

## DAFTAR PUSTAKA

- Aghamajidi, A., & Maleki Vareki, S. (2022). The Effect of the Gut Microbiota on Systemic and Anti-Tumor Immunity and Response to Systemic Therapy against Cancer. *Cancers (Basel)*, 14(15). <https://doi.org/10.3390/cancers14153563>
- Ahmadi, S., Mainali, R., Nagpal, R., Sheikh-Zeinoddin, M., Soleimanian-Zad, S., Wang, S., Deep, G., Kumar Mishra, S., & Yadav, H. (2017). Dietary Polysaccharides in the Amelioration of Gut Microbiome Dysbiosis and Metabolic Diseases. *Obes Control Ther*, 4(3). <https://doi.org/10.15226/2374-8354/4/2/00140>
- Al-Assal, K., Martinez, A. C., Torrinhas, R. S., Cardinelli, C., & Waitzberg, D. (2018). Gut microbiota and obesity. *Clinical Nutrition Experimental*, 20, 60-64. <https://doi.org/10.1016/j.yclnex.2018.03.001>
- Ali, Y. O., Escala, W., Ruan, K., & Zhai, R. G. (2011). Assaying locomotor, learning, and memory deficits in *Drosophila* models of neurodegeneration. *J Vis Exp*(49). <https://doi.org/10.3791/2504>
- Becattini, S., Taur, Y., & Pamer, E. G. (2016). Antibiotic-Induced Changes in the Intestinal Microbiota and Disease. *Trends Mol Med*, 22(6), 458-478. <https://doi.org/10.1016/j.molmed.2016.04.003>
- Broderick, N. A., Buchon, N., & Lemaitre, B. (2014). Microbiota-induced changes in *Drosophila melanogaster* host gene expression and gut morphology. *mBio*, 5(3), e01117-01114. <https://doi.org/10.1128/mBio.01117-14>
- Buchon, N., Broderick, N. A., Poidevin, M., Pradervand, S., & Lemaitre, B. (2009). *Drosophila* intestinal response to bacterial infection: activation of host defense and stem cell proliferation. *Cell Host Microbe*, 5(2), 200-211. <https://doi.org/10.1016/j.chom.2009.01.003>
- Capo, F., Wilson, A., & Di Cara, F. (2019). The Intestine of *Drosophila melanogaster*: An Emerging Versatile Model System to Study Intestinal Epithelial Homeostasis and Host-Microbial Interactions in Humans. *Microorganisms*, 7(9). <https://doi.org/10.3390/microorganisms7090336>
- Chao, H., Sun, M., Ye, M., Zheng, X., & Hu, F. (2020). World within world: Intestinal bacteria combining physiological parameters to investigate the response of *Metaphire guillelmi* to tetracycline stress. *Environ Pollut*, 261, 114174. <https://doi.org/10.1016/j.envpol.2020.114174>
- Charroux, B., & Royet, J. (2012). Gut-microbiota interactions in non-mammals: what can we learn from *Drosophila*? *Semin Immunol*, 24(1), 17-24. <https://doi.org/10.1016/j.smim.2011.11.003>
- Clark, R. I., & Walker, D. W. (2018). Role of gut microbiota in aging-related health decline: insights from invertebrate models. *Cell Mol Life Sci*, 75(1), 93-101. <https://doi.org/10.1007/s00018-017-2671-1>
- de Man, J. C., Rogosa, M. and Sharpe, M.E. (1960). A Medium for the Cultivation of Lactobacilli. *Journal of Applied Bacteriology*, 23, 130-135. <https://doi.org/https://doi.org/10.1111/j.1365-2672.1960.tb00188.x>
- de Vries, L. E., Valles, Y., Agerso, Y., Vaishampayan, P. A., Garcia-Montaner, A., Kuehl, J. V., Christensen, H., Barlow, M., & Francino, M. P. (2011). The gut as reservoir of

- antibiotic resistance: microbial diversity of tetracycline resistance in mother and infant. *PLoS One*, 6(6), e21644. <https://doi.org/10.1371/journal.pone.0021644>
- di Cerbo, A., Pezzuto, F., Guidetti, G., Canello, S., & Corsi, L. (2019). Tetracyclines: Insights and Updates of their Use in Human and Animal Pathology and their Potential Toxicity. *The Open Biochemistry Journal*, 13(1), 1-12. <https://doi.org/10.2174/1874091x01913010001>
- Dodge, R., Jones, E. W., Zhu, H., Obadia, B., Martinez, D. J., Wang, C., Aranda-Diaz, A., Aumiller, K., Liu, Z., Voltolini, M., Brodie, E. L., Huang, K. C., Carlson, J. M., Sivak, D. A., Spradling, A. C., & Ludington, W. B. (2023). A symbiotic physical niche in *Drosophila melanogaster* regulates stable association of a multi-species gut microbiota. *Nat Commun*, 14(1), 1557. <https://doi.org/10.1038/s41467-023-36942-x>
- Douglas, A. E. (2018). The *Drosophila* model for microbiome research. *Lab Anim (NY)*, 47(6), 157-164. <https://doi.org/10.1038/s41684-018-0065-0>
- Flatt, T. (2020). Life-History Evolution and the Genetics of Fitness Components in *Drosophila melanogaster*. *Genetics*, 214(1), 3-48. <https://doi.org/10.1534/genetics.119.300160>
- Flint, H. J., Scott, K. P., Louis, P., & Duncan, S. H. (2012). The role of the gut microbiota in nutrition and health. *Nat Rev Gastroenterol Hepatol*, 9(10), 577-589. <https://doi.org/10.1038/nrgastro.2012.156>
- Gail A.M. Cresci, K. I. (2019). Chapter 4 - Gut Microbiome. Academic Press, 45-54. <https://doi.org/10.1016/B978-0-12-814330-8.00004-4>
- Gilbert, J. A., Blaser, M. J., Caporaso, J. G., Jansson, J. K., Lynch, S. V., & Knight, R. (2018). Current understanding of the human microbiome. *Nat Med*, 24(4), 392-400. <https://doi.org/10.1038/nm.4517>
- Gökçe Üstündağ, K. B., Ender Büyükgüzel. (2020). Penicillin Impact on Survivorship, Development, and Adult Longevity of *Drosophila melanogaster* (Diptera: Drosophilidae). *Journal of Entomological Science*, 55(4). <https://doi.org/https://doi.org/10.18474/0749-8004-55.4.560>
- Guo, X., Yu, Z., & Yin, D. (2023). Sex-dependent obesogenic effect of tetracycline on *Drosophila melanogaster* deteriorated by dysrhythmia. *J Environ Sci (China)*, 124, 472-480. <https://doi.org/10.1016/j.jes.2021.11.029>
- Hadadi, N., Berweiler, V., Wang, H., & Trajkovski, M. (2021). Intestinal microbiota as a route for micronutrient bioavailability. *Curr Opin Endocr Metab Res*, 20, 100285. <https://doi.org/10.1016/j.coemr.2021.100285>
- Hye-Yeon Lee, S.-H. L., Ji-Hyeon Lee, Won-Jae Lee, Kyung-Jin Min. (2019). The role of commensal microbes in the lifespan of *Drosophila melanogaster*. *Aging*, 11(13). <https://doi.org/10.18632/aging.102073>
- J. Boulétreau-Merle, R. A., Y. Cohet & J. R. David. (1982). Reproductive strategy in *Drosophila melanogaster*: Significance of a genetic divergence between temperate and tropical populations. *Oecologia* 53, 323–329. <https://doi.org/https://doi.org/10.1007/BF00389008>
- James M Kinross , A. W. D. a. J. K. N. (2011). Gut microbiome host interactions in health and disease. *Genome Medicine*. <https://doi.org/https://doi.org/10.1186/gm228>

