

## DAFTAR PUSTAKA

- Adachi. K., Toriyama. K., Azekura. T., Morioka. K., Tongnunui. P., Ikejima. K., 2012. Potent Cellulase Activity in the Hepatopancreas of Mangrove Crabs. *Fish Sci* (2012) 78:1309–1314
- Aiyer, P.V.D. 2004. 'Effect of C:N Ratio on Alpha Amylase Production by *Bacillus licheniformis* SPT 27', *African J. of Biotech.* Vol. 3 (10), pp. 519-522, October 2004.
- Allardyce, B.J., 2010. *Cellulose and Hemicellulose Digestion by Herbivorous Terrestrial Crustaceans*. Disertasi (Online). Deakin University
- Anonim. Cellulase. [http://en.wikipedia.org/wiki/files.Types\\_of\\_Cellulase.org](http://en.wikipedia.org/wiki/files.Types_of_Cellulase.org). Diakses: 18 November 2012
- Asosiasi Pulp dan Kertas Indonesia. 2005. *Pedoman Pengelolaan Limbah Padat Industri Pulp dan Kertas*. Temu usaha dan pengawasan standar operasional prosedur (SOP) pengelolaan limbah B3 pada industri pulp dan kertas. Semarang.
- Bakare M.K., Adewale I.O., Ajayi. A., Shonukan O.O., 2005. Purification and characterization of cellulase from the wild-type and two improved mutants of *Pseudomonas fluorescens*. *African J. of Biotech* Vol. 4 (9), pp. 898-904.
- Bassett. J., Denney. R.C., Jeffery. G.H., Mendham. J., 1964. *Buku Ajar Vogel Kimia Analisis Kuantitatif Anorganik*. Penerbit Buku Kedokteran EGC. Jakarta.
- Campbell, M. K., Farrell, S.O. 2009. *Biochemistry*, Sixth Edition. Thomson Higher Education. Belmont, USA.
- Dahlan, Hatta. M. 2011. *Pengolahan Limbah Kertas menjadi Pulp sebagai Bahan Pengemas Produk Agroindustri*. Prosiding Seminar Nasional AVoER ke-3 Palembang. ISBN : 979-587-395-4
- El-Safey, E.M. dan Ammar, M.S. (2004). 'Purification and Characterization of  $\alpha$ -amylase Isolated from *Aspergillus falvus var.columnaris*', *Ass. Univ.Bull. Environ. Res.* Vol. 7 No. 1, March 2004.

- Erlina, Lili. 1998. *Isolasi, Karakterisasi dan Pemanfaatan Enzim Selulase dari Bekicot (Achatina fulica)*. Skripsi jurusan kimia FMIPA Universitas Diponegoro. Semarang.
- Gokhan, C., Burhan, A., M. Nisa, U., Hatice, G. 2002. Some Properties of Crude Carboxymethyl Cellulase of *Aspergillus niger* Z10 Wild-Type Strain. *Turk J Biol* 26 (2002) 209-213.
- Guo, Rui., Ming, D., Zhang, L.S., Xu, G., Zhao Fu-kun. 2008. Molecular Cloning and Characterization of Two Novel Cellulase Genes from the Mollusc *Ampullaria crossean*. *J Comp Physiol B* 178:209–215
- Ikram-ul-haq, Javed, M.M, Khan, S.T and Siddiq, Z. 2005. Cotton Saccharifying Activity of Cellulases Produced by Co-culture of *Aspergillus niger* and *Trichoderma viride*. *Res. J. Agric & Biol. Sci.* 1(3):241-245.
- Jagtap S, Rao M., 2005. Purification and Properties of a Low Molecular Weight  $\beta$ -1,4-Glucan Glucohydrolase Having One Active Site for Carboxymethyl Cellulose and Xylan from an Alkalothermophilic *Thermomonospora* sp. *Biochem Biophys Res Commun* 329(1):111–116.
- Jasman. 2002. *Aktivitas Selulase Rayap Terhadap Substrat Kertas dan Serbuk Gergaji*. Tesis. Universitas Hasanuddin. Makassar.
- Joana, L.A.B. *et al.* 2011. Structural Insights Into a Unique Cellulase Fold and Mechanism of Cellulose Hydrolysis. *Proc Natl Acad Sci USA* vol. 108 no. 20 5237-5242
- Kikuchi. T., Shibuya. H., Jones. T.J., 2005. Molecular and Biochemical Characterization of an Endo-b-1, 3-glucanase from The Pinewood Nematode *Bursaphelenchus xylophilus* Acquired by Horizontal Gene Transfer from Bacteria. *Biochem J* 389:117–125
- Kumar, G.S., Chandra, M.S., Sumanth. M., Vishnupriya. A., Reddy, B.R., Choi, Y.L. 2009. Cellulolytic Enzymes Production from Submerged Fermentation of Different Substrates by Newly Isolated Bacillus spp. *FME. J. Korean Soc. Appl. Biol. Chem.* 52(1), 17-21.
- Lembaga Kajian Ekologi dan Konservasi Lahan Basah. 2002. *Minimasi Limbah dalam Industri Pulp dan Paper*. <http://www.ecoton.or.id>. Diakses 28 Januari 2013.

- Lin, S. and Stutzenberger, F. J., 1995, Purification and Characterization of the Major Beta-1,4-endoglucanase from *Thermomonospora curvata*. *J. Appl. Bacteriol.* 79(4), 447-453.
- Linton, S.M., dan Greenaway, P., 2004. Presence and Properties of Cellulase and Hemicellulase Enzymes of the Gecarcinid Land Crabs *Gecarcoidea natalis* and *Discoplax hirtipes*. *The Journal of Experim. Bio* 207, 4095-4104
- Maisaroh. 2009. *Hidrolisis Selulosa Bagas dengan Enzim Selulase dari Bekicot Achatina fulica untuk Produksi Etanol dengan Zymonas mobilis* A3. Tesis Jurusan Kimia FMIPA ITS Surabaya.
- Moriya S, Tanaka K, Ohkuma M, Sugano S, Kudo T. 2001. Diversification of the Microtubule System in the Early Stage of Eukaryote Evolution: Elongation Factor 1 Alpha and Alpha-tubulin Protein Phylogeny of Termite Symbiotic Oxymonad and Hypermastigote Protists. *J Mol Evol.* 2001 Jan;52(1):6-16.
- Murray, R.K., Granner, D.K., and Victor, R.W. 2009. *Biokimia Harper*. EGC Penerbit Buku Kedokteran Jakarta. 68-69.
- Natsir, H. 2010. *Kajian Enzim Kitinase Termostabil dari Bakteri Termofil: Produksi, Pemurnian, Karakterisasi, dan Aplikasi dalam Hidrolisis Kitin*. Disertasi. Universitas Hasanuddin. Makassar.
- Nawaz, S., Malana, M. A., Ikram, N., Hafeez, S. 2006. Kinetic Study of Carboxymethyl Cellulase From *Trichoderma harzianum*. *Pak. J. Life Soc. Sci.* (2006), 4(1-2): 15-19
- Nelson, D.L. dan Cox, M.M. 2004. *Lehninger Principles of Biochemistry Fourth Edition*. University of Wisconsin. Madison.
- Niranjane, ajay. 2004. *Screening Diverse Cellulase Enzymes from The White Rot Fungus Phlebia Gigantea for High Activity and Large Scale Applications*. Thesis Department of Biotechnology and Environmental Biology Royal Melbourne Institute of Technology (RMIT University).
- Novalina, Y. 2002. *Pemanfaatan kertas koran bekas sebagai bahan baku dalam pembuatan kertas karton*. Skripsi. Jurusan Teknologi Hasil Hutan. Fakultas Kehutanan. Institut Pertanian Bogor. Bogor.
- Page, D.S. 1989. *Prinsip-prinsip Biokimia edisi kedua*. Alih Bahasa: R. Soendoro. Erlangga. Jakarta.

