

DAFTAR PUSTAKA

- Abbas, S., Shanbhag, T. and Kothare, A., 2021. Applications of bromelain from pineapple waste towards acne. *Saudi J. Biol. Sci.* 28(1), 1001–1009. doi.org/10.1016/j.sjbs.2020.11.032.
- Abebe, B., Park, J. H., Pyo, S., Gibtan, A., Park, S., Choi, J., Kim, M., 2020. Activities of eastern eel (*Anguilla japonica*). *Korean J. Food Preserv.* 27(10), 959–972.
- Abuine, R., Rathnayake, A. U. and Byun, H. G., 2019. Biological activity of peptidas purified from fish skin hydrolysates. *Fish Aquatic Sci.* 22(1), 1–14. doi.org/10.1186/s41240-019-0125-4.
- Affandi, R., 2005. Strategi pemanfaatan sumberdaya ikan sidat, *Anguilla* spp. di Indonesia. *J. Iktiologi Indones.* 5(2), 77–81.
- Akar, Z., Küçük, M., and Doğan, H., 2017. A new colorimetric DPPH• scavenging activity method with no need for a spectrophotometer applied on synthetic and natural antioxidants and medicinal herbs. *J. Enzyme Inhib. Med. Chem.* 32(1), 640–647. doi.org/10.1080/14756366.2017.1284068.
- Akbarian, M., Khani, A., Eghbalpour, S., and Uversky, V. N., 2022. Bioactive peptidas: synthesis, sources, applications, and proposed mechanisms of action. *Int. J. Mol. Sci.* 23(3), 1445. doi: 10.3390/ijms23031445.
- Al-Sa'ady, A., Al-Hadban, W., and Al-Zubaidy, M., 2016. Optimal conditions for bromelain extraction from pineapple fruit (*Ananas comosus*). *J. Eng. Sci. Technol.* 34(5), 675–682.
- Alam, M. N., Bristi, N. J., and Rafiquzzaman, M., 2013. Review on in vivo and in vitro methods evaluation of antioxidant activity. *Saudi Pharm. J.* 21(2), 143–152. doi.org/10.1016/j.jps.2012.05.002.
- Antoine, F. R., Wei, C. I., Littell, R. C., and Marshall, M. R., 1999. HPLC method for analysis of free amino acids in fish using o-phthaldialdehyde precolumn derivatization. *J. Agric. Food Chem.* 47(12), 5100–5107. doi.org/10.1021/jf990032+.
- Antolovich, M., Prenzler, P. D., Patsalides, E., McDonald, S., and Robards, K., 2002. Methods for testing antioxidant activity. *Analyst*, 127(1), 183–198. doi.org/10.1039/b009171p.
- Aoyama, J., 2009. Life history and evolution of migration in catadromous eels (Genus *Anguilla*). *Aqua-BioScience Monographs.* 2(1), 1–42. doi.org/10.5047/absm.2009.00201.0001.
- Apostolopoulos, V., Bojarska, J., Chai, T. T., Elnagdy, S., Kaczmarek, K., Matsoukas, J., New, R., Parang, K., Lopez, O. P., Parhiz, H., Perera, C. O., Pickholz, M., Remko, M., Saviano, M., Skwarczynski, M., Tang, Y., Wolf, W. M., Yoshiya, T., Zabrocki, J., Toth, I., 2021. A global review on short peptidas: frontiers and perspectives. *Molecules*. 26(2). doi.org/10.3390/molecules26020430.

- Arai, T., and Abdul Kadir, S. R., 2017. Diversity, distribution, and different habitat use among the tropical freshwater eels of the genus *Anguilla*. *Sci. Rep.* 7(1), 1–12. doi.org/10.1038/s41598-017-07837-x.
- Asmat, U., Abad, K., and Ismail, K., 2016. Diabetes mellitus and oxidative stress—a concise review. *Saudi Pharm J.* 24(5), 547–553. doi.org/10.1016/j.jsps.2015.03.013.
- Association of Official Analytical Chemist (AOAC), 2005. Official methods of analysis (18 Edn). Washington (US): The Association of Official Analytical Chemist Inc.
- Auwal, S. M., Zarei, M., Abdul-Hamid, A., and Saari, N., 2017. Response surface optimization for the production of antioxidant hydrolysates from stone fish protein using bromelain. *Evid.-Based Complementary Altern. Med.* 2017. doi.org/10.1155/2017/4765463.
- Azad, S., 2018. Amino acids: Its types and uses. *Int. j. clin. diagn. pathol.*, 1(1), 13–16.
- Bartsch, P., Berg, R., Gabriel, O., Henderson, I. W., Kamstra, A., Kloppmann, M., Reimer, L. W., Wirth, T., White, R. J., and Thorpe, J. E., 1877. The eel. *Nature*. 15 (380). doi.org/10.2307/1443633.
- Bashir, K. M. I., Park, Y. J., An, J. H., Choi, S. J., Kim, J. H., Baek, M. K., Kim, A., Sohn, J. H., and Choi, J. S., 2018. Antioxidant properties of *Scomber japonicus* hydrolysates prepared by enzymatic hydrolysis. *J. Aquat. Food Prod. Technol.* 27(1), 107–121. doi.org/10.1080/10498850.2017.1407013.
- Baskoro, B. D., Nugraha, R. A., Puspitawati, R., and Redjeki, S., 2017. Effect of centrifugation at 7.000 g, 8.000 g, and 9.000 g on the salivary protein profile ≥ 30 kDa. *J. Phys. Conf. Ser.* 884(1). doi.org/10.1088/1742-6596/884/1/012013.
- Blois, M., 1958. Antioxidant determination by the use of a stable free radical. *Nature*. 181, 1198–1200. doi.org/10.1038/1811199a0.
- Burgos-Morón, E., Abad-Jiménez, Z., de Marañón, A. M., Iannantuoni, F., Escribano-López, I., López-Domènech, S., Salom, C., Jover, A., Mora, V., Roldan, I., Solá, E., Rocha, M., and Víctor, V. M., 2019. Relationship between oxidative stress, ER stress, and inflammation in type 2 diabetes: The battle continues. *J. Clin. Med.* 8(9). doi.org/10.3390/jcm8091385.
- Cai, L., Wu, X., Zhang, Y., Li, X., Ma, S., and Li, J., 2015. Purification and characterization of three antioxidant peptidas from protein hydrolysate of grass carp (*Ctenopharyngodon idella*) skin. *J. Funct. Foods.* 16, 234–242. doi.org/10.1016/j.jff.2015.04.042.
- Chai, I. J., and Arai, T., 2018. Ages at maturation of tropical freshwater eels, *Anguilla bicolor bicolor* and *Anguilla bengalensis bengalensis*. *J. Appl. Anim. Res.* 46(1), 1108–1113. doi.org/10.1080/09712119.2018.1470090.
- Chakrabarti, S., Guha, S., and Majumder, K., 2018. Food-derived bioactive peptidas in human health: Challenges and opportunities. *Nutrients*. 10(11), 1–17. doi.org/10.3390/nu10111738.

- Cheng, I. C., Liao, J. X., Ciou, J. Y., Huang, L. T., Chen, Y. W., and Hou, C. Y., 2020. Characterization of protein hydrolysates from eel (*Anguilla marmorata*) and their application in herbal eel extracts. *Catalysts.* 10 (2). doi.org/10.3390/catal10020205.
- Chi, C. F., Wang, B., Hu, F. Y., Wang, Y. M., Zhang, B., Deng, S. G., and Wu, C. W., 2015. Purification and identification of three novel antioxidant peptidas from protein hydrolysate of bluefin leatherjacket (*Navodon septentrionalis*) skin. *Food Res. Int.* 73, 124–129. doi.org/10.1016/j.foodres.2014.08.038.
- Cicero AFG, Fogacci F, Colletti A., 2017. Potential role of bioactive peptidas in prevention and treatment of chronic diseases: a narrative review. *Br.J Pharmacol.*174(11), 1378–94. doi:10.1111/bph.13608.
- Corzo, C. A., Waliszewski, K. N., and Welti-Chanes, J., 2012. Pineapple fruit bromelain affinity to different protein substrates. *Food Chem.* 133 (3), 631–635. doi.org/10.1016/j.foodchem.2011.05.119.
- Craig, S., 2017. Understanding fish nutrition, feeds, and feeding. Virginia Cooperative Extension. diunduh dari: <https://digitalpubs.ext.vt.edu/vcedigitalpubs/8651631966449426/MobilePageReplica.action?pm=2&folio=1#pg1>.
- Daliri, E. B. M., Oh, D. H., and Lee, B. H., 2017. Bioactive peptidas. *Foods.* 6(5), 1–21. doi.org/10.3390/foods6050032.
- Damodaran, S., and Parkin, K. L., 2017. Fennema's food chemistry: Amino acids, peptidas, and proteins. CRC Press, Wales. doi.org/10.1201/9781315372914.
- Daniel, R. M., Danson, M. J., Eisenthal, R., Lee, C. K., and Peterson, M. E., 2008. The effect of temperature on enzyme activity: New insights and their implications. *Extremophiles,* 12(1), 51–59. doi.org/10.1007/s00792-007-0089-7.
- Das, A., Nayak, Y., and Dash, S., 2021. Fish protein hydrolysate production, treatment methods, and current potential uses: A review. *Int. J. Fish. Aquat.* 9(2), 195–200.
- Dash, P., and Ghosh, G., 2017. Amino acid composition, antioxidant and functional properties of protein hydrolysates from cucurbitaceae seeds. *J. Food Sci. Technol.* 54(13), 4162–4172. doi.org/10.1007/s13197-017-2855-6.
- De Torre, M. P., Cavero, R. Y., Calvo, M. I., and Vizmanos, J. L. W., 2019. A simple and a reliable method to quantify antioxidant activity in vivo. *Antioxidants,* 8(5), 1–11. doi.org/10.3390/antiox8050142.
- Demirhan, E., Apar, D. K., and Özbek, B., 2011. Sesame cake protein hydrolysis by alcalase: Effects of process parameters on hydrolysis, solubilization, and enzyme inactivation. *Korean J. Chem. Eng.* 28(1), 195–202. doi.org/10.1007/s11814-010-0316-2.
- Dhillon, A., Sharma, K., Rajulapati, V., and Goyal, A., 2016. Proteolytic enzymes. In current developments in biotechnology and bioengineering: production, isolation and purification of industrial products. Elsevier Inc, Amsterdam. doi.org/10.1016/B978-0-444-63662-1.00007-5.

- Di Meo, S., and Venditti, P., 2020. Evolution of the knowledge of free radicals and other oxidants. *Oxid. Med. Cell Longev.* 2020. doi.org/10.1155/2020/9829176.
- Englard, S., and Seifter, S., 1990. Guide to protein purification: Precipitation techniques. Academic Press, New York.
- Esfandi, R., Walters, M. E., and Tsopmo, A. 2019. Antioxidant properties and potential mechanisms of hydrolyzed proteins and peptidas from cereals. *Heliyon.* 5(4), 1538. doi.org/10.1016/j.heliyon.2019.e01538.
- Fahmi, M. R., 2015. Konservasi genetik ikan sidat tropis (*Anguilla* spp.) di perairan Indonesia. *JPPI.* 21(1), 45–54.
- Ferri, M., Gianotti, A., and Tassoni, A., 2013. Optimization of assay conditions for the determination of antioxidant capacity and polyphenols in cereal food components. *J. Food Compos. Anal.* 30(2), 94–101. doi.org/10.1016/j.jfca.2013.02.004.
- Froese, R., and Pauly, D., 2021. Fish Identification : Anguillidae Freshwater eels. doi.org/10.1038/2261175c0
- Gajanan, P. G., Elavarasan, K., and Shamasundar, B. A., 2016. Bioactive and functional properties of protein hydrolysates from fish frame processing waste using plant proteases. *Environ. Sci. Pollut. Res.* 23(24), 24901–24911. doi.org/10.1007/s11356-016-7618-9.
- George, S., and Abrahamse, H., 2020. Redox potential of antioxidants in cancer progression and prevention. *Antioxidant.* 9(11), 1–21. doi.org/10.3390/antiox9111156.
- Gong, X., An, Q., Le, L., Geng, F., Jiang, L., Yan, J., Xiang, D., Peng, L., Zou, L., Zhao, G., and Wan, Y., 2020. Prospects of cereal protein-derived bioactive peptidas: Sources, bioactivities diversity, and production. *Crit. Rev. Food Sci. Nutr.* 0 (0), 2855–2871. doi.org/10.1080/10408398.2020.1860897.
- Griñan-Lison, C., Blaya-Cánovas, J. L., López-Tejada, A., Ávalos-Moreno, M., Navarro-Ocón, A., Cara, F. E., González-González, A., Lorente, J. A., Marchal, J. A., and Granados-Principal, S., 2021. Antioxidants for the treatment of breast cancer: Are we there yet?. *Antioxidants.* 10(2), 1–44. doi.org/10.3390/antiox10020205.
- Grzonka, Z., Kasprzykowski, F., and Wiczk, W., 2007. Cysteine proteas: Julio Polaina (ed). Industrial enzymes. Springer. doi: 10.1007/1-4020-5377-0_11.
- Gupta, R. K., Patel, A. K., Shah, N., Chaudhary, A. K., Jha, U. K., Yadav, U. C., Gupta, P. K., and Pakuwal, U., 2014. Oxidative stress and antioxidants in disease and cancer: A review. *Asian Pac. J. Cancer Prev.* 15(11), 4405–4409. doi.org/10.7314/APJCP.2014.15.11.4405.
- Haider, K., Haider, M. R., Neha, K., and Yar, M. S., 2020. Free radical scavengers: An overview on heterocyclic advances and medicinal prospects. *Eur. J. Med. Chem.* 204. doi.org/10.1016/j.ejmech.2020.112607.
- Halliwell, B., 1995. How to characterize an antioxidant: an update. *Biochem Soc. Symposium.* 61, 73–101. doi.org/10.1042/bss0610073.