- Jia, Y., Jin, S., Hu, K., Geng, L., Han, C., Kang, R., Pang, Y., Ling, E., Tan, E. K., Pan, Y., & Liu, W. (2021). Gut microbiome modulates *Drosophila* aggression through octopamine signaling. *Nat Commun*, 12(1), 2698. <https://doi.org/10.1038/s41467-021-23041-y>
- Klein, E. Y., Van Boeckel, T. P., Martinez, E. M., Pant, S., Gandra, S., Levin, S. A., Goossens, H., & Laxminarayan, R. (2018). Global increase and geographic convergence in antibiotic consumption between 2000 and 2015. *Proc Natl Acad Sci U S A*, 115(15), E3463-E3470. <https://doi.org/10.1073/pnas.1717295115>
- Kummerer, K. (2009). Antibiotics in the aquatic environment a review part I. *Chemosphere*, 75(4), 417-434. <https://doi.org/10.1016/j.chemosphere.2008.11.086>
- Kwiatkowska, B., Maslinska, M., Przygodzka, M., Dmowska-Chalaba, J., Dabrowska, J., & Sikorska-Siudek, K. (2013). Immune system as a new therapeutic target for antibiotics. *Advances in Bioscience and Biotechnology*, 04(04), 91-101. <https://doi.org/10.4236/abb.2013.44A013>
- Landis, G. N., Doherty, D., Tower, J. (2020). Analysis of *Drosophila melanogaster* Lifespan. In: Curran, S. (eds) Aging. *Methods in Molecular Biology*, 2144. [https://doi.org/https://doi.org/10.1007/978-1-0716-0592-9\\_4](https://doi.org/https://doi.org/10.1007/978-1-0716-0592-9_4)
- Leftwich, P. T., Clarke, N. V. E., Hutchings, M. I., & Chapman, T. (2017). Gut microbiomes and reproductive isolation in *Drosophila*. *Proc Natl Acad Sci U S A*, 114(48), 12767-12772. <https://doi.org/10.1073/pnas.1708345114>
- Leulier, R. C. M. F. (2014). *Lactobacilli* Host mutualism: learning on the fly. *Microb Cell Fact*. <https://doi.org/https://doi.org/10.1186/1475-2859-13-S1-S6>
- Liu, J., Li, X., & Wang, X. (2019). Toxicological effects of ciprofloxacin exposure to *Drosophila melanogaster*. *Chemosphere*, 237, 124542. <https://doi.org/10.1016/j.chemosphere.2019.124542>
- Lu, X. M., & Lu, P. Z. (2019). Synergistic effects of key parameters on the fate of antibiotic resistance genes during swine manure composting. *Environ Pollut*, 252(Pt B), 1277-1287. <https://doi.org/10.1016/j.envpol.2019.06.073>
- Markow, T. A. (2015a). *Drosophila* reproduction: Molecules meet morphology. *Proc Natl Acad Sci U S A*, 112(27), 8168-8169. <https://doi.org/10.1073/pnas.1510121112>
- Markow, T. A. (2015b). The secret lives of *Drosophila* flies. *eLife*, 4. <https://doi.org/10.7554/eLife.06793>
- Mills, S., Stanton, C., Lane, J. A., Smith, G. J., & Ross, R. P. (2019). Precision Nutrition and the Microbiome, Part I: Current State of the Science. *Nutrients*, 11(4). <https://doi.org/10.3390/nutrients11040923>
- Molinero, N., Ruiz, L., Sanchez, B., Margolles, A., & Delgado, S. (2019). Intestinal Bacteria Interplay With Bile and Cholesterol Metabolism: Implications on Host Physiology. *Front Physiol*, 10, 185. <https://doi.org/10.3389/fphys.2019.00185>
- Nainu, F. (2018). Review : Penggunaan *Drosophila melanogaster* Sebagai Organisme Model Dalam Penemuan Obat. *Jurnal Farmasi Galenika (Galenika Journal of Pharmacy)*. <https://doi.org/10.22487/j24428744>
- Nero, L. A., Beloti, V., DE Aguiar Ferreira Barros, M., Ortolani, M. B. T., Tamanini, R., & DE Melo Franco, B. D. G. (2006). Comparison of petrifilm aerobic count plates