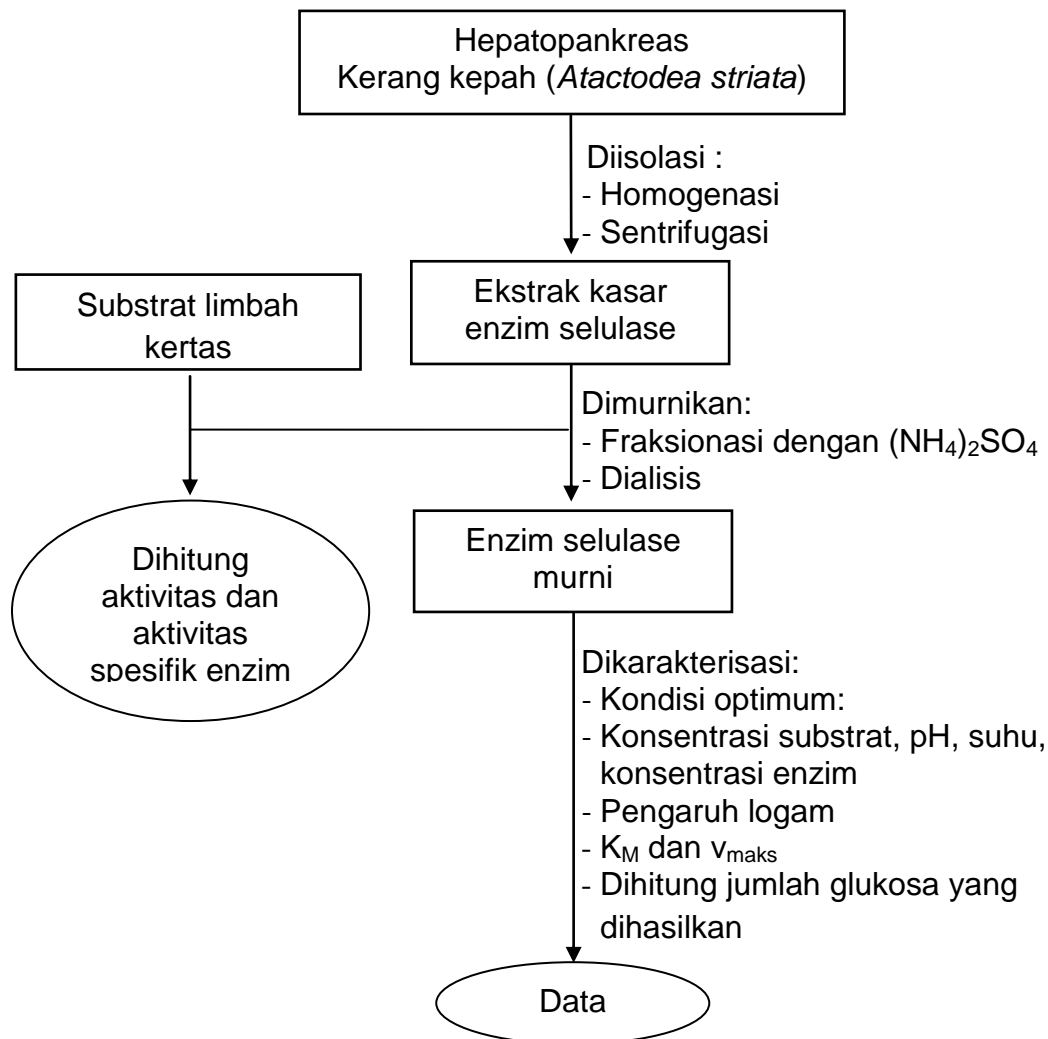
- Palmer, T. 1991. *Understanding enzymes*. Ellis harwood. Chichester, west sussex. England
- Permasih, Dian. 2011. *Metabolisme Kapsel* (Online). <http://www.scribd.com/doc/54988027/4/Teori-Gembok-dan-Kunci-Lock-and-Key-Theory>. Diakses: 19 November 2012
- Purbasari, Dian. 2008. *Produksi dan Karakterisasi Hidrolisat Protein dari Kerang Mas Ngur (Atractodes striata)*. Skripsi (Online): Program Studi Teknologi Hasil Perikanan Fakultas Perikanan Dan Ilmu Kelautan. Institut Pertanian Bogor
- Rajagukguk, S.K. 1997. *Pengaruh Umur Kertas dan Penggunaan Bahan Kolektor terhadap Penghilangan Tinta Kertas Koran Bekas*. Skripsi (Online). Fakultas Teknologi Pertanian, Institut Pertanian Bogor.
- Riaz, Tariq & Javaid, arshad. 2010. Fungi Associated With Paper Processing Materials. *Pak. J. Phytopathol. Vol. 22(1):06-08,2010*.
- Sadikin, M. 2002. *Biokimia Enzim*. Widya Medika. Jakarta.
- Sakamoto, K dan Toyohara, H. 2009. A Comparative Study of Cellulase and Hemicellulase Activities of Brackish Water Clam *Corbicula japonica* with Those of Other Marine Veneroida Bivalves. *J. Experim Biol* 212, 2812-2818.
- Santoso, U. 1987. *Limbah Bahan Ransum Unggas yang Rasional*. PT. Bhratara Karya Aksara.
- Sona. A., 2004. *Studies on Isolation, Purification and Properties of Endoglucanase from the Hepatopancreas of Perna viridi*. Thesis (Online), Central Institute of Fisheries Technology, Cochin.
- Stryer, L. 1995. *Biochemistry*. 4 th ed., W.H. Freeman and Company, New York.
- Stryer, L., Tymoczko, J.L., Berg, J.M., 2002. *Biochemistry* 5 th ed., W.H. Freeman and Company, New York.
- Sudarmadji, S., Haryono, B., Suhardi. 2003. *Analisa Bahan Makanan dan Pertanian*. Liberty. Yogyakarta.
- Sugimura. M., Watanabe. H., Lo. N., Saito. H., 2003. Purification, Characterization, cDNA Cloning and Nucleotide Sequencing of a Cellulase from the Yellow-Spotted Longicorn Beetle, *Psacotha hilaris*. *Eur J Biochem* 270:3455–3460.