- Haslaniza, H., Maskat, M. Y., Wan Aida, W. M., and Mamot, S., 2010. The effects of enzyme concentration, temperature and incubation time on nitrogen content and degree of hydrolysis of protein precipitate from cockle (*Anadara granosa*) meat wash water. Int. Food Res. J. 17(1), 147–152.
- Hidayati, A., 2019. aktivitas antioksidan hidrolisat protein miofibril belut (*Synbranchus bengalensis*) yang dihidrolisis dengan enzim papain. Tesis M.Si. IPB, Bogor. Tidak dipublikasikan.
- Hou, Y., Wu, Z., Dai, Z., Wang, G., and Wu, G., 2017. Protein hydrolysates in animal nutrition: Industrial production, bioactive peptidas, and functional significance. J. Anim. Sci. Biotechnol. 8(1), 1–13. doi.org/10.1186/s40104-017-0153-9.
- Hu, X. M., Wang, Y. M., Zhao, Y. Q., Chi, C. F., and Wang, B., 2020. Antioxidant peptidas from the protein hydrolysate of monkfish (*Lophius litulon*) Muscle: Purification, identification, and cytoprotective function on HepG2 cells damage by H₂O₂. Mar. Drugs, 18(3). doi.org/10.3390/md18030153.
- Huang, D., Boxin, O. U., and Prior, R. L., 2005. The chemistry behind antioxidant capacity assays. J. Agric. Food Chem. 53(6), 1841–1856. doi.org/10.1021/jf030723c.
- Huyen, K. T., and Linh, N. Q., 2020. Nutritional composition and lipid content of skin and muscle of wild giant mottled eel *Anguilla marmorata* in Thua Thien Hue, Vietnam. 129(3), 5–14. doi.org/10.26459/hueunijard.v129i3C.5848.
- Is, K., and Tor, I., 2007. Comparative evaluation of Fe (III) reducing power-based antioxidant capacity assays in the presence of phenanthroline, batho - phenanthroline , tripyridyltriazine (FRAP), and ferricyanide reagents. Talanta. 72, 1157–1165. doi.org/10.1016/j.talanta.2007.01.019.
- Islam, S., Hongxin, W., Admassu, H., Noman, A., Ma, C., and An, F., 2021. Degree of hydrolysis, functional and antioxidant properties of protein hydrolysates from Grass Turtle (*Chinemys reevesii*) as influenced by enzymatic hydrolysis conditions. Food Sci. Nutr. 2020, 1–17. doi.org/10.1002/fsn3.1903.
- Jakubczyk, A., Karas, M., Rybczynska-Tkaczyk, K., Zielinska, E., and Zielinski, D., 2020. Current trends of bioactive peptidas - new sources and therapeutic effect. Foods. 9(7). doi.org/10.3390/foods9070846.
- Jamaluddin, Rusli, C., Yuyun, Y., and Widodo, A., 2019. Comparative study of fish eel amino acid profile (*Anguilla marmorata* (Q.) Gaimard) on silver eel phase from Palu river and Poso lake. J. Pharm. Nutr. Sci. 9(2), 125–129. doi.org/10.29169/1927-5951.2019.09.02.12.
- Karakoltsidis, P. A., and Constantinides, S. M., 1995. The eels, *Anguilla* spp., their characteristics and uses. Food Rev. Int. 11(2), 347–361. doi.org/10.1080/87559129509541045.
- Karso, K., Wuryanti, W., and Sriatun, S., 2014. Isolasi dan karakterisasi kitinase isolat jamur akuatik kitinolitik KC3 dari kecoa (*Orthoptera*). JKSA. 17(2), 51–57. https://doi.org/10.14710/jksa.17.2.51-57.
- Kementerian Kelautan dan Perikanan (KKP), 2018. Sidat. 2018. Diakses 16 Juli 2021. https://kkp.go.id/djprl/bpsplpadang/page/323-sidat.

- Kementerian Kelautan dan Perikanan (KKP)., 2020. KKP Ajak masyarakat kembangkan potensi budidaya sidat. Diakses 16 Juli 2021. <https://kkp.go.id/artikel/21418-kkp-ajak-masyarakat-kembangkan-potensi-budidaya-sidat>.
- Krzemi, J., Wronka, M., Mlynarska, E., Franczyk, B., and Jacek Rysz, 2022. Arterial hypertension — oxidative stress and inflammation. *Antioxidant*. 2022 (11), 72. doi.org/ 10.3390/antiox11010172.
- Kusharto, C. M., Emi Widayarsi, R. A. H., Budywiryan, B., Sri Wiyono, E., and Sugengherisuseno, S., 2014. Nutritive value and fatty acids profile of fresh indonesian eel (*Anguilla bicolor*) and kabayaki. *Malaysian J. Med. Health Sci.* 12(1), 41–46. doi.org/10.17576/jskm-1201-2014-06.
- Kusumaningtyas, E., Widiastuti, R., Kusumaningrum, H. D., and Suhartono, T., 2015. protein susu kambing dengan ekstrak kasar bromelin (antibacterial and antioxidant activity of hydrolysate from goat milk protein hydrolized by crude bromelain extract). *J. Teknol. dan Industri Pangan*. 26(2), 179–188. doi.org/10.6066/jtip.2015.26.2.179.
- Leite, J. A. S., Montoya, C. A., Loveday, S. M., Maes, E., Mullaney, J. A., McNabb, W. C., and Roy, N. C., 2021. Heat-treatments affect protease activities and peptida profiles of ruminants' milk. *Front. Nutr.* 8(3), 1–10. doi.org/10.3389/fnut.2021.626475.
- Lin, J., Hong, H., Zhang, L., Zhang, C., and Luo, Y., 2019. Antioxidant and cryoprotective effects of hydrolysate from gill protein of bighead carp (*Hypophthalmichthys nobilis*) in preventing denaturation of frozen surimi. *Food Chem.* 298(5), 124868. doi.org/10.1016/j.foodchem.2019.05.142.
- Liu, J., Zhang, D., Zhu, Y., Wang, Y., He, S., and Zhang, T., 2018. Enhancing the in vitro antioxidant capacities via the interaction of amino acids. *Emirates J. Food Agr.* 30(3), 224–231. doi.org/10.9755/ejfa.2018.v30.i3.1641.
- Lobo, V., Patil, A., Phatak, A., and Chandra, N., 2010. Free radicals, antioxidants and functional foods: Impact on human health. *Pharm. Rev.* 4(8), 118–126. doi.org/10.4103/0973-7847.70902
- Martins, B. C., Rescolino, R., Coelho, D. F., Zanchetta, B., Tambourgi, E. B., and Silveira, E., 2014. Characterization of bromelain from *Ananas comosus* agroindustrial residues purified by ethanol fractional precipitation. *Chem. Eng. Trans.* 37, 781–786. doi.org/10.3303/CET143713.
- Masoodi, K. Z., Lone, S. M., and Rasool, R. S., 2021. Study of principle of centrifugation. advanced methods in molecular biology and biotechnology. Elsevier Inc. 133–137. doi.org/10.1016/b978-0-12-824449-4.00023-2.
- Miller, M. J., Westerberg, H., Sparholt, H., Wysujack, K., Sørensen, S. R., Marohn, L., Jacobsen, M. W., Freese, M., Ayala, D. J., Pohlmann, J. D., Svendsen, J. C., Watanabe, S., Andersen, L., Møller, P. R., Tsukamoto, K., Munk, P., and Hanel, R., 2019. Spawning by the European eel across 2000 km of the Sargasso Sea. *Biol. Lett.* 15(4). doi.org/10.1098/rsbl.2018.0835.
- Mohan, R., Sivakumar, V., Rangasamy, T., and Muralidharan, C., 2016. Optimisation of bromelain enzyme extraction from pineapple (*Ananas comosus*) and application in process industry. *Am. J. Biochem. Biotechnol.* 12(3), 188–195. doi.org/10.3844/ajbbsp.2016.188.195.

- Mora, L., Aristoy, M. C., and Toldrá, F., 2018. Bioactive peptidas in encyclopedia of food chemistry, 381–389. doi.org/10.1016/B978-0-08-100596-5.22397-4.
- Munteanu, I. G., and Apetrei, C., 2021. Analytical methods used in determining antioxidant activity: A review. Int. J. Mol. Sci. 22(7). doi.org/10.3390/ijms22073380.
- Munthe, I., Isa, M., Winaruddin, W., Sulasmi, S., Herrialfian, H., and Rusli, R., 2016. Analisis kadar protein ikan depik (*Rasbora tawarensis*) di danau laut tawar kabupaten Aceh Tengah. J. Med. Vet. 10(1), 67. doi.org/10.21157/j.med.vet..v10i1.4044.
- Nafsiyah, I., Nurilmala, M., and Abdullah, A., 2018. Komposisi nutrisi ikan sidat *Anguilla bicolor bicolor* dan *Anguilla marmorata*. JPHPI. 21(3), 504–512.
- Noman, A., Xu, Y., AL-Bukhaiti, W. Q., Abed, S. M., Ali, A. H., Ramadhan, A. H., and Xia, W., 2018. Influence of enzymatic hydrolysis conditions on the degree of hydrolysis and functional properties of protein hydrolysate obtained from Chinese sturgeon (*Acipenser sinensis*) by using papain enzyme. Process Biochem. 679, 19–28. doi.org/10.1016/j.procbio.2018.01.009.
- Novaes, L. C. de L., Jozala, A. F., Lopes, A. M., Santos-Ebinuma, V. de C., Mazzola, P. G., and Junior, A. P., 2015. Stability, purification, and applications of bromelain. Bioseparations and Downstream Processing. 50, 1–34. doi.org/10.1002/btpr.
- Pavan, R., Jain, S., Shraddha, and Kumar, A., 2012. Properties and therapeutic application of bromelain: A review. Biotech. Res. Int. 2012, 1–6. doi.org/10.1155/2012/976203.
- Pavia, D. L., Lampman, G. M., and Kriz, G. S., 2001. Introduction to spectroscopy : A guide for students of organic chemistry. Thomson Learning Inc. doi.org/10.1201/9781439894651-21
- Peterson, M. E., Daniel, R. M., Danson, M. J., and Eisenthal, R., 2007. The dependence of enzyme activity on temperature: Determination and validation of parameters. Biochem. J. 402(2), 331–337. doi.org/10.1042/BJ20061143.
- Pisoschi, A. M., and Pop, A., 2015. The role of antioxidants in the chemistry of oxidative stress: A review. European J. Med. Chem. 97, 55–74. doi.org/10.1016/j.ejmech.2015.04.040.
- Pizzino, G., Irrera, N., Cucinotta, M., Pallio, G., Mannino, F., Arcoraci, V., Squadrito, F., Altavilla, D., and Bitto, A., 2017. Oxidative Stress: harms and benefits for human health. Oxid. Med. Cell. Longev. 2017. doi.org/10.1155/2017/8416763.
- Poh, S. S., and Abdul Majid, F. A., 2011. Thermal stability of free bromelain and bromelain-polyphenol complex in pineapple juice. Int Food Res. J. 18(3), 1051–1060.
- Purwanto, M. G. M., 2014. Perbandingan analisis kadar protein terlarut dengan berbagai metode spektroskopi UV-Visible. JIST. 7, 1–71.
- Raghavan S, Kristinsson HG, and Leeuwenburgh C., 2008. Radical scavenging and reducing ability of tilapia (*Oreochromis niloticus*) protein hydrolysates. J. Agric. Food Chem. 56 (21), 10359–67. doi.org/10.1021/jf8017194.

