

博士論文

**Metabolomic Study of Food Derived Compounds Interacting
with ABC Drug Transporters**

ABC 薬物トランスポーターと相互作用する食物由来化合物を 対象とした
メタボロミクス研究

金沢大学大学院医薬保健学総合研究科創薬科学専攻
分子薬物治療学研究室

学籍番号	1829012007
学生氏名	RINA AGUSTINA
主任指導教員	加藤 将夫 教授
論文提出	令和 3 年 7 月

Table of Contents

Abbreviations	- 3 -
Abstract	- 4 -
General Introduction	- 5 -
I. Untargeted metabolomic analysis to identify endogenous BCRP substrates	
Introduction	- 7 -
Results	- 9 -
Discussion	- 29 -
Conclusion	- 32 -
II. Effect of P-gp inhibitors on disposition of food derived steroidal alkaloids	
Introduction	- 33 -
Results	- 35 -
Discussion	- 43 -
Conclusion	- 44 -
General Conclusion	- 45 -
Materials and Methods	- 46 -
Supplementary Figures	- 55 -
Supplementary Tables	- 57 -
References	- 58 -
Acknowledgements	- 64 -

Abbreviations

AIF	all ion fragmentation
AUC	area under the plasma concentration–time curve
BCRP	breast cancer resistance protein
BS	biochanin A sulfate
C _{max}	maximum plasma concentration
DDI	drug-drug interaction
DS	daidzein sulfate
ES	equol sulfate
ESI	electrospray ionization
GS	genistein sulfate
iPS	induced pluripotent stem
LC-MS/MS	liquid chromatography with tandem mass spectrometry
LC-TOFMS	liquid chromatography time-of-flight mass spectrometry
LY	lucifer yellow
NMQ	N-methyl-quinidine
P-gp	P-glycoprotein
PLS-DA	projection to latent structures-discriminant analysis
SULT	sulfotransferase
TKO	triples knock out (<i>Abcb1a/1b/Abcg2</i> ^{-/-})

ABSTRACT

Breast cancer resistance protein (BCRP) and P-glycoprotein (P-gp) belong to ABC drug transporters. They are localized on the apical side of plasma membranes in various organs and extrude their substrates from the cells. Potential inhibition of these transporters is a crucial issue during drug development, and utilization of its physiological substrates as biomarkers would be advantageous to evaluate the inhibition potential. The purpose of this study was to identify such BCRP/P-gp substrates in the body using metabolomics approaches. In BCRP study, mice were fed with a roasted soybean flour-containing diet which may highly include BCRP substrates isoflavones. Lapatinib was orally administered as a BCRP inhibitor, and plasma samples were collected. Ion peaks in the plasma samples were comprehensively measured by LC-TOFMS using all ion fragmentation mode and quantified by LC-MS/MS. Another screening with a similar sequence of analysis was also performed in plasma samples of *Bcrp*^{-/-} and wild-type mice. Untargeted metabolomics with neutral loss search of sulfate moiety has revealed increase in plasma peaks of daidzein sulfate (DS) and genistein sulfate (GS) by lapatinib administration and *Bcrp*^{-/-} mice. Oral administration of lapatinib significantly increased AUC_{0-7h} for DS, GS, and equal sulfate (ES) by 3.56-, 5.60-, and 1.57-fold, respectively. Another BCRP inhibitor febuxostat also increased AUC_{0-7h} of DS, GS, and ES. To demonstrate the effect of BCRP inhibitors on the intestinal disposition of the isoflavone sulfates, their appearance in the basal side of human iPS cell-derived small intestinal epithelial-like cells (F-hiSIEC) was examined to mimic physiological intestinal transport. After addition of daidzein, genistein, and equol to the medium of apical side, the appearance of DS, GS, and ES in the basal compartment significantly increased in the presence of lapatinib and febuxostat, suggesting that these BCRP inhibitors may inhibit intestinal BCRP and thereby increase appearance of the isoflavone sulfates. Both inhibitors reduced ATP-dependent uptake of DS and ES in BCRP-expressing membrane vesicles, indicating inhibition of BCRP-mediated DS and ES transport. In P-gp study, effect of P-gp inhibitors on the disposition of orally administered steroidal alkaloids derived from tomatoes, tomatidine and esculeogenin A, was examined since P-gp preferably recognizes steroidal compounds as substrates. The plasma concentration of a typical P-gp substrate fexofenadine was increased following oral administration of P-gp inhibitors elacridar and tariquidar, whereas AUC_{0-6h} and C_{max} of tomatidine was not significantly changed. On the other hand, these inhibitors significantly increased brain-to-plasma concentration ratio (K_{p,brain}) of tomatidine by 2.38- and 2.41-fold, respectively. Plasma concentration and K_{p,brain} of esculeogenin A were not changed by coadministration of elacridar. AUC_{0-6h} and C_{max} of tomatidine were significantly increased in *Abcb1a/1b/Abcg2*^{-/-} (TKO) mice compared with wild-type mice by 2.81- and 2.52-fold, respectively, whereas K_{p,brain} of tomatidine in TKO was also significantly higher than wild-type mice by 2.98-fold. ATP-dependent uptake of tomatidine was observed in P-gp expressing membrane vesicles, and its uptake was reduced by elacridar and tariquidar. On the other hand, ATP-dependent uptake of tomatidine was not observed in the BCRP-expressing membrane vesicles. These results suggest P-gp may play some roles in distribution of tomatidine to the brain, but tomatidine may not be a useful biomarker to assess P-gp inhibition by the drugs. In conclusion, isoflavone sulfates were proposed as the first evidence to be a surrogate marker of BCRP inhibition, whereas further studies are required to assess a surrogate marker of P-gp inhibition.