- Suhartono, M.T. 1989. *Enzim dan Bioteknologi*. Direktorat Jenderal Pendidikan Tinggi Antar Universitas Bioteknologi, Institut Pertanian Bogor.
- Sunarto 2001. *Remis, Kerang Suku Mesodesmatidae, Penghuni Pasir Pantai Pulau-Pulau Karang*. Warta Puslitbang Oseanologi, Volume Xv Nomor 1. Jakarta.P.8-11.
- Suzuki. K., Ojima. T., Nishita. K., 2003. Purification and cDNA Cloning of a Cellulase from Abalone *Haliotis discus hannai*. *Eur. J. Biochem.* 270, 771–778.
- Syafii W, Siregar IZ. 2006. Chemical Properties and Fiber Dimension of *Acacia mangium* Willd. From Three Provenances. *J. Tropical Wood Science and Technology* 4:28-32.
- Underwood, A.L., Day, R.A.,2002. *Analisis Kimia Kuantitatif*. Erlangga. Jakarta.
- Vocadlo D. J., Davies G. J., Laine R., Withers S. G. 2001. "Catalysis by Hen Egg-White Lysozyme Proceeds via a Covalent Intermediate". *Nature* 412 (6849): 835–8
- Waranmaselembun, Celsius. 2007. *Komposisi Kimia dan Aktivitas Inhibitor Topoisomerase i dari Kerang mas ngur (Atractodea striata)*. Tesis Institut Pertanian Bogor. Bogor.
- Watanabe, H., Tokuda, G. 2001. Review: Animal Cellulases. *CMLS, Cell. Mol. Life Sci.* 58 (2001) 1167–1178
- Winarno. F.G. 2004. *Kimia Pangan dan Gizi*. PT Gramedia Pustaka Utama. Jakarta
- Xu, Bingze. 2002. *Endoglucanase and Mannanase from Blue Mussel, Mytilus edulis Purification, Characterization, Gene and Three Dimensional Structure*. Disertasi (Online). Uppsala University, Uppsala, Sweden.
- Xu, B., Sellos, D. and Janson, J. (2002). Cloning and Expression in *Pichia pastoris* of a Blue Mussel (*Mytilus edulis*)  $\beta$ -mannanase Gene. *Eur. J. Biochem.* 269, 1753-1760.