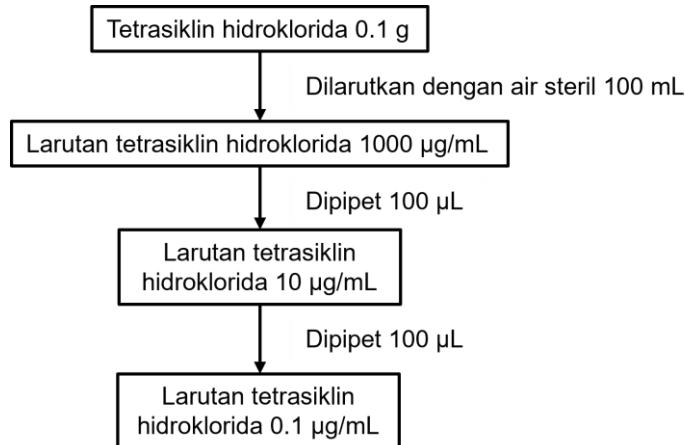
- and de man rogosa sharpe agar for enumeration of lactic acid bacteria. *Journal of Rapid Methods & Automation in Microbiology*. <https://doi.org/https://doi.org/10.1111/j.1745-4581.2006.00050.x>
- Nicholson, J. K., Holmes, E., Kinross, J., Burcelin, R., Gibson, G., Jia, W., & Pettersson, S. (2012). Host-gut microbiota metabolic interactions. *Science*, 336(6086), 1262-1267. <https://doi.org/10.1126/science.1223813>
- Nikolopoulos, N., Matos, R. C., Ravaud, S., Courtin, P., Akherraz, H., Palussiere, S., Gueguen-Chaignon, V., Salomon-Mallet, M., Guillot, A., Guerardel, Y., Chapot-Chartier, M. P., Grangeasse, C., & Leulier, F. (2023). Structure-function analysis of Lactiplantibacillus plantarum DltE reveals D-alanylated lipoteichoic acids as direct cues supporting *Drosophila* juvenile growth. *eLife*, 12. <https://doi.org/10.7554/eLife.84669>
- Pais, I. S., Valente, R. S., Sporniak, M., & Teixeira, L. (2018). *Drosophila melanogaster* establishes a species specific mutualistic interaction with stable gut colonizing bacteria. *PLoS Biol*, 16(7), e2005710. <https://doi.org/10.1371/journal.pbio.2005710>
- Panchal, K., & Tiwari, A. K. (2017). *Drosophila melanogaster* "a potential model organism" for identification of pharmacological properties of plants/plant-derived components. *Biomed Pharmacother*, 89, 1331-1345. <https://doi.org/10.1016/j.biopha.2017.03.001>
- Quigley, E. M. (2013). Gut Bacteria in Health and Disease. *Gastroenterology and Hepatology*, 9(9), 560-569.
- Ramirez, J., Guarner, F., Bustos Fernandez, L., Maruy, A., Sdepanian, V. L., & Cohen, H. (2020). Antibiotics as Major Disruptors of Gut Microbiota. *Front Cell Infect Microbiol*, 10, 572912. <https://doi.org/10.3389/fcimb.2020.572912>
- Rothschild, D. W., O. Barkan, E. Kurilshikov, A. Korem, T. Zeevi, D. Costea, P. I. Godneva, A. Kalka, I. N. Bar, N. Shilo, S. Lador, D. Vila, A. V. Zmora, N. Pevsner-Fischer, M. Israeli, D. Kosower, N. Malka, G. Wolf, B. C. Avnit-Sagi, T. Lotan-Pompan, M. Weinberger, A. Halpern, Z. Carmi, S. Fu, J. Wijmenga, C. Zhernakova, A. Elinav, E. Segal, E. (2018). Environment dominates over host genetics in shaping human gut microbiota. *Nature*, 555(7695), 210-215. <https://doi.org/10.1038/nature25973>
- Rusu, A., & Buta, E. L. (2021). The Development of Third-Generation Tetracycline Antibiotics and New Perspectives. *Pharmaceutics*, 13(12). <https://doi.org/10.3390/pharmaceutics13122085>
- Sapkota, A., Sapkota, A. R., Kucharski, M., Burke, J., McKenzie, S., Walker, P., & Lawrence, R. (2008). Aquaculture practices and potential human health risks: current knowledge and future priorities. *Environ Int*, 34(8), 1215-1226. <https://doi.org/10.1016/j.envint.2008.04.009>
- Schretter, C. E., Vielmetter, J., Bartos, I., Marka, Z., Marka, S., Argade, S., & Mazmanian, S. K. (2018). A gut microbial factor modulates locomotor behaviour in *Drosophila*. *Nature*, 563(7731), 402-406. <https://doi.org/10.1038/s41586-018-0634-9>
- Sekirov, I., Russell, S. L., Antunes, L. C., & Finlay, B. B. (2010). Gut microbiota in health and disease. *Physiol Rev*, 90(3), 859-904. <https://doi.org/10.1152/physrev.00045.2009>