Key words: Metabolomic study, BCRP, P-gp, biomarker

References

- Álvarez AI, Vallejo F, Barrera B, Merino G, Prieto JG, Tomás-Barberán F, and Espín JC (2011) Bioavailability of the glucuronide and sulfate conjugates of genistein and daidzein in breast cancer resistance protein 1 knockout mice. *Drug Metab Dispos* **39**:2008–2012.
- An G, and Morris ME (2011) The sulfated conjugate of biochanin A is a substrate of breast cancer resistant protein (ABCG2). *Biopharm Drug Dispos* **32**:446–57.
- Bankstahl JP, Bankstahl M, Römermann K, Wanek T, Stanek J, Windhorst AD, Fedrowitz M, Erker T, Müller M, Löscher W, Langer O, and Kuntner C (2013) Tariquidar and elacridar are dose-dependently transported by P-glycoprotein and Bcrp at the blood-brain barrier: A small-animal positron emission tomography and in vitro study. *Drug Metab Dispos* **41**:754–762.
- Chong J, Wishart DS, and Xia J (2019) Using MetaboAnalyst 4.0 for Comprehensive and Integrative Metabolomics Data Analysis. *Curr Protoc Bioinforma* **68**.
- Chu QSC, Cianfrocca ME, Goldstein LJ, Gale M, Murray N, Loftiss J, Arya N, Koch KM, Pandite L, Fleming RA, Paul E, and Rowinsky EK (2008) A phase I and pharmacokinetic study of lapatinib in combination with letrozole in patients with advanced cancer. *Clin Cancer Res* **14**:4484–4490.
- Cichon MJ, Riedl KM, Wan L, Thomas-Ahner JM, Francis DM, Clinton SK, and Schwartz SJ (2017) Plasma Metabolomics Reveals Steroidal Alkaloids as Novel Biomarkers of Tomato Intake in Mice. *Mol Nutr Food Res* **61**.
- da Silva DC, Andrade PB, Valentão P, and Pereira DM (2017) Neurotoxicity of the steroidal alkaloids tomatine and tomatidine is RIP1 kinase- and caspase-independent and involves the eIF2 α branch of the endoplasmic reticulum. *J Steroid Biochem Mol Biol* **171**:178–186.
- Enokizono J, Kusuhara H, and Sugiyama Y (2007a) Effect of breast cancer resistance protein (Bcrp/Abcg2) on the disposition of phytoestrogens. *Mol Pharmacol* **72**:967–75.
- Enokizono J, Kusuhara H, and Sugiyama Y (2007b) Regional expression and activity of breast cancer resistance protein (Bcrp/Abcg2) in mouse intestine: overlapping distribution with sulfotransferases. *Drug Metab Dispos* **35**:922–8.
- Friedman M (2015) Chemistry and Anticarcinogenic Mechanisms of Glycoalkaloids Produced by Eggplants, Potatoes, and Tomatoes. *J Agric Food Chem* **63**:3323–3337.