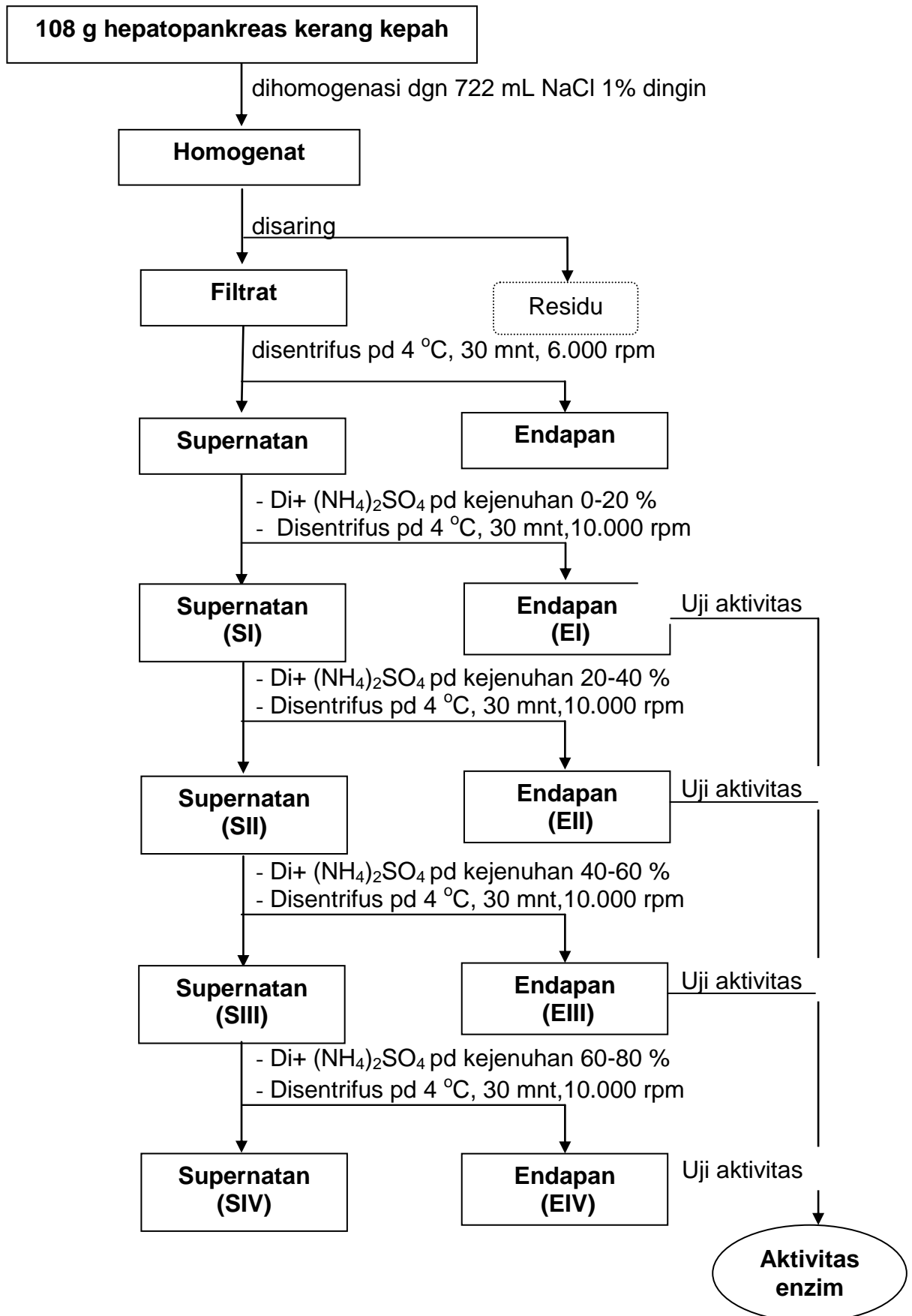
## LAMPIRAN

### Lampiran 1.

#### Gambaran Umum Prosedur Penelitian

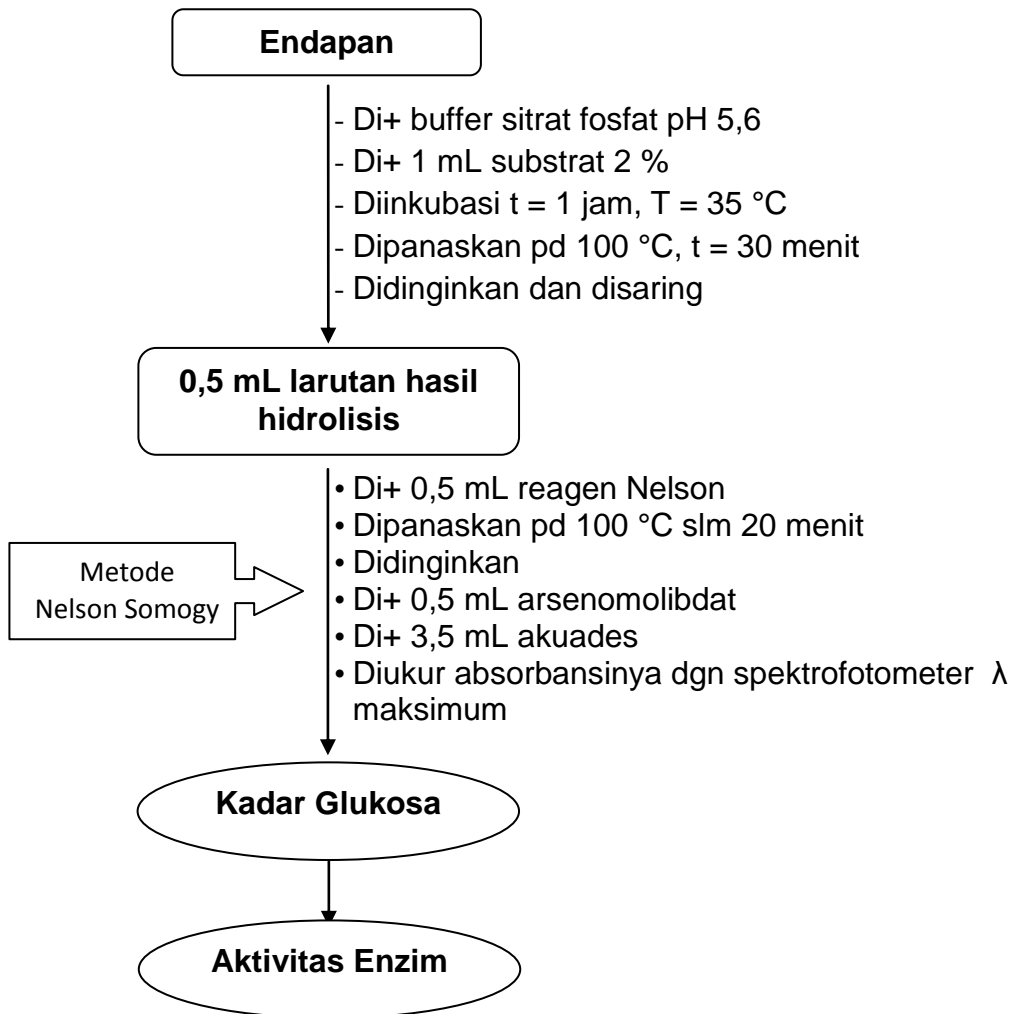


**Lampiran 2. Isolasi enzim selulase dari kerang kepah  
(*Atactodea striata*)**

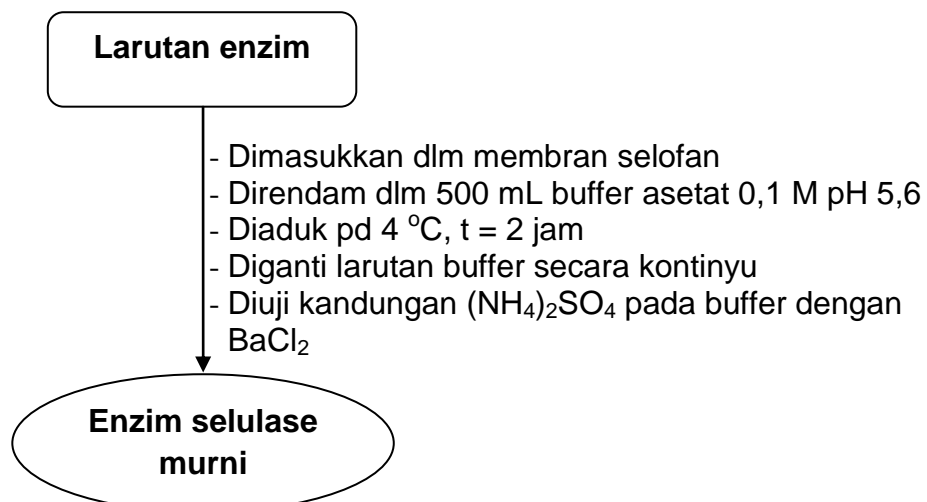


### Lampiran 3. Uji Aktivitas Enzim, Dialisis, dan Penentuan Kadar