- Rivas-Vela, C. I., Amaya-Llano, S. L., Castaño-Tostado, E., and Castillo-Herrera, G. A., 2021. Protein hydrolysis by subcritical water: A new perspective on obtaining bioactive peptidas. *Molecules*, 26(21), 1–15. doi.org/10.3390/molecules26216655.
- Robinson, P. K., 2015. Enzymes: principles and biotechnological applications. *Essays in Biochem.* 59, 1–41. doi.org/10.1042/BSE0590001.
- Rowan, A. D., and Buttle, D. J., 1994. Pineapple cysteine endopeptidases: Wurtzek (ed). *Methods in enzymology*. Academic Press, Amsterdam. 555–568. doi.org/10.1016/0076-6879(94)44040-9.
- Sadili, D., Haryono, and Kamal, M. M. 2015. Pedoman umum restoking jenis ikan terancam punah. Kementerian Kelautan dan Perikanan.
- Sánchez, A., and Vázque, A., 2017. Bioactive peptidas: A review. *Food Qual. Saf.* 1(4), 29–46. doi.org/10.1093/fqs/fyx006.
- Shahi, Z., Sayyed-alangi, S. Z., and Naja, L., 2020. Effects of enzyme type and process time on hydrolysis degree , electrophoresis bands and antioxidant properties of hydrolyzed proteins derived from defatted *Bunium persicum* Bioss. *Heliyon*. 6(2). doi.org/10.1016/j.heliyon.2020.e03365.
- Shahidi, F., and Zhong, Y., 2015. Measurement of antioxidant activity. *J. Funct. Foods*. 18, 757–781. doi.org/10.1016/j.jff.2015.01.047.
- Sharifi-Rad, M., Anil Kumar, N. V., Zucca, P., Varoni, E. M., Dini, L., Panzarini, E., Rajkovic, J., Tsouh Fokou, P. V., Azzini, E., Peluso, I., Prakash Mishra, A., Nigam, M., El Rayess, Y., Beyrouty, M. El, Polito, L., Iriti, M., Martins, N., Martorell, M., Docea, A. O., Sharifi-Rad, J., 2020. Lifestyle, oxidative stress, and antioxidants: back and forth in the pathophysiology of chronic diseases. *Front. Physiol.* 11(7), 1–21. doi.org/10.3389/fphys.2020.00694.
- Simamora, A., 2015. Asam amino, peptida, dan protein. Buku ajar blok 3 biologi sel (1). http://repository.ukrida.ac.id/bitstream/123456789/580/4/Asam%20amino%20protein_2015%20Maret%202016.pdf.
- Singh, P. K., Shrivastava, N., and Ojha, B. K., 2019. Enzymes in the meat industry: Mohammad Kuddus (Ed). *Enzymes in food biotechnology: production, applications, and future prospects*. Elsevier Inc, Amsterdam. 111–128. doi.org/10.1016/B978-0-12-813280-7.00008-6.
- Singh, R., Devi, S., and Gollen, R., 2015. Role of free radical in atherosclerosis, diabetes and dyslipidaemia: Larger-than-life. *Diabetes/Metab. Res. Rev.* 31(2), 113–126. doi.org/10.1002/dmrr.2558.
- Skoog, D. A., West, D. M., Holler, F. J., and Crouch, S. R., 2014. *Fundamentals of analytical chemistry Ninth Edition*. Mary Finch, Boston.
- Soares, P. A. G., Vaz, A. F. M., Correia, M. T. S., Pessoa, A., and Carneiro-Da-Cunha, M. G., 2012. Purification of bromelain from pineapple wastes by ethanol precipitation. *Sep. Purif. Technol.*, 98, 389–395. doi.org/10.1016/j.seppur.2012.06.042.
- Sonklin, C., Laohakunjit, N, and Kerdchoechuen, O, 2018. Assessment of antioxidant properties of membrane ultrafiltration peptidas from mungbean meal protein hydrolysates. *PeerJ*. 6, 5337; DOI 10.7717/peerj.5337.

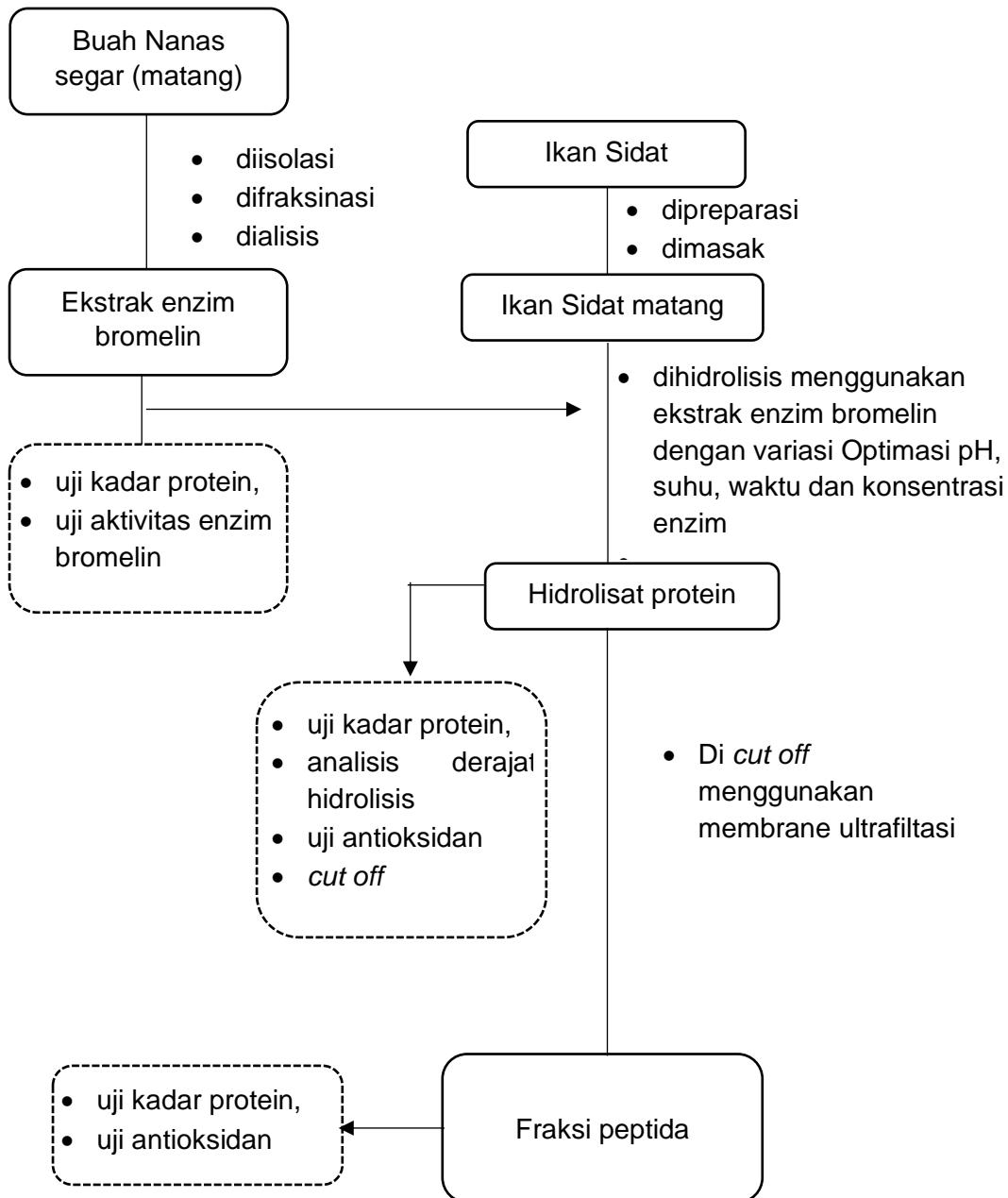
- Stephenson, F. H., 2010. Calculations for Molecular Biology and Biotechnology. Academic Press, Amsterdam. 413–421. doi.org/10.1016/B978-0-12-375690-9.00012-7.
- Subagio, A., Windrati, W. S., Fauzi, M., and Witono, Y., 2004. Karakterisasi protein miofibril dari ikan kuniran dan ikan mata besar. Jurnal Teknol. dan Industri Pangan. 15 (1), 70–78.
- Suprobo, C. O., Suprihati, S., and Wuryanti, W. 2011. Uji antikanker isolat bioaktif L-asparaginase dari kunyit putih (*Curcuma mangga* Val.) terhadap sel kanker serviks. JKSA. 14(2), 58–63. doi.org/10.14710/jksa.14.2.58-63.
- Suryati, K. 2018. Species composition and length-weight relationship of anguillid eels habited in Bengkulu waters, Indonesia. IJEMS. 2(2018), 48–53. doi.org/11.26554/ijems.2218.2.2.48-53.
- Tadesse, S. A., and Emire, S. A. 2020. Production and processing of antioxidant bioactive peptidas: A driving force for the functional food market. Heliyon. 6(8), 04765. doi.org/10.1016/j.heliyon.2020.e04765.
- Takahashi, Y., Kamata, A., and Konishi, T., 2021. Dipeptidyl peptidase - IV inhibitory peptidas derived from salmon milt and their effects on postprandial blood glucose level. Fish. Sci. https://doi.org/10.1007/s12562-021-01530-9.
- Tan, B. L., Norhaizan, M. E., Liew, W. P. P., and Rahman, H. S., 2018. Antioxidant and oxidative stress: A mutual interplay in age-related diseases. Front. Pharmacol. 9(10), 1–28. doi.org/10.3389/fphar.2018.01162.
- Tapal, A., and Tiku, P. K., 2018. Nutritional and nutraceutical improvement by enzymatic modification of food proteins. Mohammad Kuddus (Ed). Enzymes in food biotechnology: production, applications, and future prospects. Elsevier Inc, Amsterdam. doi.org/10.1016/B978-0-12-813280-7.00027-X.
- Tazkiah, N. P., Rosahdi, T. D., dan Supriadin, A., 2019 . Isolasi dan karakterisasi enzim amilase dari biji nangka (*Artocarpus heterophyllus*). Al-Kimiya, 4(1), 17–22. https://doi.org/10.15575/ak.v4i1.5079.
- Tkaczewska, J., Bukowski, M., and Mak, P., 2019. Identification of antioxidant peptidas in enzymatic hydrolysates of carp (*Cyprinus carpio*) skin gelatin. Molecules. 24(1), 1–15. doi.org/10.3390/molecules24010097.
- Touyz, R. M., Rios, F. J., Alves-Lopes, R., Neves, K. B., Camargo, L. L., and Montezano, A. C., 2020. Oxidative stress: A unifying paradigm in hypertension. CJC. 36(5), 659–670. doi.org/10.1016/j.cjca.2020.02.081.
- Wali, N., 2018. Non-vitamin and Nonmineral Nutritional Supplements Volume 1: Pineapple (*Ananas comosus*). Elsevier Inc, Amsterdam. doi.org/10.1016/B978-0-12-812491-8.00050-3.
- Wang, J., Wang, Y. M., Li, L. Y., Chi, C. F., and Wang, B., 2022. Twelve antioxidant peptidas from protein hydrolysate of skipjack tuna (*Katsuwonus pelamis*) roe prepared by flavourzyme: purification, sequence identification, and activity evaluation. Front. Nutr. 8(1), 1–14. doi.org/10.3389/fnut.2021.813780.
- Wang, X., Yu, H., Xing, R., and Li, P., 2017. Characterization, preparation, and purification of marine bioactive peptidas. BioMed Res. Int. 2017. doi.org/10.1155/2017/9746720.

- Wijayanti, I., and Setiyorini, E. S. S., 2018. Nutritional Content of wild and cultured eel (*Anguilla bicolor*) from Southern Coast of Central Java. Ilmu Kelautan: Indonesian Journal of Marine Sciences. 23(1), 37. doi.org/10.14710/ik.ijms.23.1.37-44.
- Wijeratnam, S. W., 2016. Pineapple: Benjamin Caballero (Ed) . Encyclopedia of Food and Health. Academic Press, Amsterdam. 380–384. doi.org/10.1016/B978-0-12-384947-2.00547-X.
- World Register of Marine Species (WORMS)., 2021. *Anguilla marmorata* Quoy and Gaimard, 1824. diakses 23 Mei 2021. <http://www.marinespecies.org/aphia.php?p=taxdetails&andid=217457>.
- Worsfold, P., 2005. Encyclopedia of Analytical Science: Spectrophotometry. Elsevier Ltd, Amsterdam. 318–321.
- Yadav, A., Kumari, R., Yadav, A., Mishra, J. P., Seweta, S., and Prabha, S., 2016. Antioxidants and its functions in human body. Res. Environ. Life Sci. 9(11), 1328–1331.
- Jay Kant Yadav & V. Prakash (2011) Stabilization of α -amylase, the key enzyme in carbohydrates properties alterations, at low pH. Int. J. Food Prop. 14 (6), 1182-1196. doi.org/10.1080/10942911003592795.
- Yi, T., Li, S., Fan, J-Y., Fan, L., Zhang, F.F., Luo, P., Zhang, X. J., Wang, J.G., Zhu, L., Zhao, Z.Z., and Chen, H. B., 2014. Comparative analysis of EPA and DHA in fish oil nutritional capsules by GC-MS. Lipids Health Dis. 13, 190. doi:10.1186/1476-511X-13-190.
- Yi-Cheng, H., Yu-San, H., Hsiang-Yi, H., and Yen-Ting, L., 2020. Skin coloration and habitat preference of the freshwater *Anguilla* eels. Int. J. Aqua. Fishery Sci. 6, 096–101. doi.org/10.17352/2455-8400.000063.
- Young, I. ., and Woodside, J., 2001. Antioxidants in health and disease. J Clin. Pathol. 54, 176–186. doi.org/10.1201/b18539.
- Zamora-Sillero, J., Gharsallaoui, A., and Prentice, C., 2018. Peptidas from fish by-product protein hydrolysates and its functional properties: an overview. Marine Biotech. 20(2), 118–130. doi.org/10.1007/s10126-018-9799-3.
- Zhang, Y., Dong, Y., and Dai, Z., 2021. Antioxidant and cryoprotective effects of bone hydrolysates from bighead carp (*Aristichthys nobilis*) in freeze-thawed fish fillets. Foods. 10(6). doi.org/10.3390/foods10061409.
- Zhao, G. X., Yang, X. R., Wang, Y. M., Zhao, Y. Q., Chi, C. F., and Wang, B., 2019 . Antioxidant peptidas from the protein hydrolysate of spanish mackerel (*Scomberomorus niphonius*) muscle by in vitro gastrointestinal digestion and their in vitro activities. Mar. Drugs. 17(9), 1–17. doi.org/10.3390/md17090531.
- Zhong, Y., and Shahidi, F., 2015. Handbook of Antioxidants for Food Preservation: Methods for the assessment of antioxidant activity in foods. Elsevier Ltd, Amsterdam. doi.org/10.1016/B978-1-78242-089-7.00012-9 .
- Zou, T. Bin, He, T. P., Li, H. Bin, Tang, H. W., and Xia, E. Q., 2016. The structure-activity relationship of the antioxidant peptidas from natural proteins. Molecules. 21(1), 1–14. doi.org/10.3390/molecules21010072.