- Selkirk, J., Mohammad, F., Ng, S. H., Chua, J. Y., Tumkaya, T., Ho, J., Chiang, Y. N., Rieger, D., Pettersson, S., Helfrich-Forster, C., Yew, J. Y., & Claridge-Chang, A. (2018). The *Drosophila* microbiome has a limited influence on sleep, activity, and courtship behaviors. *Sci Rep*, 8(1), 10646. <https://doi.org/10.1038/s41598-018-28764-5>
- Siva-Jothy, J. A., Prakash, A., VasanthaKrishnan, R. B., Monteith, K. M., & Vale, P. F. (2018). Oral Bacterial Infection and Shedding in *Drosophila melanogaster*. *J Vis Exp*(135). <https://doi.org/10.3791/57676>
- Sommer, F., & Backhed, F. (2013). The gut microbiota masters of host development and physiology. *Nat Rev Microbiol*, 11(4), 227-238. <https://doi.org/10.1038/nrmicro2974>
- Storelli, G., Defaye, A., Erkosar, B., Hols, P., Royet, J., & Leulier, F. (2011). *Lactobacillus plantarum* promotes *Drosophila* systemic growth by modulating hormonal signals through TOR-dependent nutrient sensing. *Cell Metab*, 14(3), 403-414. <https://doi.org/10.1016/j.cmet.2011.07.012>
- Sun, M., Ye, M., Jiao, W., Feng, Y., Yu, P., Liu, M., Jiao, J., He, X., Liu, K., Zhao, Y., Wu, J., Jiang, X., & Hu, F. (2018). Changes in tetracycline partitioning and bacteria/phage-comediated ARGs in microplastic-contaminated greenhouse soil facilitated by sophorolipid. *J Hazard Mater*, 345, 131-139. <https://doi.org/10.1016/j.jhazmat.2017.11.036>
- Tafesh-Edwards, G., & Eleftherianos, I. (2023). The role of *Drosophila* microbiota in gut homeostasis and immunity. *Gut Microbes*, 15(1), 2208503. <https://doi.org/10.1080/19490976.2023.2208503>
- Tefit, M. A., & Leulier, F. (2017). *Lactobacillus plantarum* favors the early emergence of fit and fertile adult *Drosophila* upon chronic undernutrition. *J Exp Biol*, 220(Pt 5), 900-907. <https://doi.org/10.1242/jeb.151522>
- Trinder, M., Daisley, B. A., Dube, J. S., & Reid, G. (2017). *Drosophila melanogaster* as a High-Throughput Model for Host-Microbiota Interactions. *Front Microbiol*, 8, 751. <https://doi.org/10.3389/fmicb.2017.00751>
- Vajro, P., Paolella, G., & Fasano, A. (2013). Microbiota and gut-liver axis: their influences on obesity and obesity related liver disease. *J Pediatr Gastroenterol Nutr*, 56(5), 461-468. <https://doi.org/10.1097/MPG.0b013e318284abb5>
- Vaz, L. E., Kleinman, K. P., Raebel, M. A., Nordin, J. D., Lakoma, M. D., Dutta-Linn, M. M., & Finkelstein, J. A. (2014). Recent trends in outpatient antibiotic use in children. *Pediatrics*, 133(3), 375-385. <https://doi.org/10.1542/peds.2013-2903>
- Vrieze, A., Out, C., Fuentes, S., Jonker, L., Reuling, I., Koote, R. S., van Nood, E., Holleman, F., Knaapen, M., Romijn, J. A., Soeters, M. R., Blaak, E. E., Dallinga-Thie, G. M., Reijnders, D., Ackermans, M. T., Serlie, M. J., Knop, F. K., Holst, J. J., van der Ley, C., . . . Nieuwdorp, M. (2014). Impact of oral vancomycin on gut microbiota, bile acid metabolism, and insulin sensitivity. *J Hepatol*, 60(4), 824-831. <https://doi.org/10.1016/j.jhep.2013.11.034>
- Vuong, H. E., Yano, J. M., Fung, T. C., & Hsiao, E. Y. (2017). The microbiome and host behavior. *Annu Rev Neurosci*, 40, 21-49. <https://doi.org/10.1146/annurev-neuro-072116-031347>
- Wang, Y., Wu, J., Lv, M., Shao, Z., Hungwe, M., Wang, J., Bai, X., Xie, J., Wang, Y., & Geng, W. (2021). Metabolism Characteristics of Lactic Acid Bacteria and the

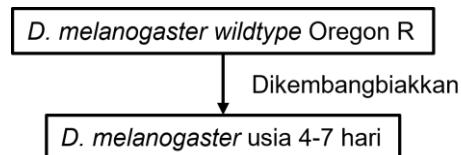
- Expanding Applications in Food Industry. *Front Bioeng Biotechnol*, 9, 612285. <https://doi.org/10.3389/fbioe.2021.612285>
- World, O. H. (2018). WHO report on surveillance of antibiotic consumption: 2016-2018 early implementation. <https://www.who.int/publications/i/item/9789241514880>
- Yu, Z., Shen, J., Li, Z., Yao, J., Li, W., Xue, L., Vandenberg, L. N., & Yin, D. (2020). Obesogenic effect of Sulfamethoxazole on *Drosophila melanogaster* with simultaneous disturbances on eclosion rhythm, glucolipid metabolism, and microbiota. *Environ Sci Technol*, 54(9), 5667-5675. <https://doi.org/10.1021/acs.est.9b07889>
- Zhang, K., Hornef, M. W., & Dupont, A. (2015). The intestinal epithelium as guardian of gut barrier integrity. *Cell Microbiol*, 17(11), 1561-1569. <https://doi.org/10.1111/cmi.12501>
- Zhang, X. P. a. B. (2021). Environmental Toxicology and Toxicogenomics. *Springer Science*, 2326. <https://doi.org/https://doi.org/10.1007/978-1-0716-1514-0>