Protein

#### I. Uji Aktivitas

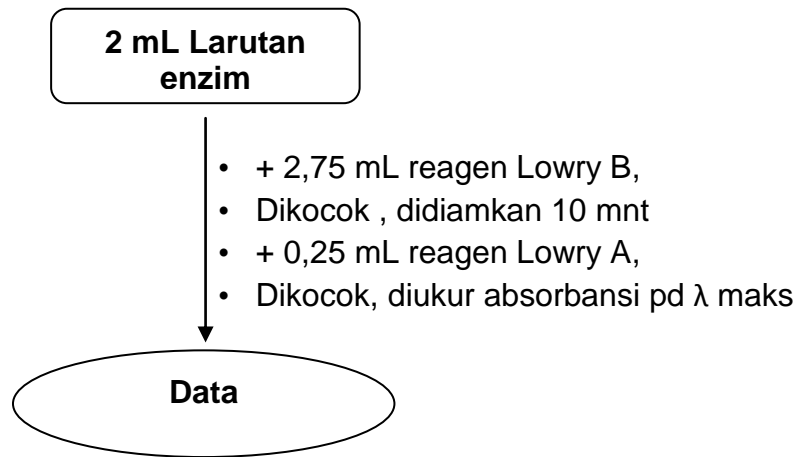


#### II. Dialisis



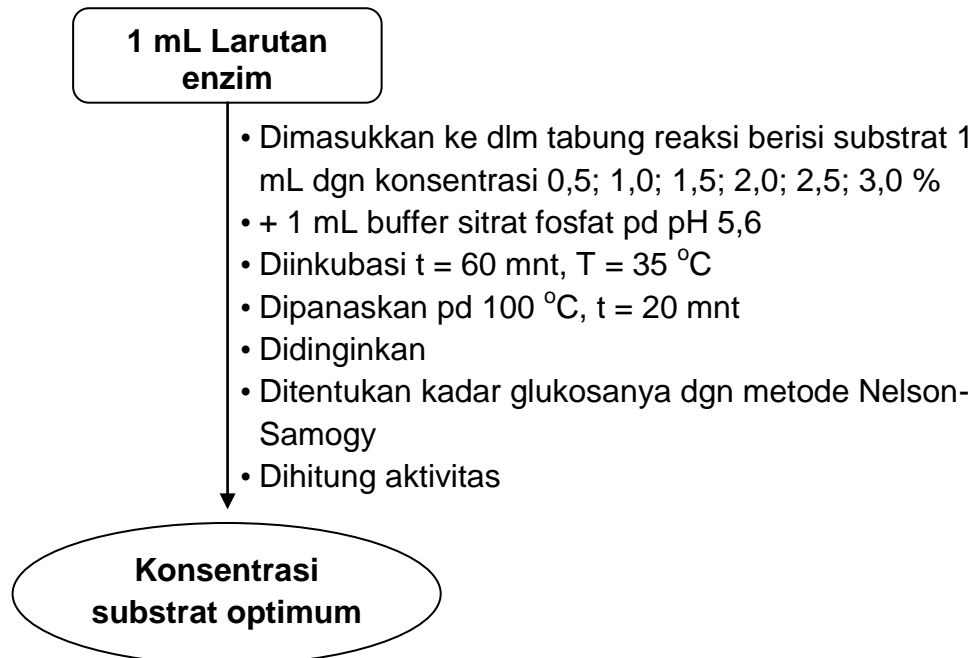


### III. Penentuan Kadar Protein

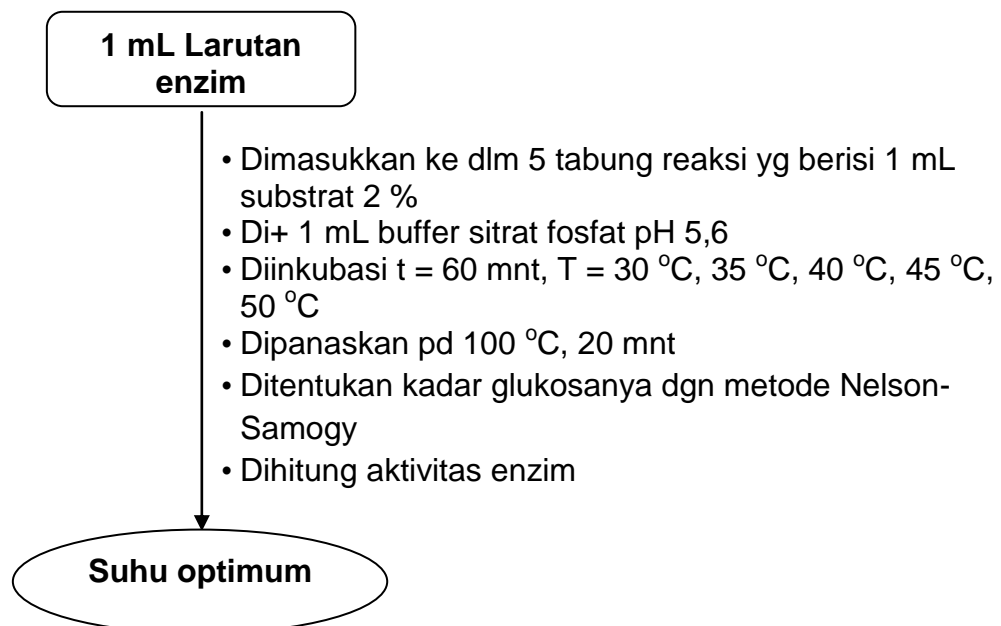


## Lampiran 4. Karakterisasi Enzim Selulase

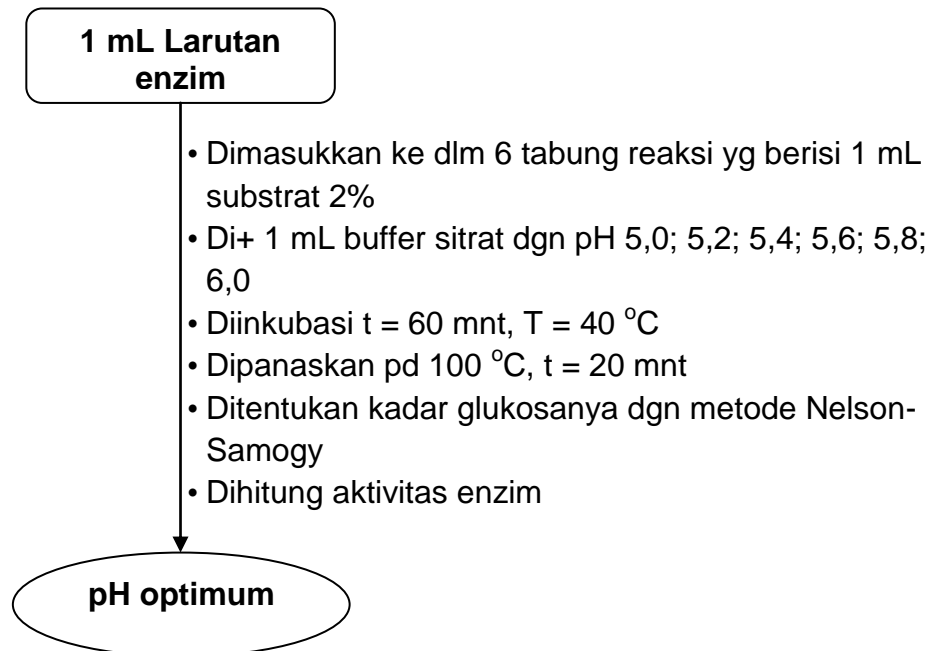
### a. Penentuan konsentrasi substrat optimum