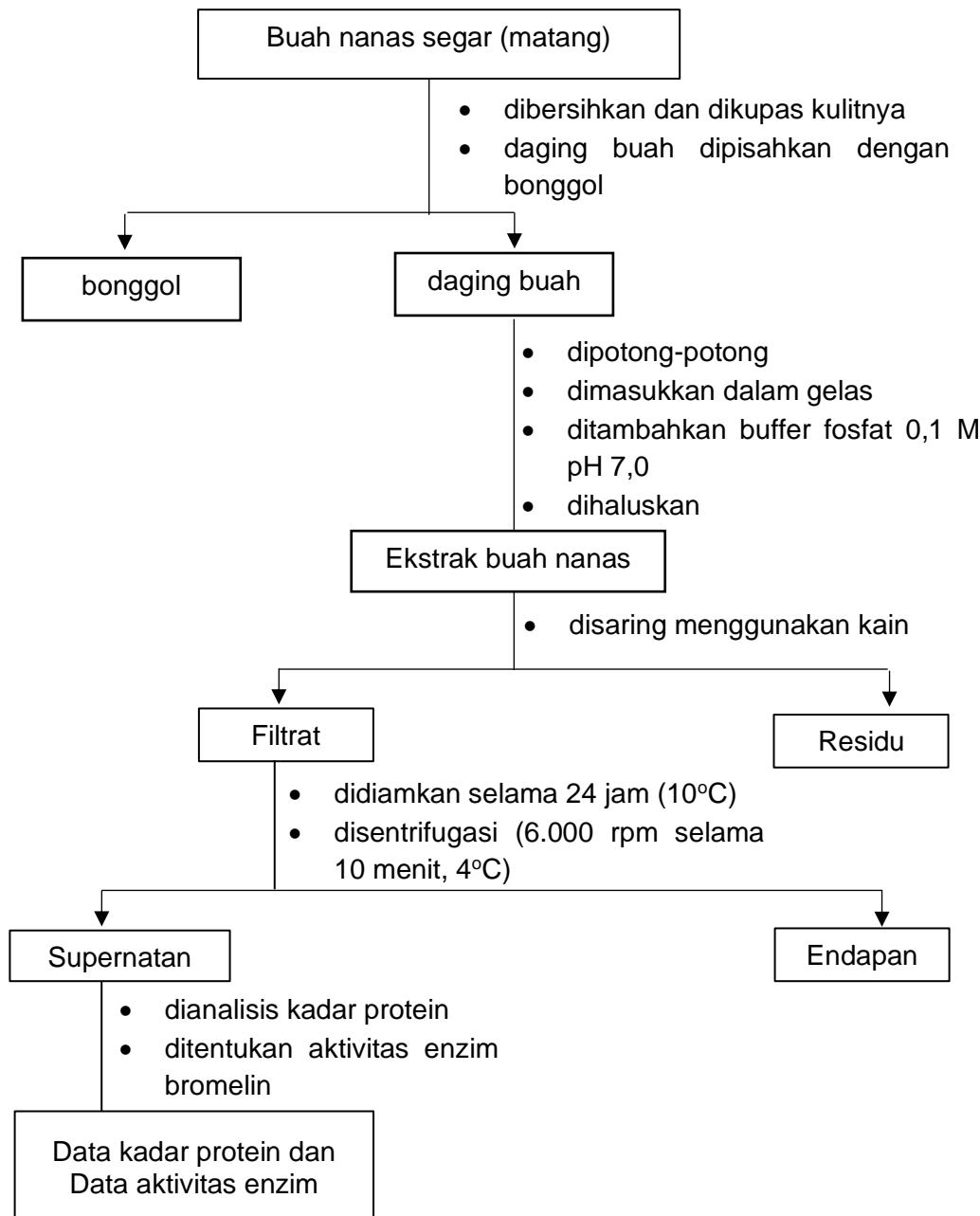
Zhou, W., Ye, C., Geng, L., Chen, G., Wang, X., Chen, W., Sa, R., Zhang, J., and Zhang, X., 2021. Purification and characterization of bromelain from pineapple (*Ananas comosus* L.) peel waste. *J. Food Sci.* 86(2), 385–393. doi.org/10.1111/1750-3841.15563.

LAMPIRAN

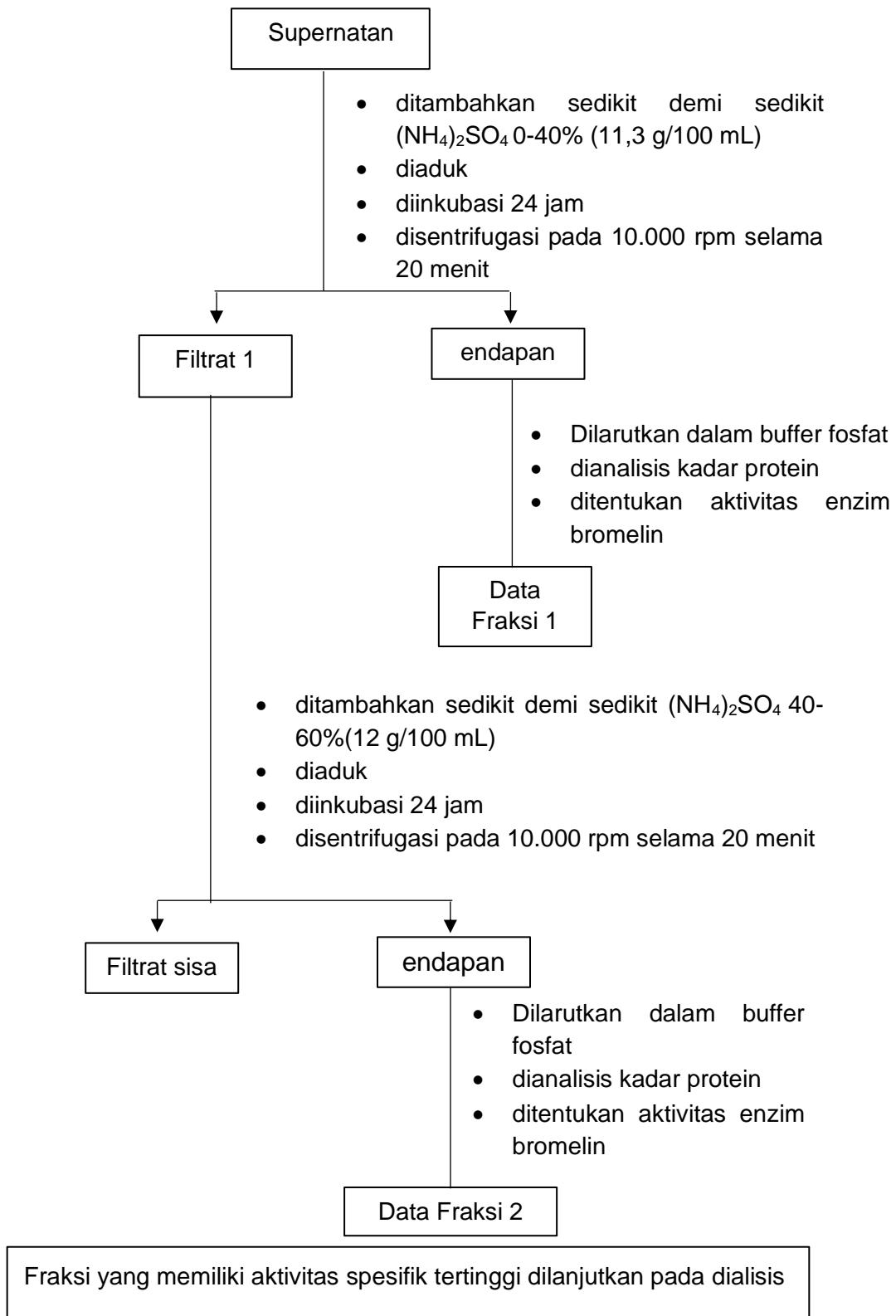
Lampiran 1: Tahapan Penelitian



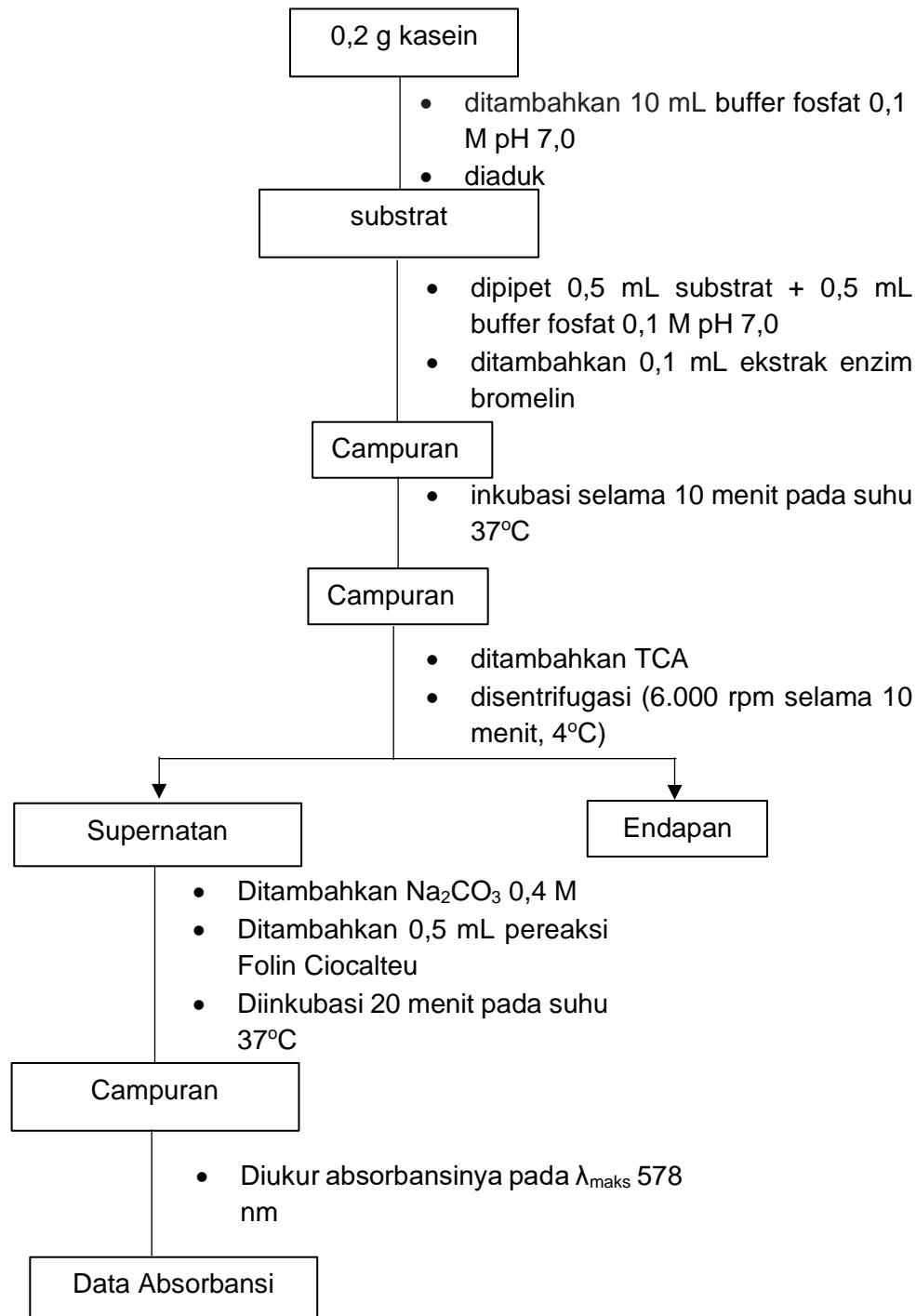
Lampiran 2: Skema Kerja Preparasi dan Isolasi Ekstrak Enzim Bromelin