- Fujiwara Y, Kiyota N, Tsurushima K, Yoshitomi M, Horlad H, Ikeda T, Nohara T, Takeya M, and Nagai R (2012) Tomatidine, a tomato saponin, ameliorates hyperlipidemia and atherosclerosis in ApoE-deficient mice by inhibiting acyl-CoA:cholesterol acyl-transferase (ACAT). *J Agric Food Chem* **60**:2472–2479.
- Gotanda K, Tokumoto T, Hirota T, Fukae M, and Ieiri I (2015) Sulfasalazine disposition in a subject with 376C>T (nonsense mutation) and 421C>A variants in the ABCG2 gene. *Br J Clin Pharmacol* **80**:1236–1237.
- Harvey RD, Aransay NR, Isambert N, Lee J-S, Arkenau T, Vansteenkiste J, Dickinson PA, Bui K, Weilert D, So K, Thomas K, and Vishwanathan K (2018) Effect of multiple-dose osimertinib on the pharmacokinetics of simvastatin and rosuvastatin. *Br J Clin Pharmacol* **84**:2877–2888.
- Hosoda K, Furuta T, and Ishii K (2011) Metabolism and disposition of isoflavone conjugated metabolites in humans after ingestion of kinako. *Drug Metab Dispos* **39**:1762–7.
- Ichida K, Matsuo H, Takada T, Nakayama A, Murakami K, Shimizu T, Yamanashi Y, Kasuga H, Nakashima H, Nakamura T, Takada Y, Kawamura Y, Inoue H, Okada C, Utsumi Y, Ikebuchi Y, Ito K, Nakamura M, Shinohara Y, Hosoyamada M, Sakurai Y, Shinomiya N, Hosoya T, and Suzuki H (2012) Decreased extra-renal urate excretion is a common cause of hyperuricemia. *Nat Commun* **3**, Nat Commun.
- Ichikawa M, Negoro R, Kawai K, Yamashita T, Takayama K, and Mizuguchi H (2021) Vinblastine treatment decreases the undifferentiated cell contamination of human iPSC-derived intestinal epithelial-like cells. *Mol Ther Methods Clin Dev* **20**:463–472.
- Iwao T, Kodama N, Kondo Y, Kabeya T, Nakamura K, Horikawa T, Niwa T, Kurose K, and Matsunaga T (2015) Generation of enterocyte-like cells with pharmacokinetic functions from human induced pluripotent stem cells using small-molecule compounds. *Drug Metab Dispos* **43**:603–610.
- Jonker JW, Buitelaar M, Wagenaar E, Van Der Valk M a, Scheffer GL, Scheper RJ, Plosch T, Kuipers F, Elferink RPJO, Rosing H, Beijnen JH, and Schinkel AH (2002) The breast cancer resistance protein protects against a major chlorophyll-derived dietary phototoxin and protoporphyria. *Proc Natl Acad Sci U S A* **99**:15649–54.

- Kabeya T, Mima S, Imakura Y, Miyashita T, Ogura I, Yamada T, Yasujima T, Yuasa H, Iwao T, and Matsunaga T (2020) Pharmacokinetic functions of human induced pluripotent stem cell-derived small intestinal epithelial cells. *Drug Metab Pharmacokinet* **35**:374–382.
- Keskitalo JE, Zolk O, Fromm MF, Kurkinen KJ, Neuvonen PJ, and Niemi M (2009) ABCG2 polymorphism markedly affects the pharmacokinetics of atorvastatin and rosuvastatin. *Clin Pharmacol Ther* **86**:197–203.
- Kim D-H (2015) Gut Microbiota-Mediated Drug-Antibiotic Interactions. *Drug Metab Dispos* **43**:1581–9.
- Kim M, Laramy JK, Gampa G, Parrish KE, Brundage R, Sarkaria JN, and Elmquist WF (2019) Brain distributional kinetics of a novel MDM2 inhibitor SAR405838: Implications for use in brain tumor therapy. *Drug Metab Dispos* **47**:1403–1414.
- Kobayashi Y, Fukami T, Nakajima A, Watanabe A, Nakajima M, and Yokoi T (2012) Species differences in tissue distribution and enzyme activities of arylacetamide deacetylase in human, rat, and mouse. *Drug Metab Dispos* **40**:671–9.
- Kodama N, Iwao T, Katano T, Ohta K, Yuasa H, and Matsunaga T (2016) Characteristic Analysis of Intestinal Transport in Enterocyte-Like Cells Differentiated from Human Induced Pluripotent Stem Cells. *Drug Metab Dispos* **44**:1662–1667.
- Lafaye A, Junot C, Ramounet-Le Gall B, Fritsch P, Ezan E, and Tabet JC (2004) Profiling of sulfoconjugates in urine by using precursor ion and neutral loss scans in tandem mass spectrometry. Application to the investigation of heavy metal toxicity in rats. *J Mass Spectrom* **39**:655–664.
- Lai Y, Mandlekar S, Shen H, Holenarsipur VK, Langish R, Rajanna P, Murugesan S, Gaud N, Selvam S, Date O, Cheng Y, Shipkova P, Dai J, Humphreys WG, and Marathe P (2016) Coproporphyrins in plasma and urine can be appropriate clinical biomarkers to recapitulate drug-drug interactions mediated by organic anion transporting polypeptide inhibition. *J Pharmacol Exp Ther* **358**:397–404.
- Lee CA, O'Connor MA, Ritchie TK, Galetin A, Cook JA, Ragueneau-Majlessi I, Ellens H, Feng B, Taub ME, Paine MF, Polli JW, Ware JA, and Zamek-Gliszczynski MJ (2015) Breast cancer resistance protein (ABCG2) in clinical pharmacokinetics and drug interactions: practical recommendations for clinical victim and perpetrator drug-drug interaction study design. *Drug Metab Dispos* **43**:490–509.
- Lehtisalo M, Keskitalo JE, Tornio A, Lapatto-Reiniluoto O, Deng F, Jaatinen T, Viinamäki J, Neuvonen M, Backman JT, and Niemi M (2020) Febuxostat, But Not Allopurinol, Markedly Raises the Plasma