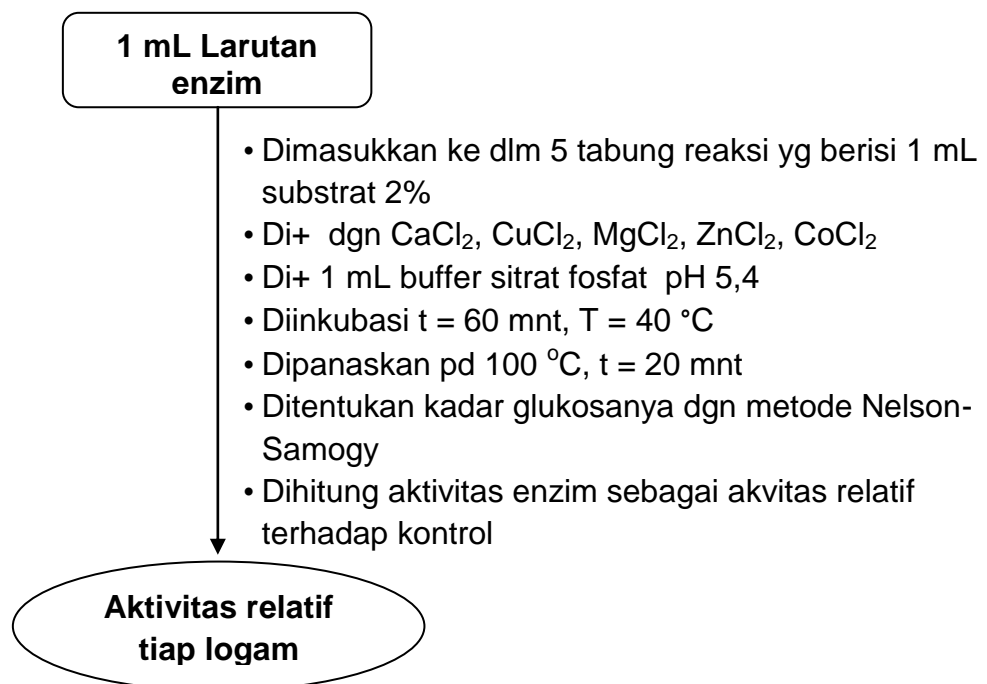
### b. Penentuan suhu optimum inkubasi



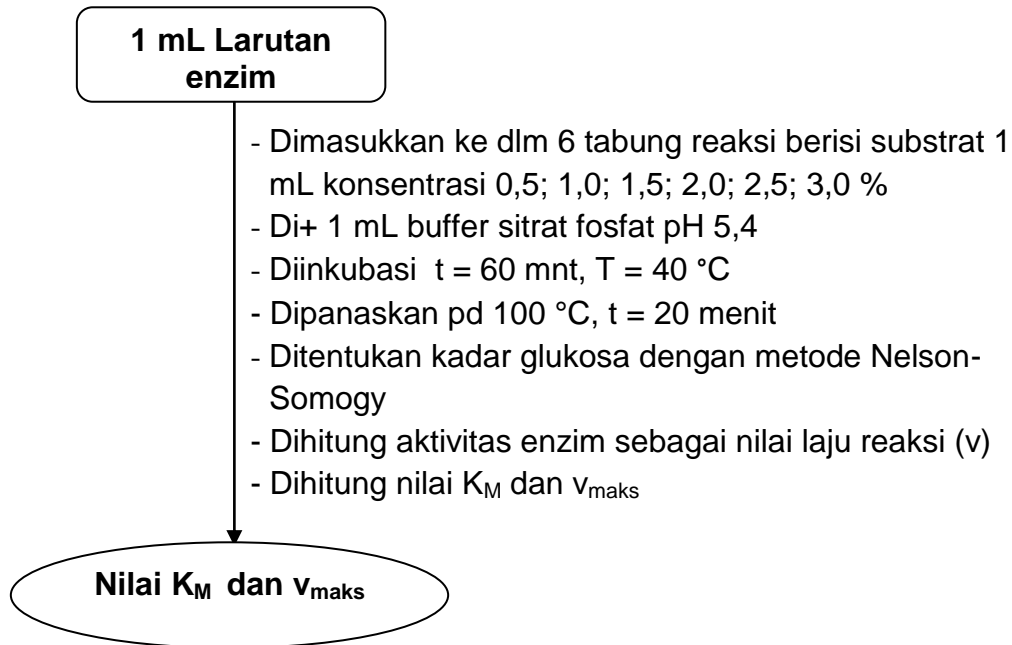
### c. Penentuan pH optimum



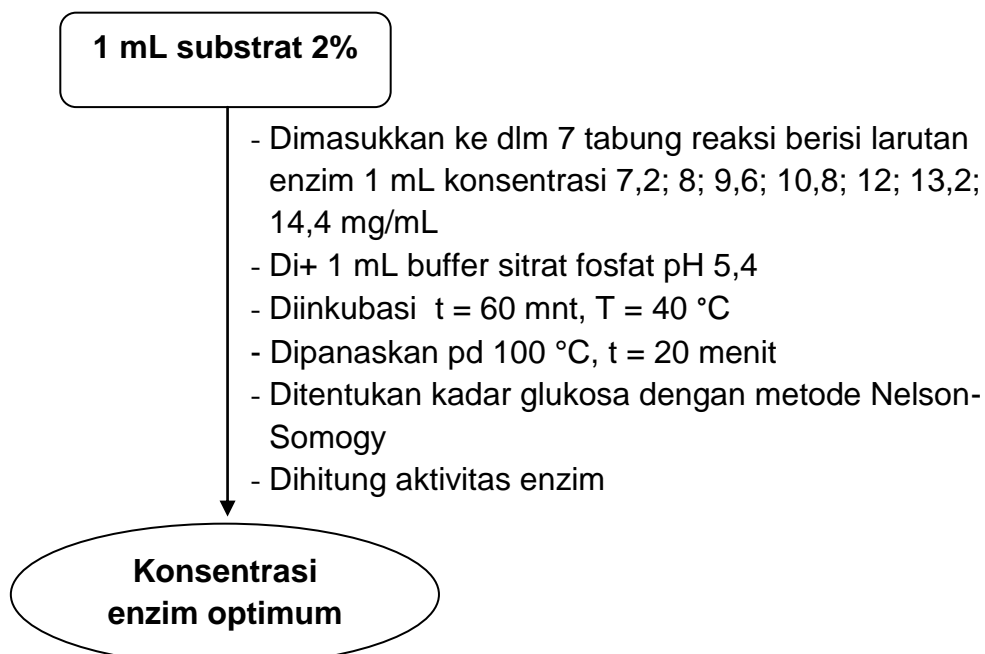
### d. Penentuan Pengaruh Aktivator dan Inhibitor



### e. Penentuan parameter kinetik nilai $K_M$ dan $v_{maks}$

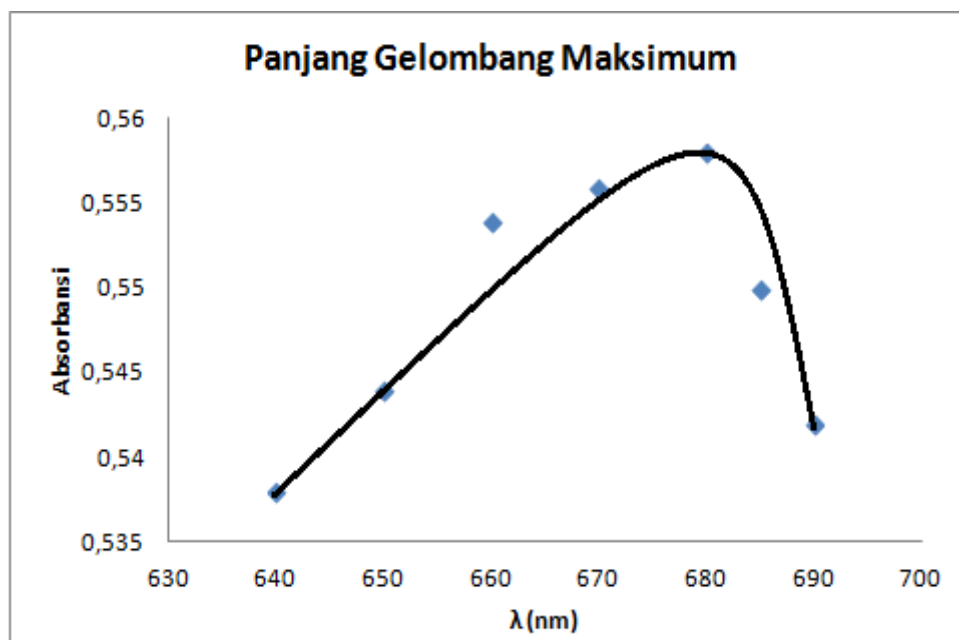


### f. Penentuan konsentrasi enzim optimum



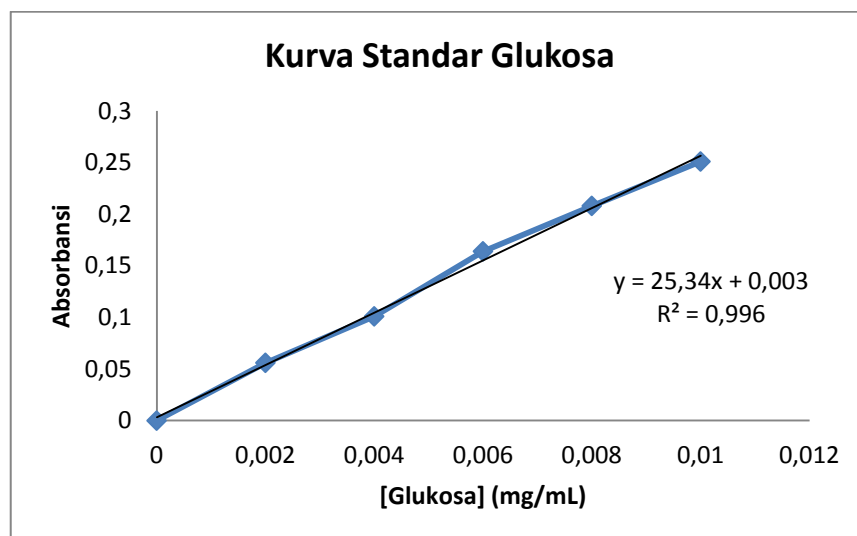
**Lampiran 5. Hasil pengukuran panjang gelombang maksimum untuk penentuan kurva standar dan kadar glukosa**

$\lambda$ (nm)	Absorbansi
640	0,538
650	0,544
660	0,554
670	0,556
680	0,558
685	0,550
690	0,542