## LAMPIRAN

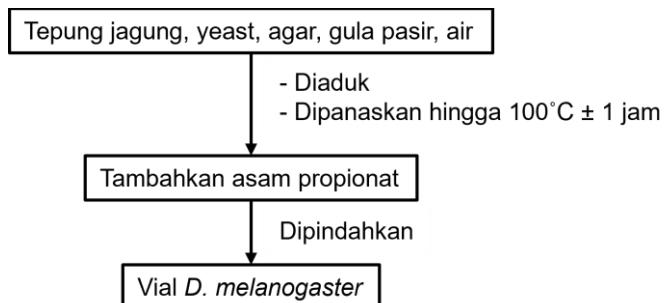
### Lampiran 1. Preparasi sampel



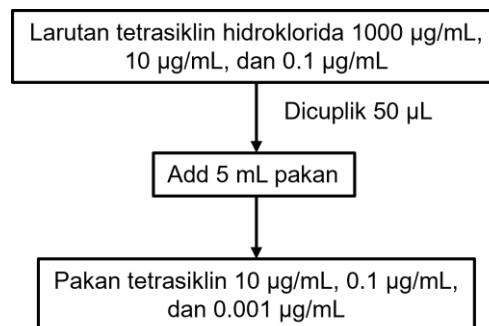
### Lampiran 2. Penyiapan hewan uji



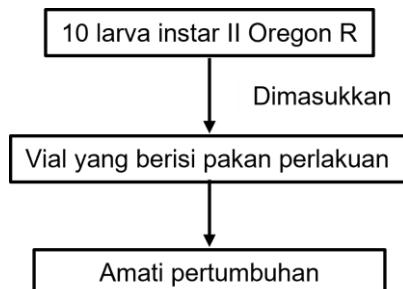
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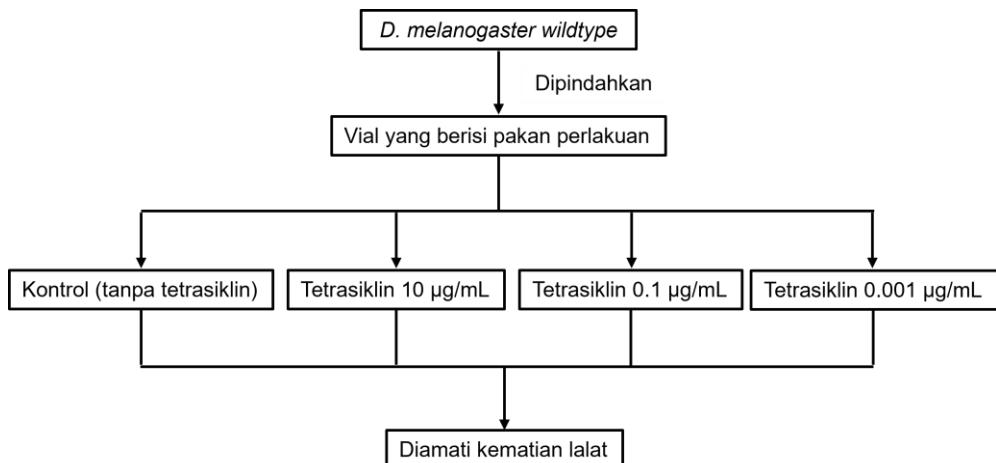
**Lampiran 4.** Penyiapan pakan pengujian



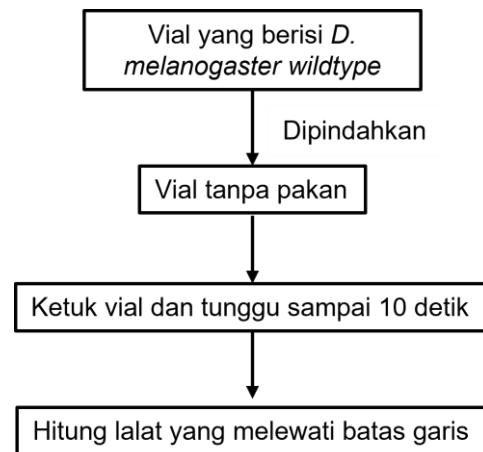
**Lampiran 5.** Uji perkembangan



**Lampiran 6.** Uji survival lalat dewasa



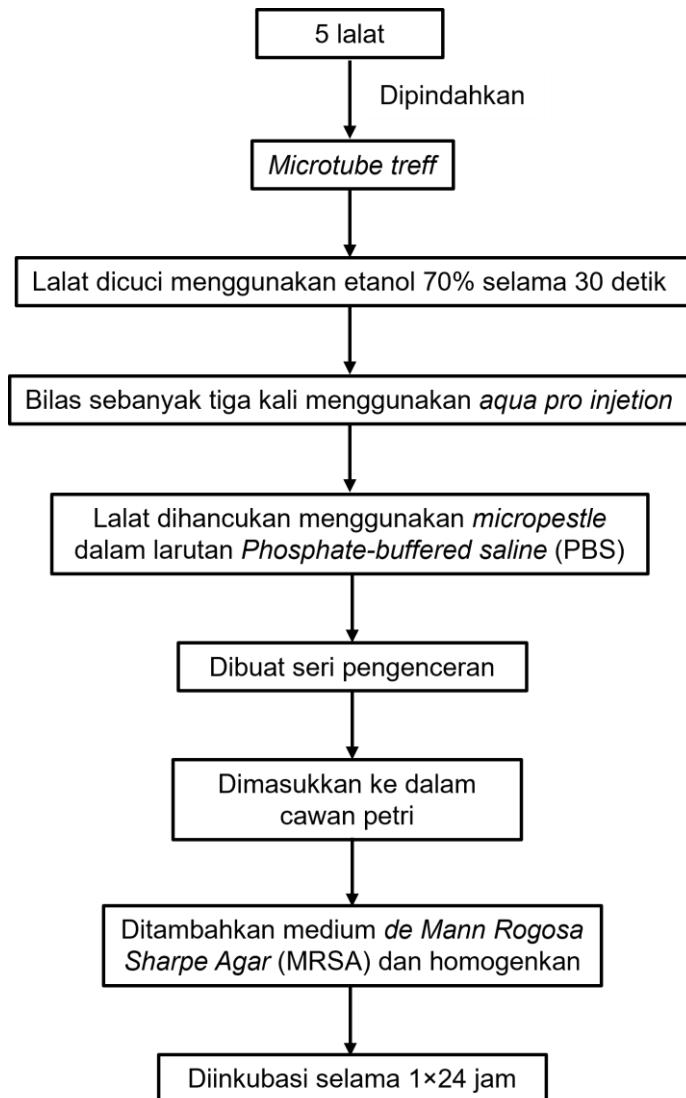
**Lampiran 7. Uji lokomotor lalat dewasa**



