikian rupa sehingga hasil riset ini dapat berguna bagi orang lain.

Riset ini telah dibahas oleh Komisi Etik Penelitian Kesehatan, Fakultas Kedokteran Universitas Hasanuddin. Kalau saudara ingin mendapat informasi lebih rinci tentang komisi etik tersebut, saudara dapat menghubungi dr. Agussalim Bukhari, M.Med, Ph.D, Sp.GK, Sekretaris Komisi Etik, di nomor: 0411-5780103; 081225704670

Bagian II: Surat Persetujuan Keikutsertaan

Saya telah membaca informasi di atas, atau informasi tersebut telah dibacakan kepada saya. Saya telah diberi kesempatan bertanya dan setiap pertanyaan yang saya ajukan telah dijawab dengan memuaskan. Saya menyetujui secara sukarela untuk berpartisipasi dalam riset ini.

- _Nama Sukarelawan :
- _Tanda tangan :
- _Tanggal (Hari/bulan/tahun)

Tanda tangan Saksi : (Tanda tangan Saksi dan cap jari sukarelawan hanya diperlukan bila sukarelawan buta huruf. Jika memungkinkan, saksi adalah seseorang yang dipilih oleh partisipan dan tidak memiliki hubungan dengan tim riset ini).

Saya telah menyaksikan pembacaan secara akurat dari lembar PSP ini ke orang yang mungkin dapat berpartisipasi, yang telah memiliki kesempatan untuk mengajukan pertanyaan. Saya menegaskan bahwa Partisipan telah memberikan persetujuan secara sukarela.

- _Nama Saksi :
- _Tanda tangan Saksi :
- _Tanggal (Hari/bulan/tahun)

Cap Jempol Partisipan :

Tanda Tangan Peneliti : Saya telah membacakan atau bersaksi mendengarkan pembacaan lembar informasi PSP pada calon partisipan yang memiliki kesempatan untuk bertanya. Saya menegaskan bahwa partisipan telah memberikan persetujuannya secara sukarela

- _Nama Peneliti :
- _Tanda tangan Peneliti :
- _Tanggal (Hari/bulan/tahun)

Salinan dari lembar persetujuan ini akan diberikan kepada Partisi

Lampiran 6. Hasil Analisis Normalitas Data S1, RBD dan NP

Tests of Normality^{b,d,e,f,g,h,i,j,k}

	group	Kolmogorov-Smirnov ^a			Shapiro-Wilk		
		Statistic	df	Sig.	Statistic	df	Sig.
S1_0	A	.227	13	.065	.854	13	.032
	B	.180	39	.003	.886	39	.001
	C	.226	35	.000	.740	35	.000
RBD_0	A	.199	13	.167	.903	13	.147
	B	.169	39	.007	.940	39	.037
	C	.119	35	.200 [*]	.935	35	.040
NP_0	A	.139	13	.200 [*]	.921	13	.258
	B	.119	39	.173	.915	39	.006
	C	.174	35	.009	.915	35	.010
S1_30	A	.211	13	.116	.793	13	.006
	B	.131	39	.087	.939	39	.035
	C	.263	35	.000	.698	35	.000
RBD_30	A	.235	13	.048	.805	13	.008
	B	.125	39	.130	.968	39	.320
	C	.134	35	.114	.946	35	.085
N_30	A	.151	13	.200 [*]	.914	13	.207
	B	.111	39	.200 [*]	.933	39	.023
	C	.104	35	.200 [*]	.974	35	.572
S1_90	A	.209	13	.125	.851	13	.030
	B	.100	39	.200 [*]	.969	39	.345
	C	.154	35	.036	.921	35	.016
RBD_90	A	.129	13	.200 [*]	.927	13	.310
	B	.092	39	.200 [*]	.970	39	.364
	C	.127	35	.170	.935	35	.041
N_90	A	.181	13	.200 [*]	.909	13	.175
	B	.089	39	.200 [*]	.957	39	.138
	C	.075	35	.200 [*]	.959	35	.211

Hasil analisis Normalitas Data neutralizing

Tests of Normality							
	Kelompok	Kolmogorov-Smirnov ^a			Shapiro-Wilk		
		Statistic	df	Sig.	Statistic	df	Sig.
Internalization 0 day	B	.254	12	.032	.840	12	.028
	A	.186	12	.200*	.893	12	.127
	C	.170	12	.200*	.906	12	.191
Internalization 30 day	B	.272	12	.014	.813	12	.013
	A	.175	12	.200*	.912	12	.223
	C	.224	12	.099	.802	12	.010
Internalization 90 day	B	.270	12	.016	.818	12	.015
	A	.219	12	.115	.798	12	.009
	C	.233	12	.071	.872	12	.069

*. This is a lower bound of the true significance.

a. Lilliefors Significance Correction

Untuk uji Normalitas data, data dikatakan normal jika hasil statistik menunjukkan nilai $p > 0,05$, dari tes normality menunjukkan bahwa kelompok data yang normal yaitu nilai RBD hari ke-0 pada kelompok A ; NP pada hari ke-0 pada kelompok A; RBD hari ke-30 pada kelompok B dan C; NP pada hari ke-30 pada kelompok C; nilai S1 hari ke-90 pada kelompok C; RBD hari ke-90 pada kelompok A dan B ; NP pada hari ke-90 pada kelompok A,B,C
Sedangkan pada data neutralizing hanya kelompok A dan C hari ke-0 dan kelompok A hari ke-30 yang berdistribusi normal.

Untuk melakukan uji statistik parametrik seperti Uji t Dependent/ Ujit t Independent atau Uji One way Anova, kelompok yang akan diuji semuanya harus terdistribusi normal, jika ada salah satu kelompok saja yang tidak normal, maka sebaiknya menggunakan uji non patametrik. Melihat hasil uji normalitas data yang ada, disarankan untuk melakukan uji non-parametrik