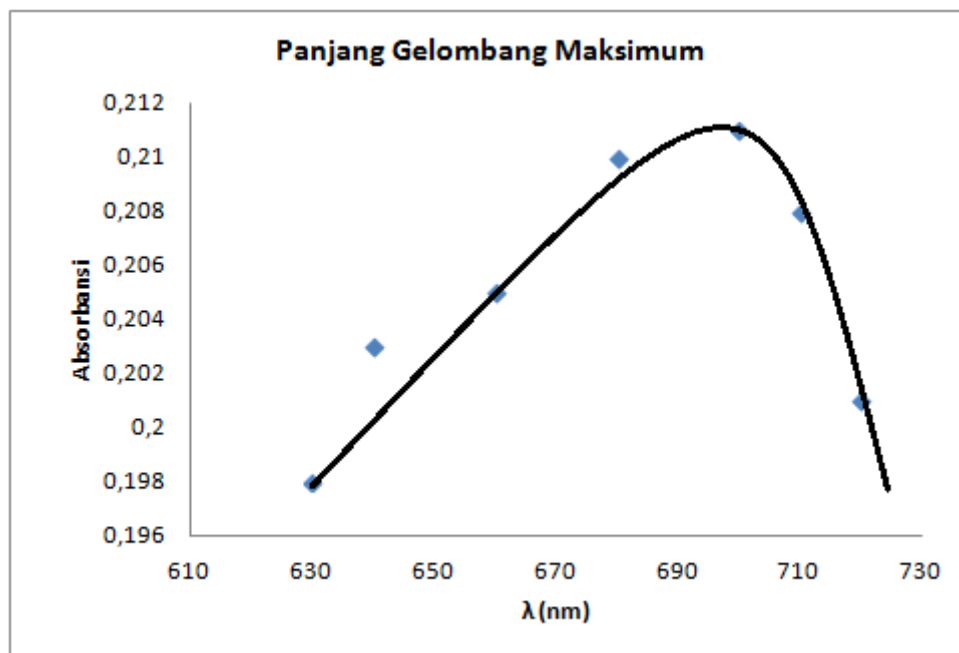
### Lampiran 6. Hasil pengukuran kurva standar glukosa

[Glukosa] (mg/mL)	Absorbansi
0	0
0,002	0,056
0,004	0,101
0,006	0,164
0,008	0,208
0,010	0,251



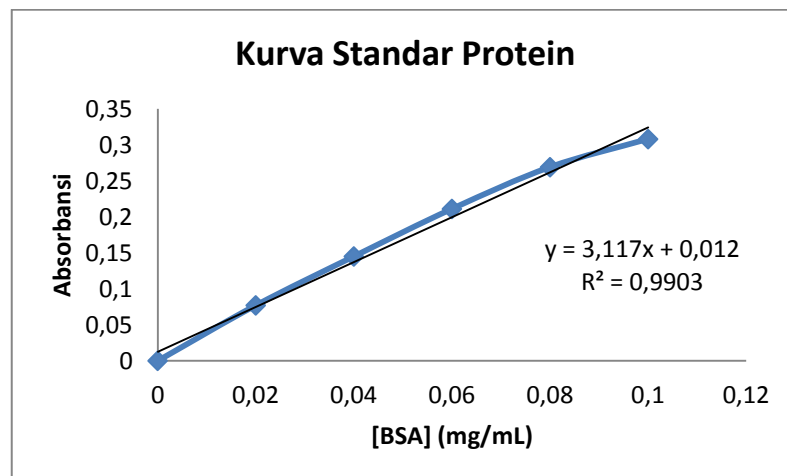
**Lampiran 7. Hasil pengukuran panjang gelombang maksimum untuk penentuan kurva standar dan kadar protein**

$\lambda$ (nm)	Absorbansi
630	0,198
640	0,203
660	0,205
680	0,210
700	0,211
710	0,208
720	0,201



**Lampiran 8. Hasil pengukuran kurva standar protein**

[BSA] (mg/mL)	Absorbansi
0	0
0,02	0,077
0,04	0,145
0,06	0,211
0,08	0,269
0,10	0,308





**Lampiran 9. Pembuatan larutan buffer sitrat fosfat**

20 mL campuran dari X mL  $\text{Na}_2\text{HPO}_4$  0,2 M dan Y mL asam sitrat 0,1 M

<b>X (mL) <math>\text{Na}_2\text{HPO}_4</math></b>	<b>Y (mL) Asam sitrat</b>	<b>pH</b>
7,71	12,29	4,0
8,28	11,72	4,2
8,82	11,18	4,4
9,35	10,65	4,6
9,86	10,14	4,8
10,30	9,70	5,0
10,72	9,28	5,2
11,15	8,85	5,4
11,60	8,40	5,6
12,09	7,91	5,8
12,63	7,37	6,0
13,22	6,78	6,2
13,85	6,15	6,4

**Lampiran 10. Contoh perhitungan penentuan aktivitas enzim**

Fraksi 70% → Absorbansi = 0,303

Persamaan garis lurus:  $y = 25,34x + 0,003$

$$\text{Kadar glukosa} = \frac{0,303 - 0,003}{25,34}$$

$$= 0,0118$$

$$\text{Aktivitas enzim} = \frac{\text{Kadar glukosa} \times 1000}{\text{BM} \times t}$$

$$= \frac{0,0118 \times 1000}{180 \times 60}$$

$$= 1,09 \times 10^{-3} \text{ U/mL}$$

### Lampiran 11. Contoh perhitungan penentuan kadar protein dan aktivitas spesifik

#### a. Penentuan kadar protein

Fraksi 70% → Absorbansi = 0,736 dengan fp = 100

Persamaan garis lurus:  $y = 3,117x + 0,012$

$$\text{Kadar Protein} = \frac{0,736 - 0,012}{3,117}$$

$$= 0,23227$$

$$\text{Kadar Protein} = 0,23227 \times \text{fp}$$

$$= 0,23227 \times 100$$

$$= 23,227 \text{ mg/mL}$$

#### b. Penentuan aktivitas spesifik

$$\text{Aktivitas spesifik} = \frac{\text{Aktivitas enzim (U/mL)}}{\text{Kadar protein (mg/mL)}}$$

$$= \frac{0,00109 \text{ U/mL}}{23,227 \text{ mg/mL}}$$

$$= 4,719 \times 10^{-5} \text{ U/mg}$$

**Lampiran 12. Data aktivitas enzim setiap fraksi**

<b>Fraksi</b>	<b>Absorbansi [glukosa]</b>	<b>[Glukosa] (mg/mL)</b>	<b>Aktivitas Enzim (U/mL)</b>
0-20 %	0,120	$4,60 \times 10^{-3}$	$4,30 \times 10^{-4}$
0-30 %	0,115	$4,40 \times 10^{-3}$	$4,10 \times 10^{-4}$
20-40 %	0,170	$6,60 \times 10^{-3}$	$6,10 \times 10^{-4}$
30-50 %	0,210	$8,20 \times 10^{-3}$	$7,60 \times 10^{-4}$
40-60 %	0,266	$1,04 \times 10^{-2}$	$9,60 \times 10^{-4}$
50-70 %	0,303	$1,18 \times 10^{-2}$	$1,10 \times 10^{-3}$
60-80 %	0,104	$3,90 \times 10^{-3}$	$3,70 \times 10^{-4}$

**Lampiran 13. Data Penentuan Konsentrasi Substrat Optimum**

<b>[substrat] (%)</b>	<b>Absorbansi</b>	<b>Kadar Glukosa (mg/mL)</b>	<b>Aktivitas Enzim (U/mL)</b>
0,5	0,091	$3,47 \times 10^{-3}$	$3,22 \times 10^{-4}$
1	0,159	$6,16 \times 10^{-3}$	$5,70 \times 10^{-4}$
1,5	0,173	$6,71 \times 10^{-3}$	$6,21 \times 10^{-4}$
2	0,205	$7,97 \times 10^{-3}$	$7,38 \times 10^{-4}$
2,5	0,203	$7,89 \times 10^{-3}$	$7,31 \times 10^{-4}$
3	0,200	$7,77 \times 10^{-4}$	$7,20 \times 10^{-4}$