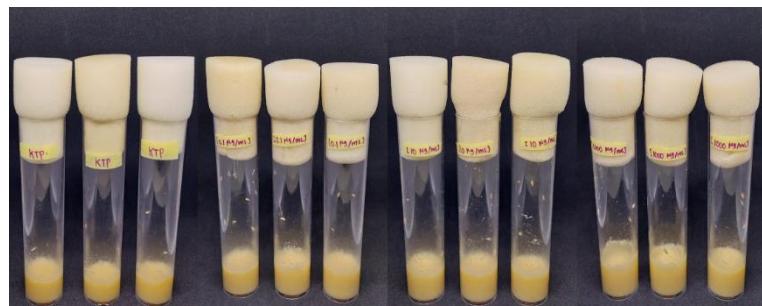
**Lampiran 8. Uji reproduksi**



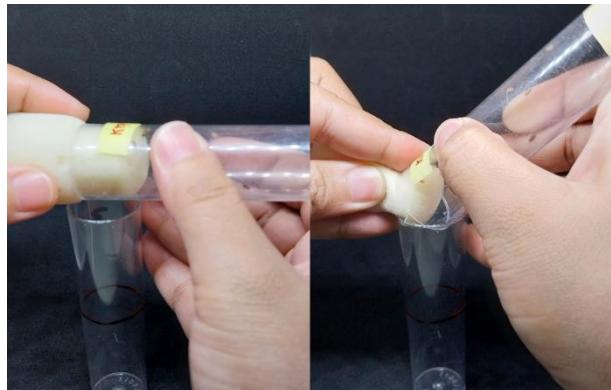
**Lampiran 9.** Uji colony forming unit (CFU)



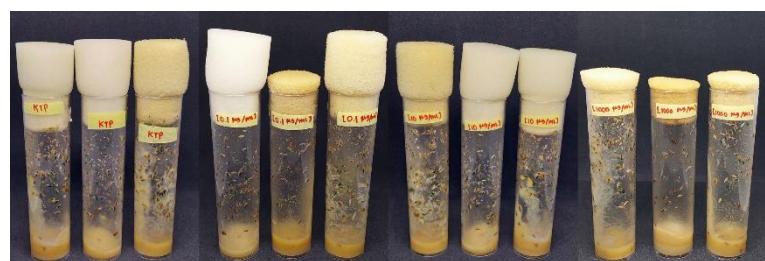
**Lampiran 10.** Gambar penelitian



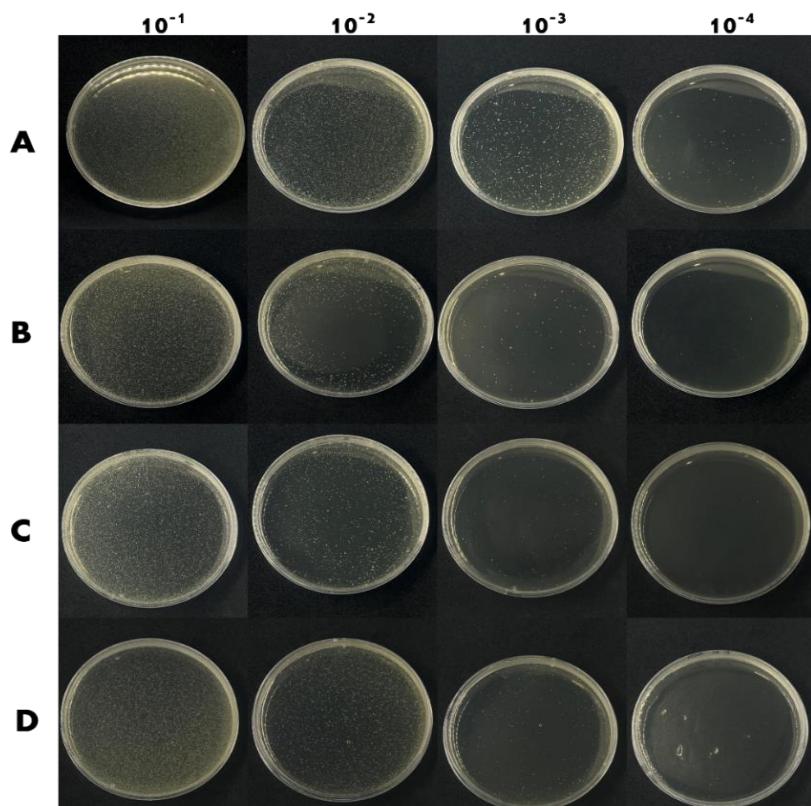
**Gambar 18.** Uji perkembangan larva menjadi pupa dan pupa menjadi lalat yang dilanjutkan ke uji survival lalat dewasa



**Gambar 19.** Uji lokomotor lalat dewasa



**Gambar 20.** Uji reproduksi



**Gambar 21.** Kultur bakteri pada *Drosophila* menggunakan medium *de Mann Rogosa Sharpe Agar* (MRSA). Kontrol tanpa tetrasiiklin (A), tetrasiiklin 0.001 µg/mL (B), tetrasiiklin 0.1 µg/mL (C), tetrasiiklin 10 µg/mL (D).