Concentrations of the Breast Cancer Resistance Protein Substrate Rosuvastatin. *Clin Transl Sci* **13**:1236–1243.

Li W, Sparidans RW, Wang Y, Lebre MC, Beijnen JH, and Schinkel AH (2018) P-glycoprotein and breast cancer resistance protein restrict brigatinib brain accumulation and toxicity, and, alongside CYP3A, limit its oral availability. *Pharmacol Res* **137**:47–55, Elsevier.

Lin JH, and Yamazaki M (2003) Role of P-glycoprotein in pharmacokinetics: clinical implications. *Clin Pharmacokinet* **42**:59–98.

Mao Q, and Unadkat JD (2015) Role of the Breast Cancer Resistance Protein (BCRP/ABCG2) in Drug Transport—an Update. *AAPS J* **17**:65–82.

Margier M, Collet X, Le May C, Desmarchelier C, André F, Lebrun C, Defoort C, Bluteau A, Borel P, Lespine A, and Reboul E (2019) ABCB1 (P-glycoprotein) regulates Vitamin D absorption and contributes to its transintestinal efflux. *FASEB J* **33**:2084–2094.

Miyata H, Takada T, Toyoda Y, Matsuo H, Ichida K, and Suzuki H (2016) Identification of Febuxostat as a New Strong ABCG2 Inhibitor: Potential Applications and Risks in Clinical Situations. *Front Pharmacol* **7**:518.

Mizuno T, Fukudo M, Terada T, Kamba T, Nakamura E, Ogawa O, Inui KI, and Katsura T (2012) Impact of genetic variation in breast cancer resistance protein (BCRP/ABCG2) on sunitinib pharmacokinetics. *Drug Metab Pharmacokinet* **27**:631–639.

Nardone-White DT, Bissada JE, Abouda AA, and Jackson KD (2021) Detoxication versus Bioactivation Pathways of Lapatinib In Vitro: UGT1A1 Catalyzes the Hepatic Glucuronidation of Debenzylated Lapatinib. *Drug Metab Dispos* **49**:233–244.

Obara A, Kinoshita M, Hosoda K, Yokokawa A, Shibasaki H, and Ishii K (2019) Identification of equol-7-glucuronide-4'-sulfate, monoglucuronides and monosulfates in human plasma of 2 equol producers after administration of kinako by LC-ESI-MS. *Pharmacol Res Perspect* **7**:1–11.

Omote H, and Moriyama Y (2018) Reconstitution and Transport Analysis of Eukaryotic Transporters in the Post-Genomic Era. *Methods Mol Biol* **1700**:343–352.

Polli JW, Humphreys JE, Harmon KA, Castellino S, O'Mara MJ, Olson KL, John-Williams LS, Koch KM, and Serabjit-Singh CJ (2008) The role of efflux and uptake transporters in [N-{3-chloro-4-[(3-fluorobenzyl)oxy]phenyl}-6-[5-({[2-(methylsulfonyl)ethyl]amino}methyl)-2-furyl]-4-

quinazolinamine (GW572016, lapatinib) disposition and drug interactions. *Drug Metab Dispos* **36**:695–701.

Sánchez-Mata MC, Yokoyama WE, Hong YJ, and Prohens J (2010) α -Solasonine and α -solamargine contents of gboma (*solanum macrocarpon* l.) and scarlet (*solanum aethiopicum* l.) eggplants. *J Agric Food Chem* **58**:5502–5508.