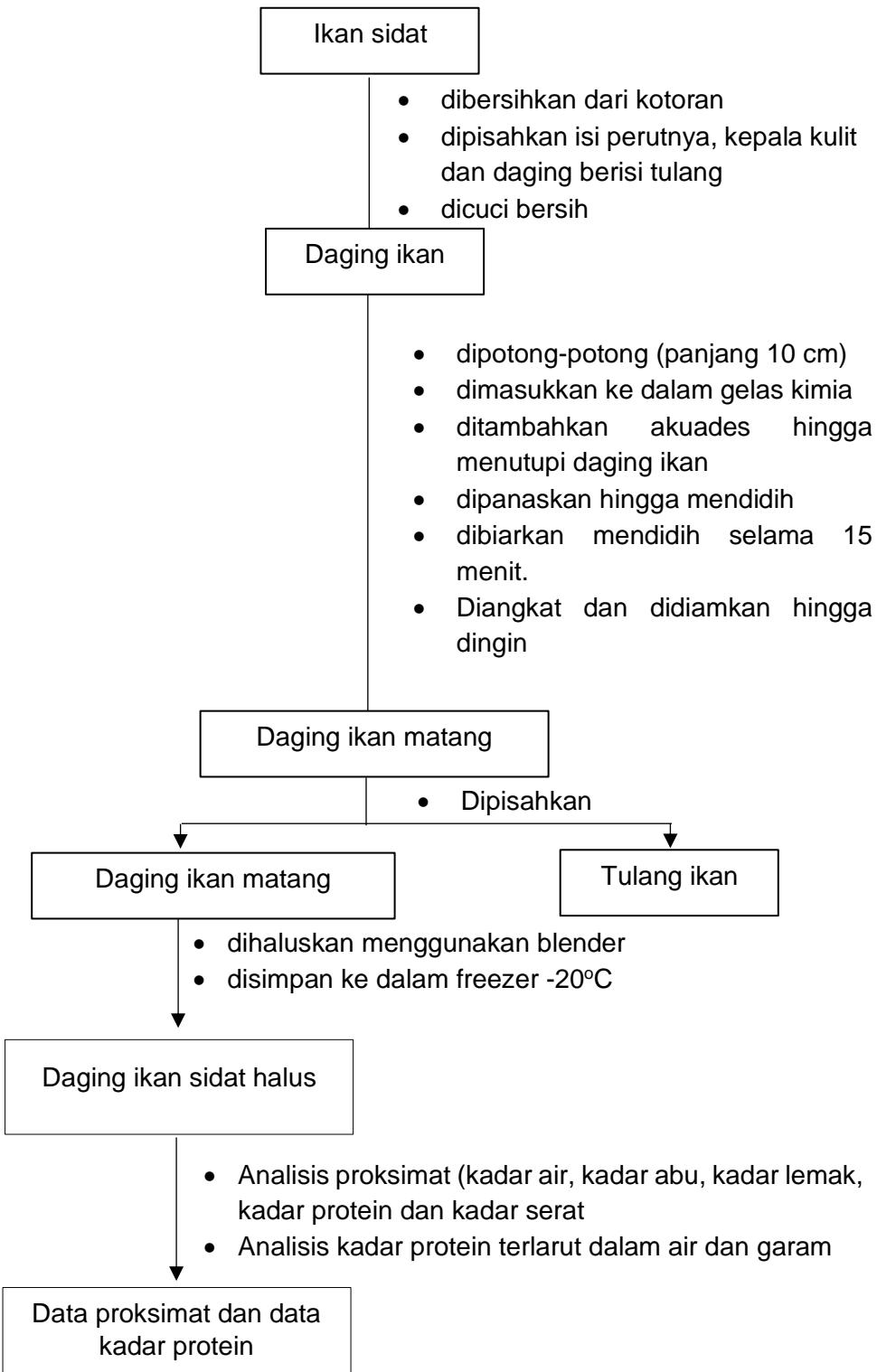
Lampiran 3: Skema Kerja Pemurnian Parsial Ekstrak Enzim Bromelin



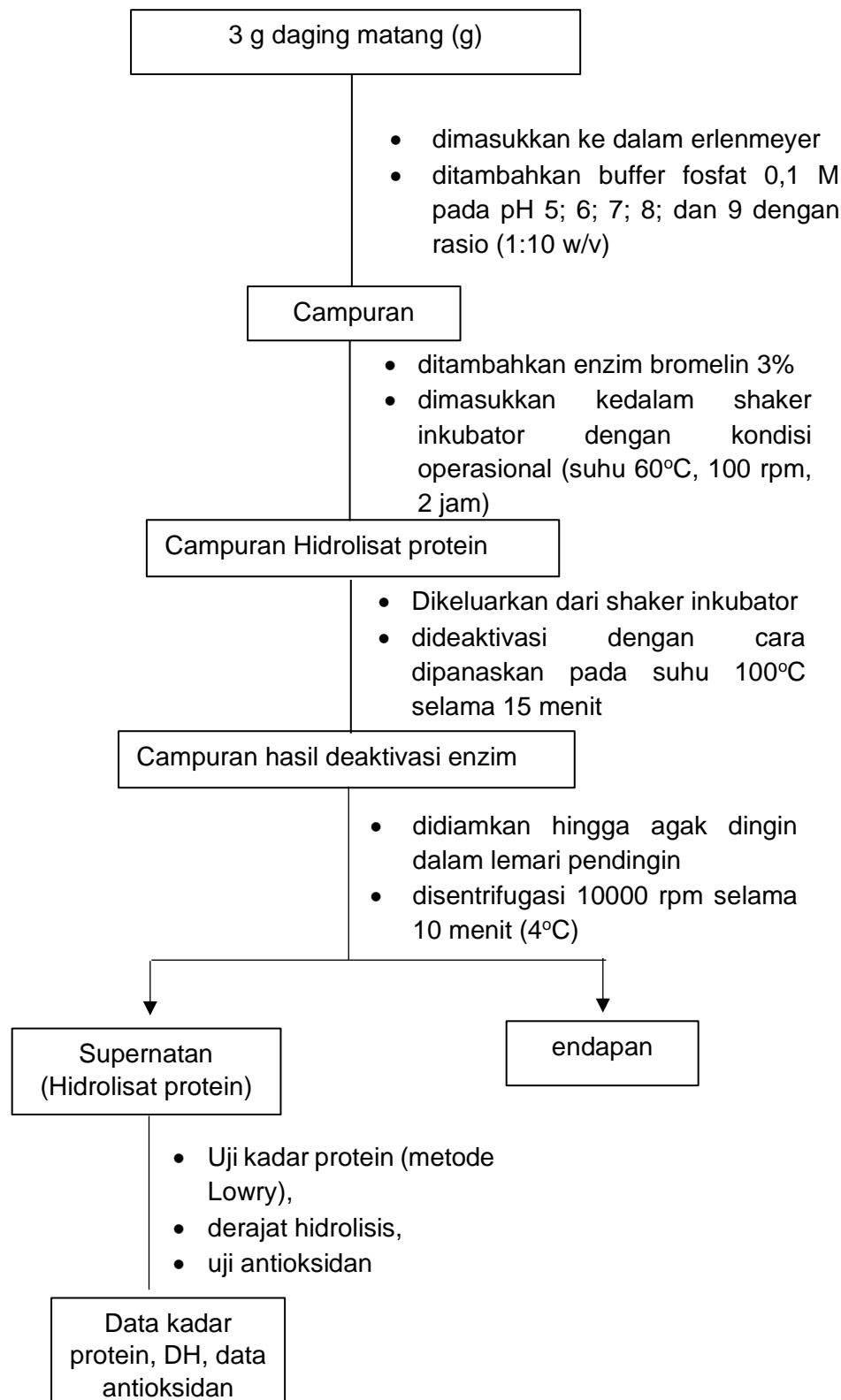
Lampiran 4: Skema Kerja Penentuan Aktivitas Bromelin Terhadap Kasein



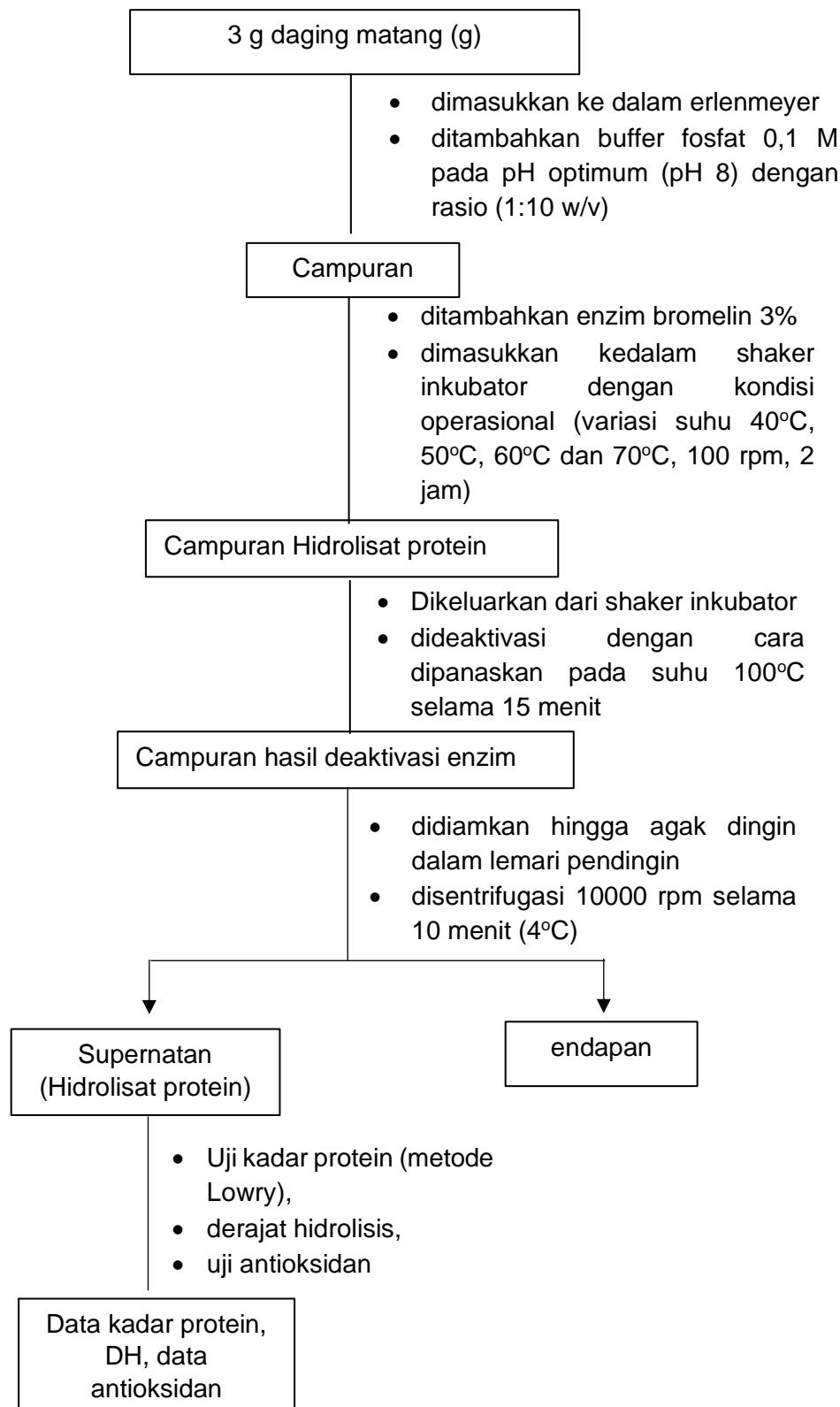
Lampiran 5: Skema Kerja Preparasi Ikan Sidat



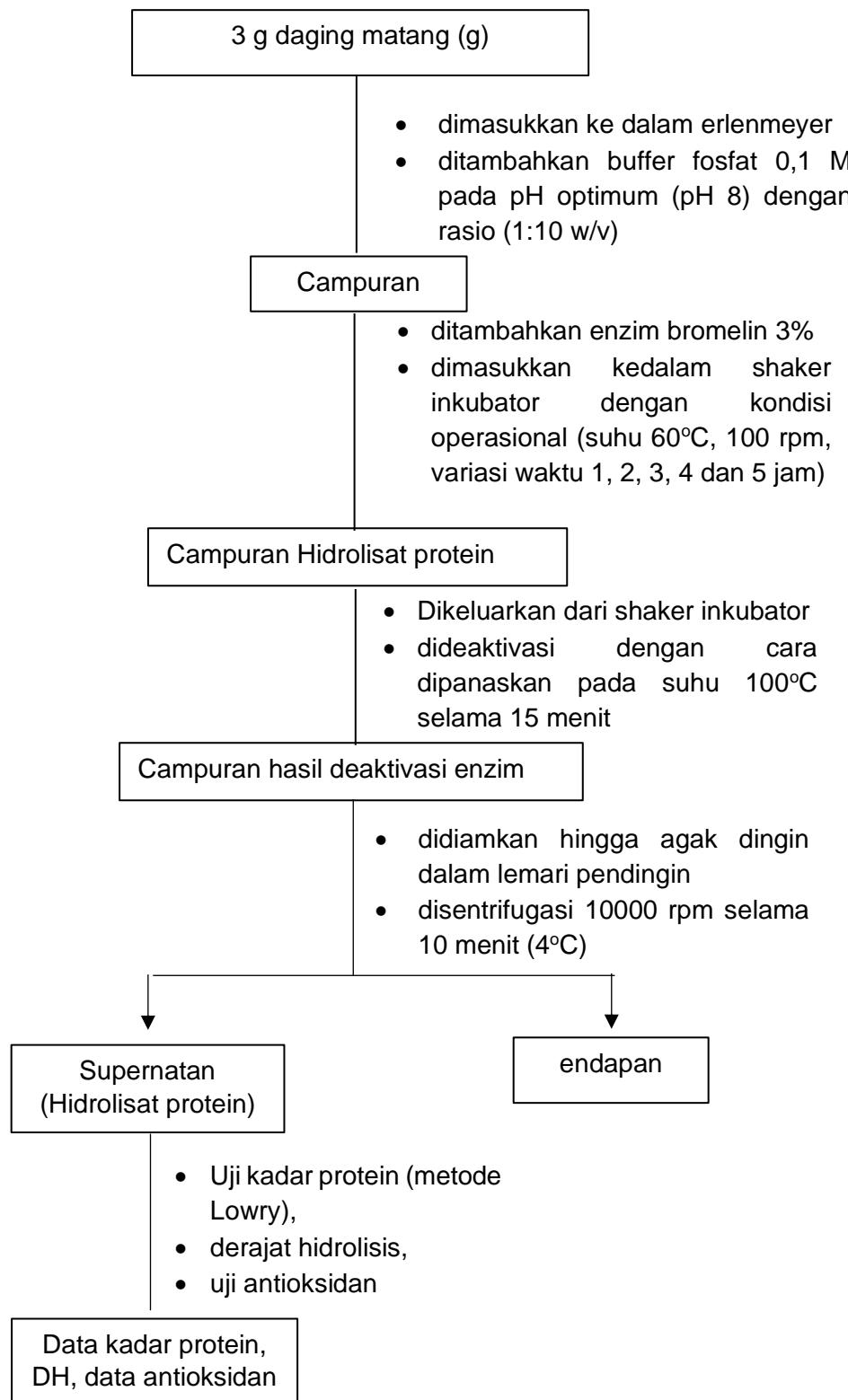
Lampiran 6: Skema Kerja Penentuan pH Optimum Hidrolisis Protein Ikan Sidat