### Lampiran 11. Perhitungan pengenceran larutan stok

- Dibuat pengenceran untuk konsentrasi 10 µg/mL**

$$V_1 \times N_1 = V_2 \times N_2$$

$$V_1 \times 1000 \text{ } \mu\text{g/mL} = 10 \text{ mL} \times 10 \text{ } \mu\text{g/mL}$$

$$V_1 = \frac{10 \text{ mL} \times 10 \text{ } \mu\text{g/mL}}{1000 \text{ } \mu\text{g/mL}} = 0.1 \text{ mL}$$

- Dibuat pengenceran untuk konsentrasi 0.1 µg/mL**

$$V_1 \times N_1 = V_2 \times N_2$$

$$V_1 \times 10 \text{ } \mu\text{g/mL} = 10 \text{ mL} \times 0.1 \text{ } \mu\text{g/mL}$$

$$V_1 = \frac{10 \text{ mL} \times 0.1 \text{ } \mu\text{g/mL}}{10 \text{ } \mu\text{g/mL}} = 0.1 \text{ mL}$$

### Lampiran 12. Contoh penentuan volume tetrasiklin pada pakan 10 µg/mL

- Konsentrasi 10 µg/mL**

$$V_1 \times N_1 = V_2 \times N_2$$

$$V_1 \times 1000 \text{ } \mu\text{g/mL} = 5 \text{ mL} \times 10 \text{ } \mu\text{g/mL}$$

$$V_1 = \frac{5 \text{ mL} \times 10 \text{ } \mu\text{g/mL}}{1000 \text{ } \mu\text{g/mL}} = 0,05 \text{ mL}$$

### Lampiran 13. Analisis statistik

**Tabel 1.** Hasil perbandingan tukey uji perkembangan larva menjadi pupa setelah terpapar tetrasiklin

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
Kontrol vs. 0.001 µg/mL	0,5	-11,26 to 12,26	NS	0,9993
Kontrol vs. 0.1 µg/mL	0	-11,09 to 11,09	NS	>0,9999
Kontrol vs. 10 µg/mL	2	-9,088 to 13,09	NS	0,953
0.001 µg/mL] vs. 0.1 µg/mL	-0,5	-12,26 to 11,26	NS	0,9993
0.001 µg/mL vs. 10 µg/mL	1,5	-10,26 to 13,26	NS	0,9824
0.1 µg/mL vs. 10 µg/mL	2	-9,088 to 13,09	NS	0,953

**Tabel 2.** Hasil perbandingan tukey uji perkembangan pupa menjadi lalat setelah terpapar tetrasiklin

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
Kontrol vs. 0.001 µg/mL	8	-7,139 to 23,14	NS	0,4537
Kontrol vs. 0.1 µg/mL	2	-13,14 to 17,14	NS	0,981
Kontrol vs. 10 µg/mL	2	-13,14 to 17,14	NS	0,981
0.001 µg/mL vs. 0.1 µg/mL	-6	-21,14 to 9,139	NS	0,6747
0.001 µg/mL vs. 10 µg/mL	-6	-21,14 to 9,139	NS	0,6747
0.1 µg/mL vs. 10 µg/mL	0	-15,14 to 15,14	NS	>0,9999

**Tabel 3.** Hasil perbandingan tukey uji survival lalat dewasa pada hari ke- 35 setelah terpapar tetrasiklin

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
Kontrol vs. 0.001 µg/mL	20	-4,290 to 44,29	NS	0,0975
Kontrol vs. 0.1 µg/mL	20	-4,290 to 44,29	NS	0,0975
Kontrol vs. 10 µg/mL	31,67	9,493 to 53,84	*	0,0122
0.001 µg/mL vs. 0.1 µg/mL	0	-24,29 to 24,29	NS	>0,9999
0.001 µg/mL vs. 10 µg/mL	11,67	-10,51 to 33,84	NS	0,3198
0.1 µg/mL vs. 10 µg/mL	11,67	-10,51 to 33,84	NS	0,3198

**Tabel 4.** Hasil perbandingan tukey uji lokomotor lalat dewasa antara kelompok kontrol dan 0.001 µg/mL

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
1:Kontrol vs. 1:0.001 µg/mL	2	-21,46 to 25,46	NS	>0,9999
3:Kontrol vs. 3:0.001 µg/mL	5,667	-17,79 to 29,13	NS	>0,9999
6:Kontrol vs. 6:0.001 µg/mL	-13	-36,46 to 10,46	NS	0,8382
9:Kontrol vs. 9:0.001 µg/mL	-13	-36,46 to 10,46	NS	0,8382
12:Kontrol vs. 12:0.001 µg/mL	-6,667	-30,13 to 16,79	NS	0,9999
15:Kontrol vs. 15:0.001 µg/mL	-2,667	-26,13 to 20,79	NS	>0,9999
18:Kontrol vs. 18:0.001 µg/mL	14	-9,458 to 37,46	NS	0,7479
21:Kontrol vs. 21:0.001 µg/mL	11,33	-12,13 to 34,79	NS	0,9414
24:Kontrol vs. 24:0.001 µg/mL	0,3333	-23,79 to 23,13	NS	>0,9999
27:Kontrol vs. 27:0.001 µg/mL	5,333	-18,13 to 28,79	NS	>0,9999

**Tabel 5.** Hasil perbandingan tukey uji lokomotor lalat dewasa antara kelompok kontrol dan 0.1 µg/mL

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
1:Kontrol vs. 1:0.1 µg/mL	3,667	-31,18 to 38,52	NS	>0,9999
3:Kontrol vs. 3:0.1 µg/mL	6,333	-28,52 to 41,18	NS	>0,9999
6:Kontrol vs. 6:0.1 µg/mL	-17,33	-52,18 to 17,52	NS	0,9259
9:Kontrol vs. 9:0.1 µg/mL	-0,3333	-35,18 to 34,52	NS	>0,9999
12:Kontrol vs. 12:0.1 µg/mL	-5	-39,85 to 29,85	NS	>0,9999
15:Kontrol vs. 15:0.1 µg/mL	2,333	-32,52 to 37,18	NS	>0,9999
18:Kontrol vs. 18:0.1 µg/mL	16,67	-18,18 to 51,52	NS	0,9462
21:Kontrol vs. 21:0.1 µg/mL	11	-23,85 to 45,85	NS	0,9994
24:Kontrol vs. 24:0.1 µg/mL	-14,33	-49,18 to 20,52	NS	0,987
27:Kontrol vs. 27:0.1 µg/mL	38,33	3,482 to 73,18	*	0,0186