**Lampiran 14. Data Penentuan pH Optimum**

<b>pH</b>	<b>Absorbansi</b>	<b>Kadar Glukosa (mg/mL)</b>	<b>Aktivitas Enzim (U/mL)</b>
5	0,032	$1,14 \times 10^{-3}$	$1,06 \times 10^{-4}$
5,2	0,102	$3,91 \times 10^{-3}$	$3,62 \times 10^{-4}$
5,4	0,118	$4,54 \times 10^{-3}$	$4,20 \times 10^{-4}$
5,6	0,076	$2,88 \times 10^{-3}$	$2,67 \times 10^{-4}$
5,8	0,063	$2,37 \times 10^{-3}$	$2,19 \times 10^{-4}$
6	0,034	$1,22 \times 10^{-3}$	$1,13 \times 10^{-4}$

**Lampiran 15. Data Penentuan Suhu Optimum**

<b>Suhu (°C)</b>	<b>Absorbansi</b>	<b>Kadar Glukosa (mg/mL)</b>	<b>Aktivitas Enzim (U/mL)</b>
30	0,039	$1,42 \times 10^{-3}$	$1,32 \times 10^{-4}$
35	0,105	$4,03 \times 10^{-3}$	$3,73 \times 10^{-4}$
40	0,110	$4,22 \times 10^{-3}$	$3,91 \times 10^{-4}$
45	0,071	$2,68 \times 10^{-3}$	$2,48 \times 10^{-4}$
50	0,046	$1,69 \times 10^{-3}$	$1,57 \times 10^{-4}$

**Lampiran 16. Data penentuan pengaruh konsentrasi enzim terhadap aktivitas enzim**

<b>[Enzim] (mg/mL)</b>	<b>Absorbansi</b>	<b>Kadar Glukosa (mg/mL)</b>	<b>Aktivitas enzim (U/mL)</b>
7,2	0,089	$3,39 \times 10^{-3}$	$3,14 \times 10^{-4}$
8	0,196	$7,62 \times 10^{-3}$	$7,05 \times 10^{-4}$
9,6	0,309	$1,21 \times 10^{-2}$	$1,12 \times 10^{-3}$
10,8	0,418	$1,64 \times 10^{-2}$	$1,52 \times 10^{-3}$
12	0,52	$2,04 \times 10^{-2}$	$1,89 \times 10^{-3}$
13,2	0,515	$2,02 \times 10^{-2}$	$1,88 \times 10^{-3}$
14,4	0,5	$1,96 \times 10^{-2}$	$1,82 \times 10^{-3}$



**Lampiran 17. Data penentuan pengaruh logam terhadap aktivitas enzim**

<b>Logam</b>	<b>Absorbansi</b>	<b>Kadar Glukosa (mg/mL)</b>	<b>Aktivitas Enzim (U/mL)</b>	<b>Aktivitas Relatif (%)</b>
Kontrol	0,183	$7,10 \times 10^{-3}$	$6,58 \times 10^{-4}$	100
Mg	0,155	$5,99 \times 10^{-3}$	$5,55 \times 10^{-4}$	84,44
Co	0,198	$7,69 \times 10^{-3}$	$7,13 \times 10^{-4}$	108,33
Ca	0,138	$5,33 \times 10^{-3}$	$4,93 \times 10^{-4}$	75
Cu	0,086	$3,28 \times 10^{-3}$	$3,03 \times 10^{-4}$	46,11
Zn	0,092	$3,51 \times 10^{-3}$	$3,25 \times 10^{-4}$	49,44

### Lampiran 18. Data Aktivitas Spesifik Enzim Selulase pada Kondisi Optimum

Kondisi Optimum:[substrat]=2 %; suhu=40 °C; pH=5,4; [enzim]= 12 mg/mL

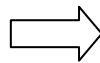
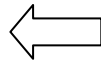
<b>[Glukosa] (mg/mL)</b>	<b>Aktifitas Enzim (U/mL)</b>	<b>Kadar Protein (mg/mL)</b>	<b>Faktor Pengenceran (fp)</b>	<b>Kadar Protein x fp</b>	<b>Aktivitas Spesifik</b>
0,0204	$1,89 \times 10^{-3}$	0,23	50	11,65	$1,62 \times 10^{-4}$

## Lampiran 20. Foto Penelitian

### Preparasi kertas koran sebagai substrat



## Proses isolasi enzim selulase dari kerang kepah

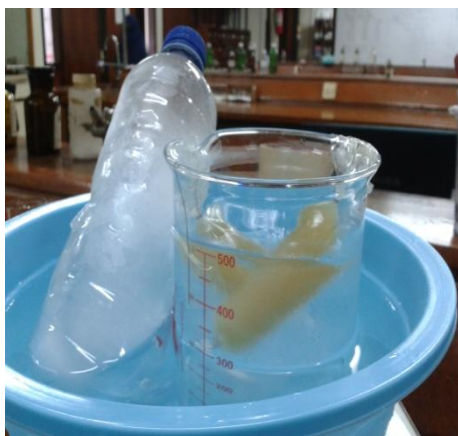




## Fraksinasi dengan amonium sulfat



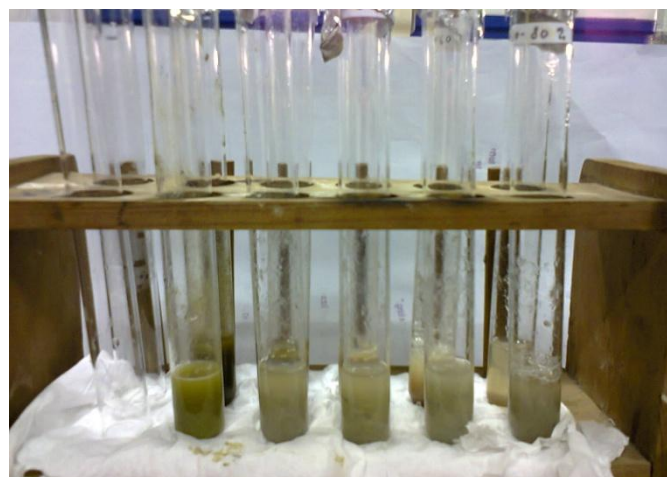
## Proses Dialisis



## Penentuan Aktivitas Enzim



Larutan standar

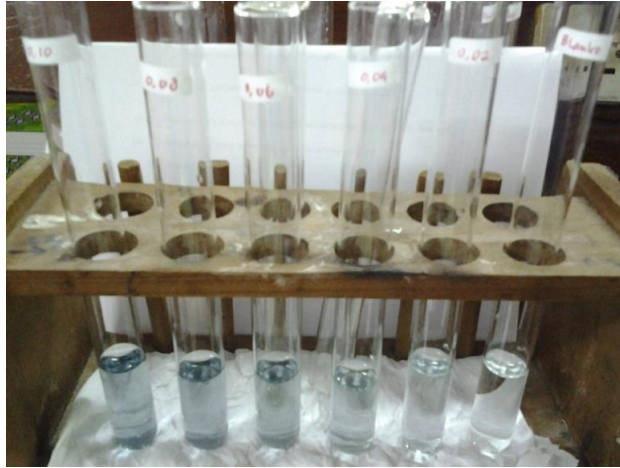


Proses hidrolisis selulosa (enzim selulase + substrat kertas + buffer)



Penentuan kadar glukosa (hasil hidrolisis + reagen Nelson + arsenomolibdar + H<sub>2</sub>O)

## Penentuan kadar protein



Larutan standar



Sampel



Pengukuran absorbansi dengan spektronik 20 D<sup>+</sup>