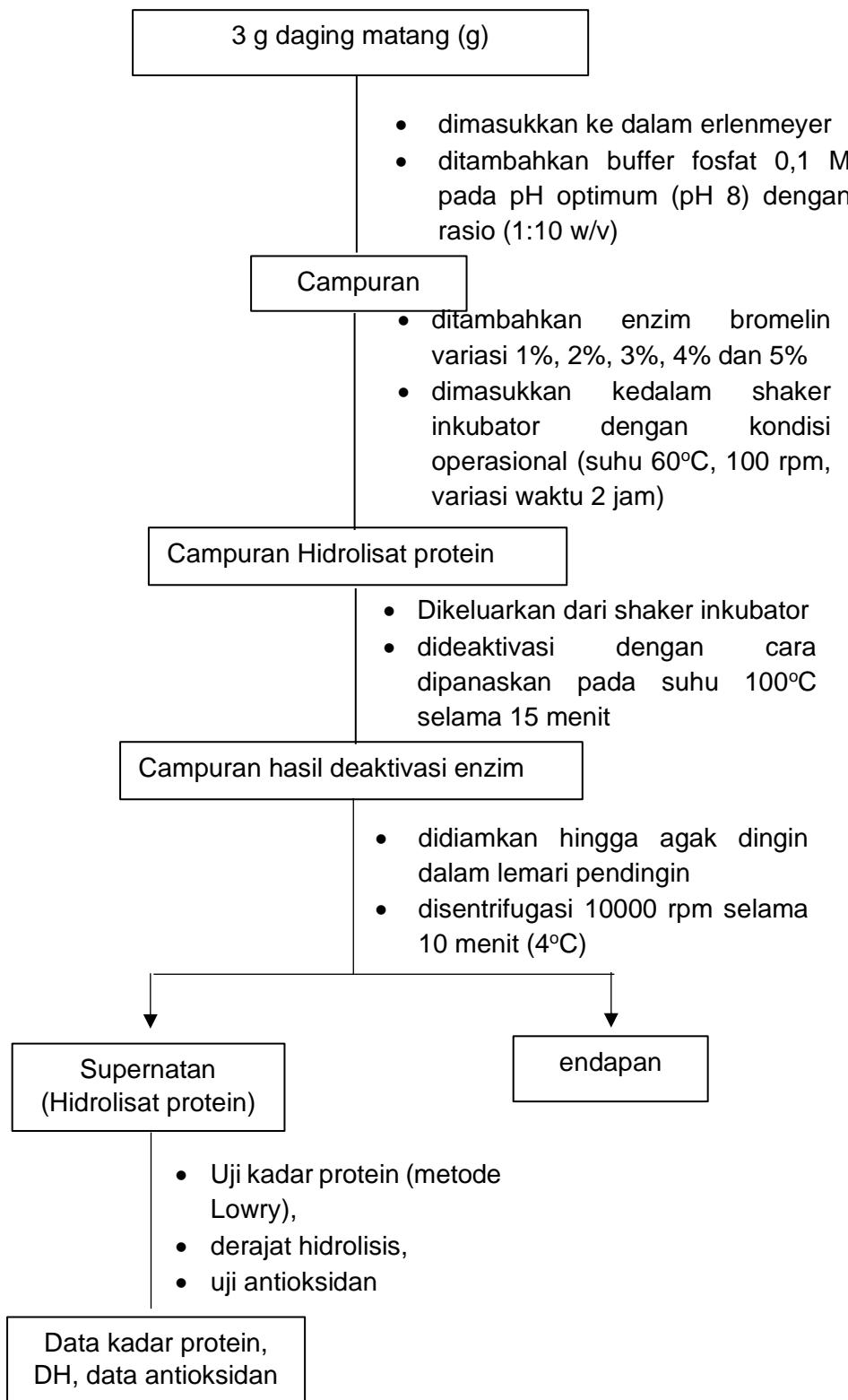
Lampiran 7: Skema Kerja Penentuan Suhu Optimum Hidrolisis Protein Ikan Sidat



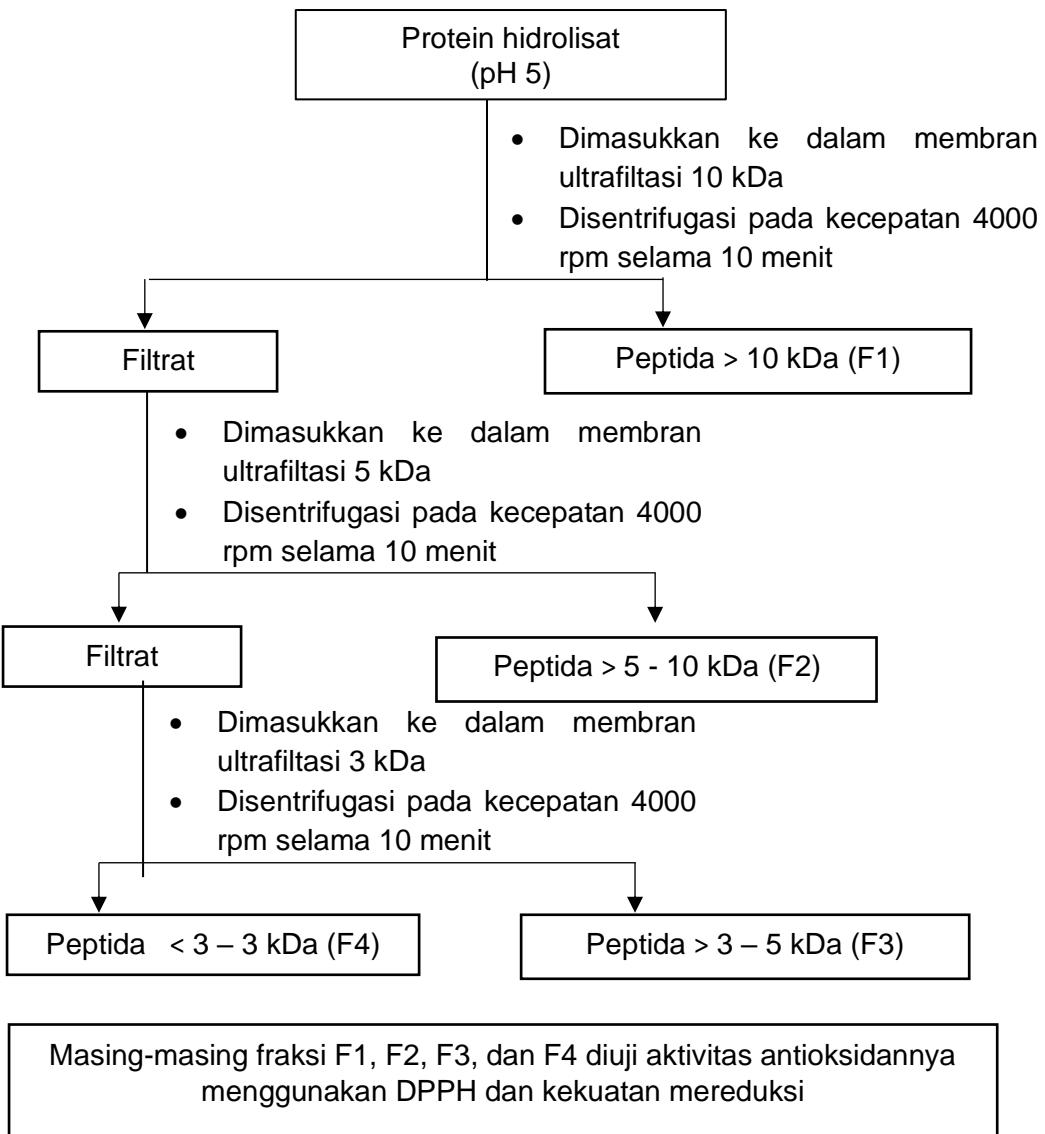
Lampiran 8: Skema Kerja Penentuan Waktu Hidrolisis Optimum Protein Ikan Sidat



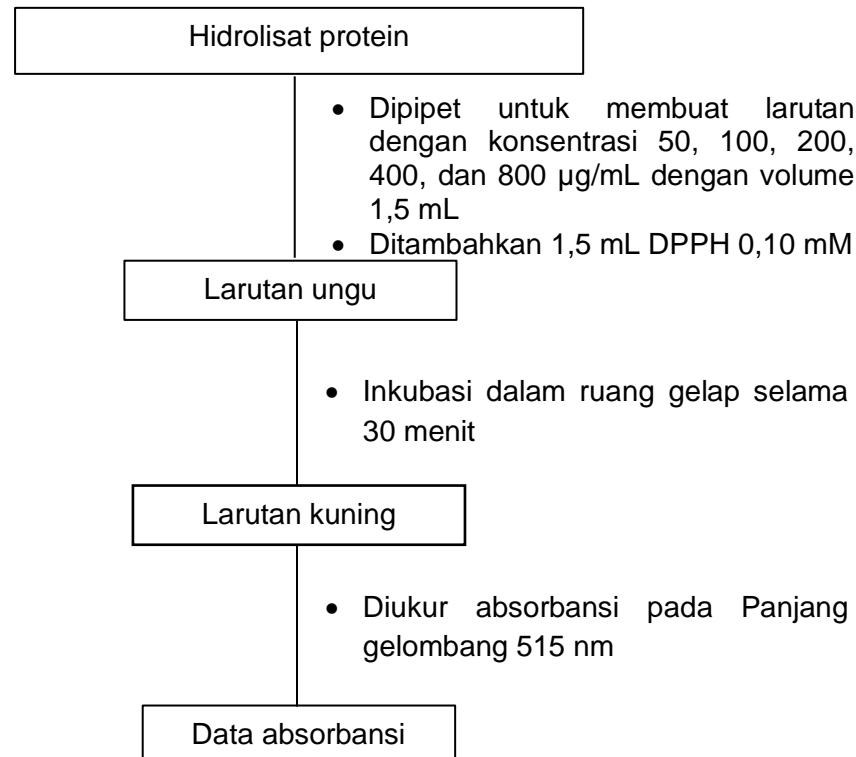
Lampiran 9: Skema Kerja Optimum Konsentrasi Enzim pada Hidrolisis Protein Ikan Sidat



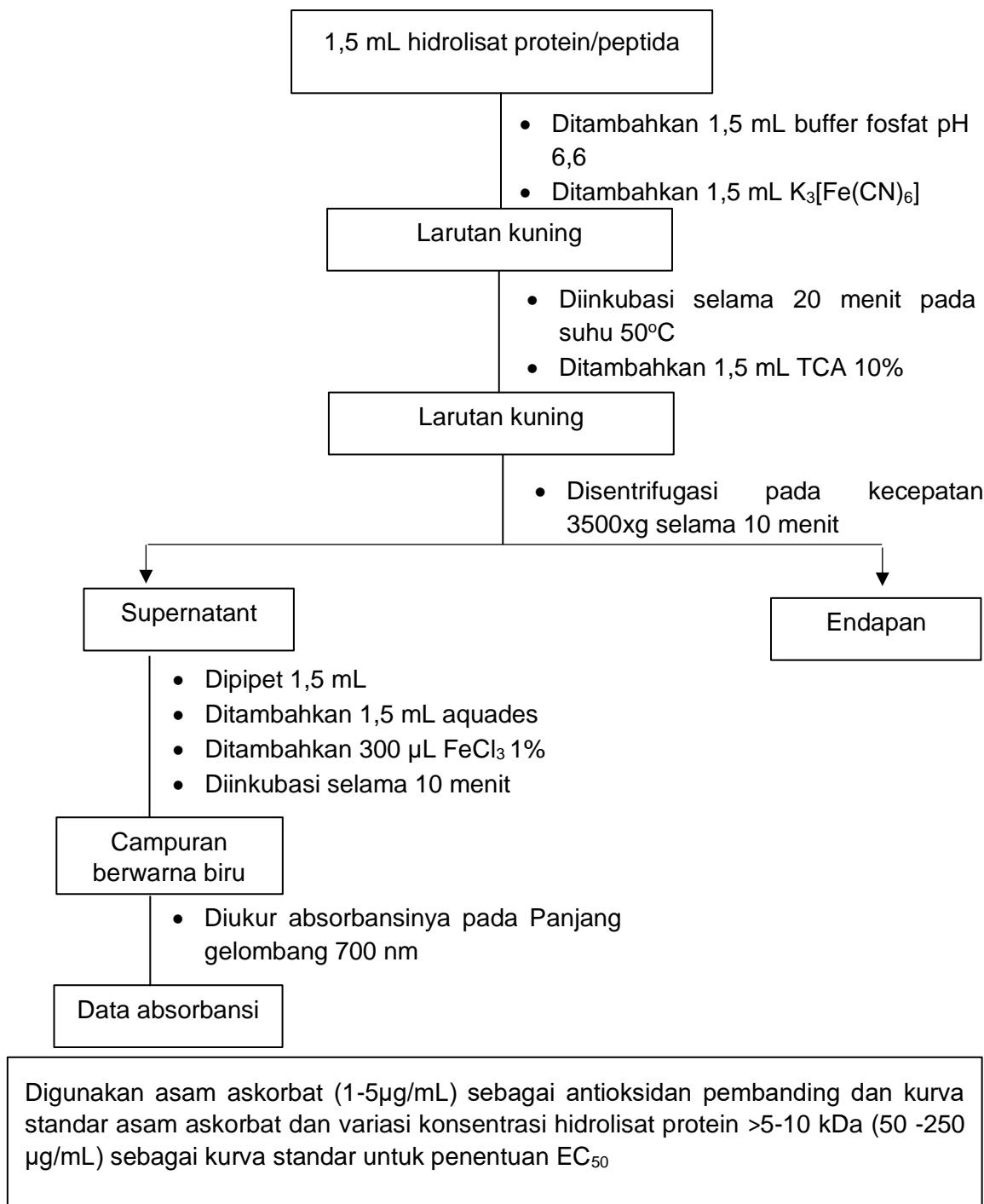
Lampiran 10: Skema Kerja Ultrafiltrasi Hidrolisat Protein Ikan Sidat