**Tabel 6.** Hasil perbandingan tukey uji lokomotor lalat dewasa antara kelompok kontrol dan 10 µg/mL

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
1:Kontrol vs. 1:10 µg/mL	2,333	-23,77 to 28,44	NS	>0,9999
3:Kontrol vs. 3:10 µg/mL	1	-25,10 to 27,10	NS	>0,9999
6:Kontrol vs. 6:10 µg/mL	-2,333	-28,44 to 23,77	NS	>0,9999
9:Kontrol vs. 9:10 µg/mL	-0,3333	-26,44 to 25,77	NS	>0,9999
12:Kontrol vs. 12:10 µg/mL	11	-15,10 to 37,10	NS	0,9834
15:Kontrol vs. 15:10 µg/mL	4,333	-21,77 to 30,44	NS	>0,9999
18:Kontrol vs. 18:10 µg/mL	25,33	-0,7698 to 51,44	NS	0,0659
21:Kontrol vs. 21:10 µg/mL	20,67	-5,437 to 46,77	NS	0,2803
24:Kontrol vs. 24:10 µg/mL	12	-14,10 to 38,10	NS	0,9619
27:Kontrol vs. 27:10 µg/mL	33,33	7,230 to 59,44	**	0,0027

**Tabel 7.** Hasil perbandingan tukey uji reproduksi jumlah pupa setelah terpapar tetrasiklin selama 5 hari

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
Kontrol vs. 0,001 µg/mL	-1,633	-19,07 to 15,80	NS	0,9899
Kontrol vs. 0,1 µg/mL	1,133	-16,30 to 18,57	NS	0,9965
Kontrol vs. 10 µg/mL	1,433	-16,00 to 18,87	NS	0,9931
0,001 µg/mL vs. 0,1 µg/mL	2,767	-14,67 to 20,20	NS	0,9548
0,001 µg/mL vs. 10µg/mL	3,067	-14,37 to 20,50	NS	0,9402
0,1 µg/mL vs. 10 µg/mL	0,3	-17,13 to 17,73	NS	>0,9999

**Tabel 8.** Hasil perbandingan tukey uji reproduksi jumlah lalat setelah terpapar tetrasiklin selama 5 hari

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
Kontrol vs. 0,001 µg/mL	-2,633	-17,26 to 12,00	NS	0,9364
Kontrol vs. 0,1 µg/mL	-0,7667	-15,40 to 13,86	NS	0,9982
Kontrol vs. 10 µg/mL	1,933	-12,70 to 16,56	NS	0,9729
0,001 µg/mL vs. 0,1 µg/mL	1,867	-12,76 to 16,50	NS	0,9754
0,001 µg/mL vs. 10 µg/mL	4,567	-10,06 to 19,20	NS	0,754
0,1 µg/mL vs. 10 µg/mL	2,7	-11,93 to 17,33	NS	0,932

**Tabel 9.** Hasil perbandingan tukey jumlah koloni bakteri lalat dewasa setelah terpapar tetrasiklin selama 35 hari

Tukey's multiple comparisons test	Mean Diff	95,00% CI of diff	Summary	Adjusted P Value
Kontrol vs. 0.001 µg/mL	1623000	1209774 to 2036226	****	<0,0001
Kontrol vs. 0.1 µg/mL	1555000	1141774 to 1968226	****	<0,0001
Kontrol vs. 10 µg/mL	1410000	996774 to 1823226	****	<0,0001
0.001 µg/mL vs. 0.1 µg/mL	-68000	-481226 to 345226	NS	0,9501
0.001 µg/mL vs. 10 µg/mL	-213000	-626226 to 200226	NS	0,4057
0.1 µg/mL vs. 10 µg/mL	-145000	-558226 to 268226	NS	0,6863

**Lampiran 14.** Perhitungan jumlah koloni bakteri asam laktat**Tabel 10.** Hasil perhitungan jumlah koloni lalat dewasa pada hari ke-35 hari

Kelompok	Pengenceran				ALT Rerata
	10 <sup>-1</sup>	10 <sup>-2</sup>	10 <sup>-3</sup>	10 <sup>-4</sup>	
Kontrol	TBUD	TBUD	TBUD	151	1.5x10 <sup>6</sup>
	TBUD	TBUD	TBUD	211	2.1x10 <sup>6</sup>
	TBUD	TBUD	TBUD	167	16 x 10 <sup>6</sup>
0.001 µg/mL	TBUD	TBUD	122	12	1.2 x 10 <sup>5</sup>
	TBUD	TBUD	108	10	1.1 x 10 <sup>5</sup>
	TBUD	TBUD	191	15	1.9 x 10 <sup>5</sup>
0.1 µg/mL	TBUD	TBUD	230	14	2.3 x 10 <sup>5</sup>
	TBUD	TBUD	201	15	2.0 x 10 <sup>5</sup>
	TBUD	TBUD	194	12	1.9 x 10 <sup>5</sup>
10 µg/mL	TBUD	TBUD	TBUD	33	3.3 x 10 <sup>5</sup>
	TBUD	TBUD	TBUD	39	3.9 x 10 <sup>5</sup>
	TBUD	TBUD	TBUD	34	3.4 x 10 <sup>5</sup>

Ket: Tidak bisa untuk dihitung (TBUD)