Schinkel AH, Smit JJM, van Tellingen O, Beijnen JH, Wagenaar E, van Deemter L, Mol CAAM, van der Valk MA, Robanus-Maandag EC, te Riele HPJ, Berns AJM, and Borst P (1994) Disruption of the mouse *mdr1a* P-glycoprotein gene leads to a deficiency in the blood-brain barrier and to increased sensitivity to drugs. *Cell* **77**:491–502.

Shelnutt SR, Cimino CO, Wiggins PA, Ronis MJJ, and Badger TM (2002) Pharmacokinetics of the glucuronide and sulfate conjugates of genistein and daidzein in men and women after consumption of a soy beverage. *Am J Clin Nutr* **76**:588–94.

Shimizu M, Uno T, Sugawara K, and Tateishi T (2006) Effects of itraconazole and diltiazem on the pharmacokinetics of fexofenadine, a substrate of P-glycoprotein. *Br J Clin Pharmacol* **61**:538–44.

Soukup ST, Helppi J, Müller DR, Zierau O, Watzl B, Vollmer G, Diel P, Bub A, and Kulling SE (2016) Phase II metabolism of the soy isoflavones genistein and daidzein in humans, rats and mice: a cross-species and sex comparison. *Arch Toxicol* **90**:1335–1347.

Tada I, Chaleckis R, Tsugawa H, Meister I, Zhang P, Lazarinis N, Dahlén B, Wheelock CE, and Arita M (2020) Correlation-Based Deconvolution (CorrDec) To Generate High-Quality MS2 Spectra from Data-Independent Acquisition in Multisample Studies. *Anal Chem* **92**:11310–11317.

Tsugawa H, Nakabayashi R, Mori T, Yamada Y, Takahashi M, Rai A, Sugiyama R, Yamamoto H, Nakaya T, Yamazaki M, Kooke R, Bac-Molenaar JA, Oztolan-Erol N, Keurentjes JJB, Arita M, and Saito K (2019) A cheminformatics approach to characterize metabolomes in stable-isotope-labeled organisms. *Nat Methods* **16**:295–298.

Tsuruya Y, Kato K, Sano Y, Imamura Y, Maeda K, Kumagai Y, Sugiyama Y, and Kusuhara H (2016) Investigation of Endogenous Compounds Applicable to Drug-Drug Interaction Studies Involving the Renal Organic Anion Transporters, OAT1 and OAT3, in Humans. *Drug Metab Dispos* **44**:1925–1933.

- Uhr M, Holsboer F, and Müller MB (2002) Penetration of endogenous steroid hormones corticosterone, cortisol, aldosterone and progesterone into the brain is enhanced in mice deficient for both *mdr1a* and *mdr1b* P-glycoproteins. *J Neuroendocrinol* **14**:753–759.
- van de Wetering K, and Sapth S (2012) ABCG2 functions as a general phytoestrogen sulfate transporter in vivo. *FASEB J* **26**:4014–24.
- van Herwaarden AE, Wagenaar E, Merino G, Jonker JW, Rosing H, Beijnen JH, and Schinkel AH (2007) Multidrug Transporter ABCG2/Breast Cancer Resistance Protein Secretes Riboflavin (Vitamin B2) into Milk. *Mol Cell Biol* **27**:1247–1253.
- Vlaming MLH, Lagas JS, and Schinkel AH (2009) Physiological and pharmacological roles of ABCG2 (BCRP): recent findings in *Abcg2* knockout mice. *Adv Drug Deliv Rev* **61**:14–25.
- Watanabe S, Yamaguchi M, Sobue T, Takahashi T, Miura T, Arai Y, Mazur W, Wähälä K, and Adlercreutz H (1998) Pharmacokinetics of soybean isoflavones in plasma, urine and feces of men after ingestion of 60 g baked soybean powder (kinako). *J Nutr* **128**:1710–1715.
- Wessler JD, Grip LT, Mendell J, and Giugliano RP (2013) The P-glycoprotein transport system and cardiovascular drugs. *J Am Coll Cardiol* **61**:2495–2502.
- Yang Z, Zhu W, Gao S, Yin T, Jiang W, and Hu M (2012) Breast cancer resistance protein (ABCG2) determines distribution of genistein phase II metabolites: Reevaluation of the roles of ABCG2 in the disposition of genistein. *Drug Metab Dispos* **40**:1883–1893.
- Zamek-Gliszczyński MJ, Goldstein KM, Paulman A, Baker TK, and Ryan TP (2013) Minor compensatory changes in SAGE *Mdr1a* (P-gp), *Bcrp*, and *Mrp2* knockout rats do not detract from their utility in the study of transporter-mediated pharmacokinetics. *Drug Metab Dispos* **41**:1174–8.