Lampiran 11: Skema Kerja Penentuan Aktivitas Antioksidan Hidrolisat Protein dan peptida dengan Metode DPPH

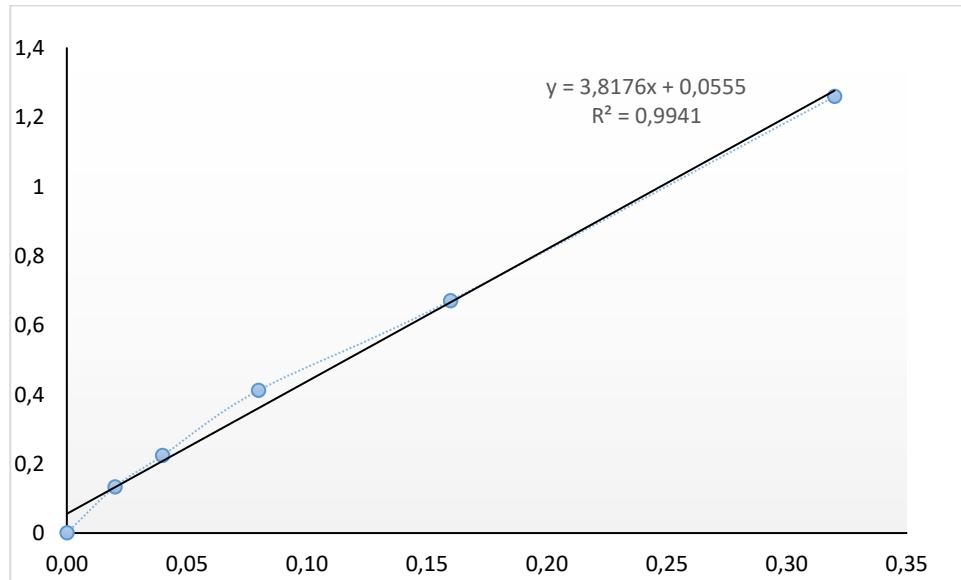


Lampiran 12: Skema Kerja Pengujian Kekuatan Reduksi Fe³⁺ Hidrolisat Protein dan Peptida



Lampiran 13. Penentuan Kadar Protein

1. Pembuatan kurva standar



2. Data penentuan kadar protein ekstrak kasar, fraksi 1, fraksi 2, filtrat sisa dan dialisat

Fraksi	Absorbansi Simplo	Absorbansi Duplo	Rata-rata	a	b	fp	Kadar Protein (mg/mL)
Ekstrak Kasar (EK)	0,482	0,482	0,482	3,8176	0,0555	50	5,586
F1	0,758	0,825	0,7915	3,8176	0,0555	50	9,640
F2	0,325	0,354	0,3395	3,8176	0,0555	50	3,720
Dialisat F2	0,139	0,135	0,137	3,8176	0,0555	50	1,067

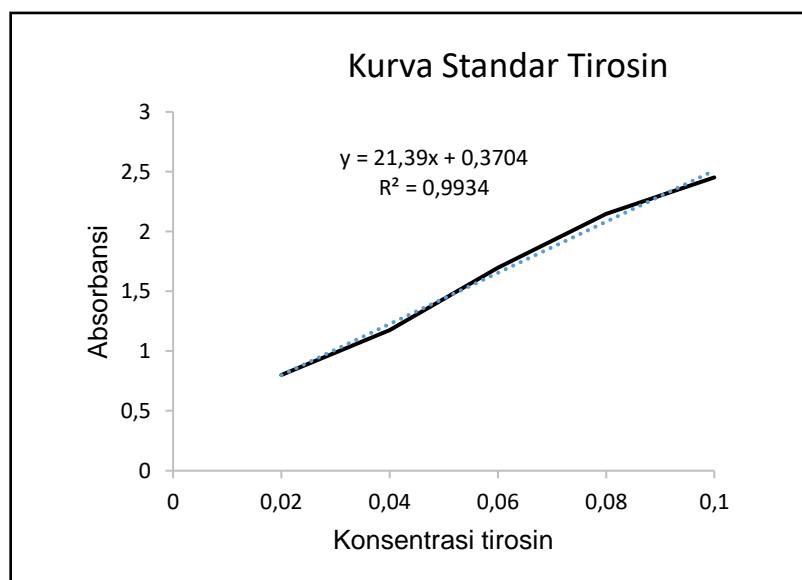
Contoh perhitungan kadar protein:

$$\text{Kadar protein} = \frac{\text{Absorbansi rata-rata} - 0,0555}{3,8176} \times fp$$

$$\text{Kadar protein EK} = \frac{0,482 - 0,0555}{3,8176} \times 50 = 5,586$$

Lampiran 14. Penentuan Aktivitas Katalitik Enzim Bromelin

1. Pembuatan Kurva Standar Tirozin



Data aktivitas

Fraksi	Abs	Kadar tirozin yang dilepaskan ($\mu\text{g/mL}$)	Kadar protein (mg/mL)	Aktivitas Enzim (U/mL)	Aktivitas spesifik (U/mg)	Kemurnian
EK	1,235	40,421	5,586	113,178	20,261	1,00
0-40% (F1)	1,384	47,387	9,640	132,683	13,764	0,68
40-60% (F2)	1,275	42,291	3,720	118,414	31,835	1,57
Dialisat F2	1,535	54,446	1,067	152,449	142,819	7,05

Contoh perhitungan aktivitas enzim, aktivitas spesifik dan tingkat kemurnian:

- Aktivitas Enzim =
$$\frac{(\text{Total } \frac{\mu\text{g}}{\text{mL}} \text{tyrosine dilepas} \times \text{total volume reaksi})}{(\text{Volume enzim} \times \text{waktu reaksi} \times \text{volume analisis})}$$

$$\text{Aktivitas Enzim} = \frac{(47,387 \times 2,1 \text{ ml})}{(0,1 \text{ mL} \times 10 \text{ menit} \times 0,75 \text{ ml})} = 132,68$$

- Aktivitas spesifik enzim = $\frac{\text{Aktivitas enzim}}{\text{kadar protein}}$
 $= \frac{132,68 \text{ U/mL}}{9,640 \text{ mg/mL}} = 13,764 \text{ U/mg}$
- Tingkat kemurnian = $\frac{\text{Aktivitas spesifik enzim setelah fraksinasi}}{\text{Aktivitas spesifik enzim ekstrak kasar}}$
 $= \frac{13,764 \text{ U/mg}}{20,261 \text{ U/mg}} = 0,68$

**Lampiran 15. Data Hasil Analisis Proksimat pada Sampel Ikan Sidat
*A.marmorata***



LABORATORIUM BIOTEKNOLOGI TERPADU PETERNAKAN
FAKULTAS PETERNAKAN
UNIVERSITAS HASANUDDIN
Alamat: Jl. Perintis Kemerdekaan KM. 10 Tamalanrea, Makassar
Email: labbioternakfapetuh@gmail.com

No.Dok.: FSPO-LBTK-UH-12.2

SERTIFIKAT HASIL UJI

No.: 008/T/LBTK-UH/I/2022

Informasi Pelanggan

Nama Perusahaan/Pelanggan	:	Jumardi
Alamat Lengkap	:	Pascasarjana Universitas Hasanuddin
No. Telp./faks./e-mail	:	085342617902
Personel Penghubung	:	081241981874

Informasi Sampel

No. Identitas Laboratorium	:	008/LBTK-RK/I-2022
Uraian/Matriks Sampel	:	-
Kondisi Saat Diterima	:	Baik
Tanggal Diterima	:	18/1/2022
Tanggal Pengujian	:	20/1/2022
Tujuan Pengujian	:	-

Informasi Hasil Pengujian

No	Kode Sampel	PARAMETER UJI				
		Kadar Air (%) (AOAC 930.15)	Kadar Abu (% BK) (AOAC 942.05)	Kadar Protein Kasar (% BK) (AOAC 984.13)	Kadar Lemak Kasar (% BK) (AOAC 920.39)	Kadar Serat Kasar (% BK) (AOAC 962.09)
1	Ikan	73,52	1,94	27,00	19,79	0,11

Ket: 1. Kadar air ditetapkan sesuai sampel uji; 2. Selain kadar air, parameter ditetapkan berdasarkan 100% BK; 3. Lembaran sertifikat hasil uji ini tertelusur; 4. Hasil hanya berhubungan dengan contoh yang diuji dan laporan ini tidak boleh digandakan

Makassar, 7 Februari 2022
 Devisi Teknis

Dr. Ir. Syahriani Syahrir, M.Si.
 NIP.: 196511121990032001

**Lampiran 16. Data Hasil Analisis Asam-Asam Amino pada Sampel Ikan Sidat
A. marmorata**



Result of Analysis | Page 2 of 3

RESULT OF ANALYSIS

Laporan Hasil Pengujian : SIG.LHP.VII.2021.084174

No.	Parameter	Unit	Result		Limit Of Detection	Method
			Simplo	Duplo		
1	L-Metionin	mg / kg	3931.70	3930.48	-	18-12-38/MU/SMM-SIG (LC MS/MS)
2	L-Sistin	mg / kg	5242.79	5240.48	-	18-12-38/MU/SMM-SIG (LC MS/MS)
3	L-Histidin	mg / kg	7865.17	7898.91	-	18-5-17/MU/SMM-SIG (UPLC)
4	L-Threonin	mg / kg	9672.58	9712.21	-	18-5-17/MU/SMM-SIG (UPLC)
5	L-Prolin	mg / kg	6596.73	6626.59	-	18-5-17/MU/SMM-SIG (UPLC)
6	L-Tirosin	mg / kg	6917.11	6953.73	-	18-5-17/MU/SMM-SIG (UPLC)
7	L-Leusin	mg / kg	15493.54	15583.70	-	18-5-17/MU/SMM-SIG (UPLC)
8	L-Asam Aspartat	mg / kg	14223.50	14282.49	-	18-5-17/MU/SMM-SIG (UPLC)
9	L-Lisin	mg / kg	14940.74	15049.09	-	18-5-17/MU/SMM-SIG (UPLC)
10	Glisin	mg / kg	10370.35	10447.09	-	18-5-17/MU/SMM-SIG (UPLC)

No.	Parameter	Unit	Result		Limit Of Detection	Method
			Simplo	Duplo		
11	L-Arginin	mg / kg	12836.29	12910.10	-	18-5-17/MU/SMM-SIG (UPLC)
12	L-Alanin	mg / kg	10164.87	10223.26	-	18-5-17/MU/SMM-SIG (UPLC)
13	L-Valin	mg / kg	9650.25	9688.92	-	18-5-17/MU/SMM-SIG (UPLC)
14	L-Isoleusin	mg / kg	9003.04	9036.41	-	18-5-17/MU/SMM-SIG (UPLC)
15	L-Fenilalanin	mg / kg	9460.69	9494.26	-	18-5-17/MU/SMM-SIG (UPLC)
16	L-Asam glutamat	mg / kg	23857.76	23980.31	-	18-5-17/MU/SMM-SIG (UPLC)
17	L-Serin	mg / kg	8324.97	8340.97	-	18-5-17/MU/SMM-SIG (UPLC)

Bogor, 08 Juli 2021
PT. Saraswanti Indo Genetech



Dwi Yulianto Laksono, S.Si
GM Laboratorium

Lampiran 17. Data Perhitungan Derajat Hidrolisis

1. Kontrol

	Kadar Protein Terlarut (mg/mL)	Kadar Protein dalam TCA (mg/mL)	DH (%)
Kontrol	6,5	0,1768	2,72

2. Pengaruh pH

pH	Kadar Protein Terlarut (mg/mL)	Kadar Protein dalam TCA (mg/mL)	DH (%)
5	6,50	2,796	43,0
6	6,50	1,631	25,1
7	6,50	3,602	55,4
8	6,50	3,949	60,8
9	6,50	3,556	54,7

3. Pengaruh suhu

Suhu (°C)	Kadar Protein Terlarut (mg/mL)	Kadar Protein dalam TCA (mg/mL)	DH (%)
40	6,50	2,397	36,9
50	6,50	2,528	38,9
60	6,50	2,986	45,9
70	6,50	1,015	15,6

4. Pengaruh waktu

Waktu (Jam)	Kadar Protein Terlarut (mg/mL)	Kadar Protein dalam TCA (mg/mL)	DH (%)
1	6,50	2,4819	38,2
2	6,50	3,3660	51,8
3	6,50	2,6522	40,8
4	6,50	2,1545	33,1
5	6,50	2,3444	36,1

5. Pengaruh konsentrasi enzim

Konsentrasi Enzim (%)	Kadar Protein Terlarut (mg/mL)	Kadar Protein Dalam TCA (mg/mL)	DH (%)
1%	6,50	1,6110	24,8
2%	6,50	2,7177	41,8
3%	6,50	3,9816	61,3
4%	6,50	3,5952	55,3
5%	6,50	3,4577	53,2

Contoh perhitungan derajat hidrolisis (DH)

$$DH = \frac{\text{Protein terlarut dalam TCA}}{\text{Protein total terlarut dalam sampel}} \times 100\%$$

$$DH = \frac{0,1768 \text{ mg/mL}}{6,500 \text{ mg/mL}} \times 100\% = 2,72\%$$

Lampiran 18. Pengujian Antioksidan dengan DPPH

Hidrolisat protein

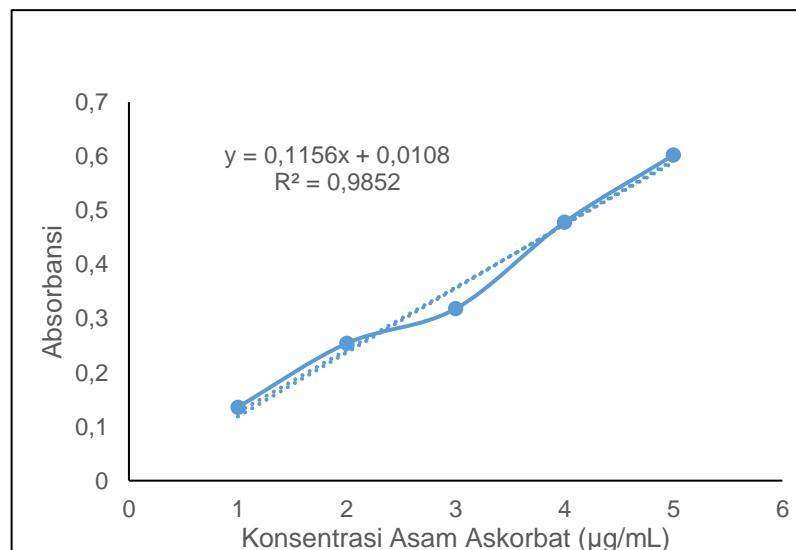
Data hasil uji antioksidan dari 20 sampel hidrolisat protein

No	Sampel	DPPH	
		Abs	% inhibisi
1	Blanko	0,654	-
2	Kontrol	0,577	11,8
3	pH 5	0,130	80,1
4	pH 6	0,368	43,7
5	pH 7	0,474	27,5
6	pH 8	0,455	30,4
7	pH 9	0,514	21,4
8	40°C	0,575	12,1
9	50 °C	0,535	18,2
10	60 °C	0,582	11,0
11	70 °C	0,415	36,5
12	1 h	0,519	20,6
13	2 h	0,430	34,3
14	3 h	0,534	18,3
15	4 h	0,462	29,4
16	5 h	0,441	32,6
17	1%	0,569	13,0
18	2%	0,561	14,2
19	3%	0,505	22,8
20	4%	0,445	32,0
21	5%	0,434	33,6

Lampiran 19. Kekuatan Mereduksi Fe³⁺ dari Hidrolisat Protein

Kurva Standar Asam askorbat

Asam Askorbat		
No	Conc (µg/mL)	Abs
1	1	0,136
2	2	0,254
3	3	0,318
4	4	0,478
5	5	0,602



Lanjutan lampiran 19

Data hasil kekuatan mereduksi dari 20 sampel hidrolisat protein

No	Sampel	Power Reduksi	
		Abs	FRAP (mg AA/g HP)
1	Blanko	0	0,00
2	Kontrol	0,12	0,91
3	pH 5	1,1	9,08
4	pH 6	0,5	4,11
5	pH 7	0,2	1,75
6	pH 8	0,4	3,19
7	pH 9	0,8	6,86
8	40°C	0,3	0,94
9	50 °C	0,3	2,71
10	60 °C	0,3	2,59
11	70 °C	0,4	2,53
12	1 h	0,4	3,54
13	2 h	0,5	0,94
14	3 h	0,2	3,33
15	4 h	0,2	3,97
16	5 h	0,4	1,94
17	1%	0,6	2,05
18	2%	0,15	3,07
19	3%	0,3	0,94
20	4%	0,3	4,70
21	5%	0,3	1,19

Contoh perhitungan nilai FRAP :

Dari persamaan regresi pada kurva standar asam askorbat diperoleh persamaan:

$$y = 0,1156x + 0,0108$$

$$x = \frac{y - 0,0108}{0,1156}$$

Nilai Y = nilai absorbansi hidrolisat protein

$$x = \frac{1,1 - 0,0108}{0,1156} = 9,08$$

nilai FRAP = 3,38 mg asam askorbat/ g hidrolisat protein

Lampiran 20. Kadar Protein Hasil Ultrafiltrasi

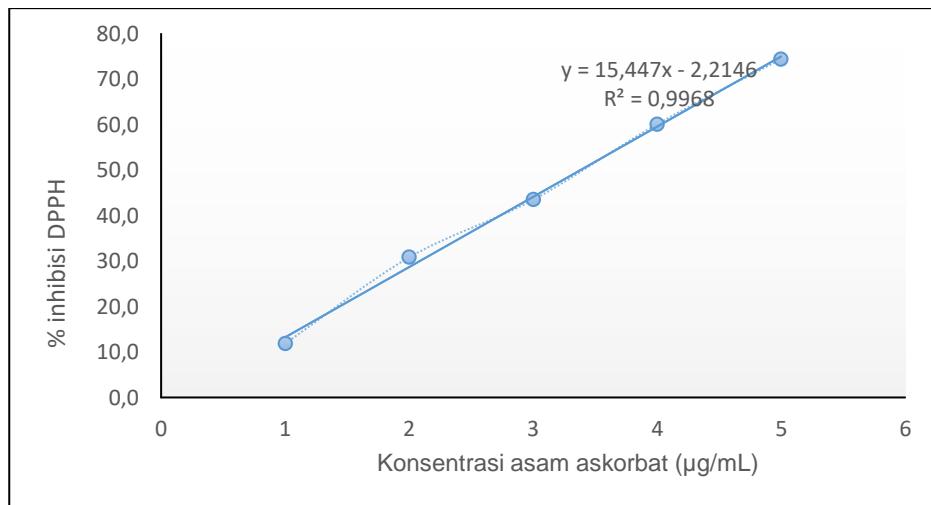
Kadar Protein fraksi hidrolisat protein hasil ultrafiltrasi

Protein	Abs		Rata-Rata	a	b	Conc mg/mL I
	I	II				
>10 kDa	0,351	0,365	0,358	3,8176	0,0555	3,962
10-5 kDa	0,462	0,469	0,466	3,8176	0,0555	5,370
5-3 kDa	0,373	0,365	0,369	3,8176	0,0555	4,107
<3 kDa	0,288	0,278	0,283	3,8176	0,0555	2,980

Lampiran 21. Pengujian Antioksidan Dengan DPPH Dan Kekuatan Mereduksi Fe³⁺ dari Peptida Hasil Ultrafiltrasi

1. Kurva Standar asam askorbat

No	Konsentrasi ($\mu\text{g/mL}$)	Abs	Abs	% inhibisi
1	Kontrol	0,727	0,727	-
2	1	0,638	0,641	11,8
3	2	0,515	0,503	30,8
4	3	0,401	0,411	43,5
5	4	0,291	0,290	60,1
6	5	0,179	0,186	74,4



Contoh perhitungan persen inhibisi:

$$\text{Perhitungan \% inhibisi} = \frac{\text{Abs Blanko} - \text{Abs sampel}}{\text{Abs Blanko}} \times 100\%$$

$$\text{Perhitungan \% inhibisi} = \frac{0,727 - 0,641}{0,727} \times 100\% = 11,8\%$$

Contoh perhitungan IC₅₀:

Dari persamaan regresi pada kurva standar DPPH diperoleh persamaan:

$$y = 15,447x - 2,2146$$

$$x = \frac{y + 2,2146}{15,447}$$

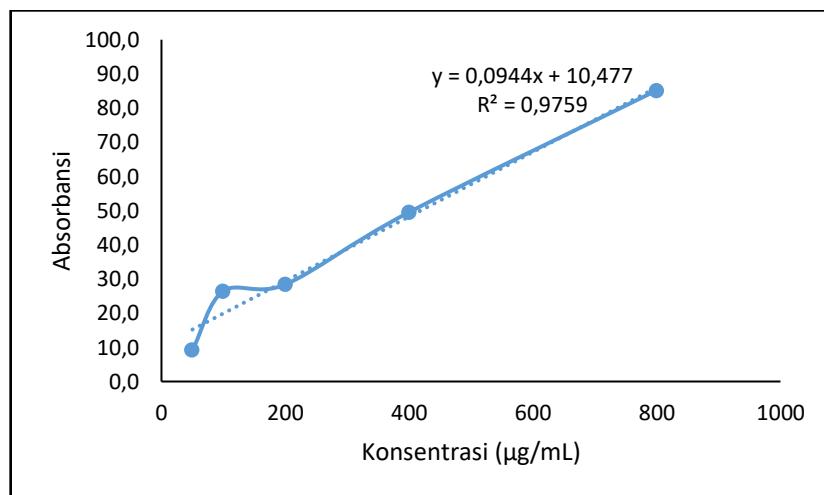
$$\text{Nilai Y} = 50$$

$$x = \frac{50 + 2,2146}{15,447} = 3,38$$

$$\text{nilai IC}_{50} = 3,38 \mu\text{g/mL} \text{ atau } 0,0034 \text{ mg/mL}$$

2. Kurva Standar DPPH untuk sampel 10 kDa

No	Conc ($\mu\text{g/mL}$)	Abs	Abs	Abs rata-rata	% inhibisi
1	Kontrol	0,603	0,637	0,620	-
2	50	0,570	0,556	0,563	9,2
3	100	0,450	0,462	0,456	26,5
4	200	0,447	0,441	0,444	28,4
5	400	0,311	0,315	0,313	49,5
6	800	0,094	0,090	0,092	85,2



Contoh perhitungan persen inhibisi:

$$\text{Perhitungan \% inhibisi} = \frac{\text{Abs Blanko} - \text{Abs sampel}}{\text{Abs Blanko}} \times 100\%$$

$$\text{Perhitungan \% inhibisi} = \frac{0,631 - 0,466}{0,631} \times 100\% = 26,1\%$$

Contoh perhitungan IC₅₀:

Dari persamaan regresi pada kurva standar DPPH diperoleh persamaan:

$$y = 0,0944x + 10,477$$

$$x = \frac{y - 10,477}{0,0944}$$

$$\text{Nilai } Y = 50$$

$$x = \frac{50 - 10,477}{0,0944} = 418$$

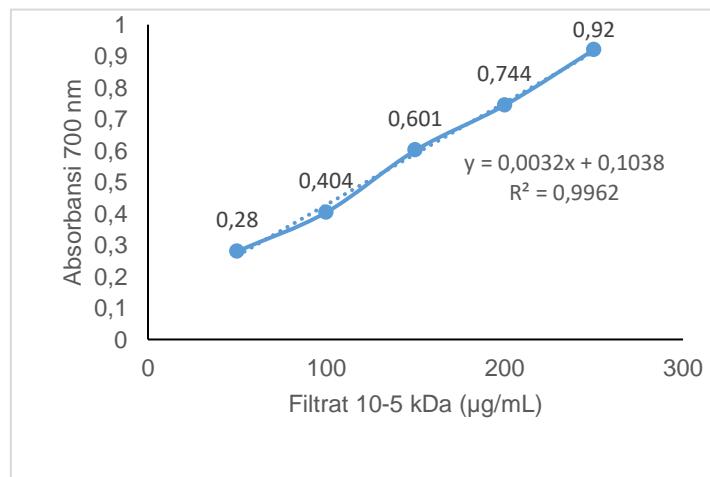
$$\text{nilai IC}_{50} = 418 \mu\text{g/mL} \text{ atau } 0,418 \text{ mg/mL}$$

3. Aktivitas antioksidan terhadap DPPH

No	Sampel	DPPH		
		Abs	% inhibisi	IC ₅₀ (mg/mL)
1	Kontrol	0,631	-	-
2	> 10 kDa	0,485	23,1	-
3	10-5 kDa	0,409	35,2	0,418
4	5-3 kDa	0,465	26,3	-
5	<3 kDa	0,466	26,1	-
6	Asam askorbat			0,0034

4. Kekuatan mereduksi Fe³⁺ dari fraksi hidrolisat

No	Sampel	Metode FRAP		
		Abs	Nilai FRAP	EC ₅₀ (mg/mL)
1	Kontrol	0,12	0,91	-
2	> 10 kDa	0,65	8,1	-
3	10-5 kDa	0,95	7,9	15,59
4	5-3 kDa	0,92	7,6	-
5	<3 kDa	0,89	5,6	-



Dari persamaan regresi pada kurva standar DPPH diperoleh persamaan:

$$y = 0,0032x + 0,1038$$

$$x = \frac{y - 0,1038}{0,0032}$$

$$\text{Nilai Y} = 50$$

$$x = \frac{50 - 0,1038}{0,0032} = 15592,56$$

nilai IC₅₀ = 15592, 56 µg/mL atau 15, 59 mg/mL

Lampiran 22. Dokumentasi Penelitian



Isolasi Enzim bromelin dari buah nanas



Sampel Ikan sidat (*Anguilla marmorata*)



Hidrolisis protein ikan sidat menggunakan enzim bromelin



Pengujian protein



Pengujian antioksidan DPPH dan kekuatan mereduksi hidrolisat protein



Pengujian antioksidan DPPH dan kekuatan mereduksi hasil ultrafiltrasi